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PROBLEMS

of Infectious and Parasitic Diseases

**NATIONAL CENTER OF INFECTIOUS AND PARASITIC DISEASES
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PROBLEMS OF INFECTIOUS AND PARASITIC DISEASES
VOLUME 30, NUMBER 2/2002

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EFFECT OF A FUNGIAL Cu/Zn SUPEROXIDE DISMUTASE ON THE FUNCTIONS OF PHAGOCYtic CELLS IN *YERSINIA ENTEROCOLITICA* PYV (+) INFECTED MICE

E. Ivanova¹, D. Donkova¹, Z. Stefanova¹,
R. Toshkova², M. Angelova¹, W. Voelter³,
H. Najdenski¹

1. Institute of Microbiology, Bulgarian Academy of Sciences, Sofia, Bulgaria
2. Institute of Experimental Pathology and Parasitology, Bulgarian Academy of Sciences, Sofia, Bulgaria
3. University of Tübingen, Tübingen, Germany

SUMMARY

Effect of Cu/Zn superoxide dismutase (SOD) isolated from the fungal strain *Humicola lutea* 103 (HLSOD) on the functions of phagocyte cells in *Yersinia enterocolitica* pYV(+) (YE) infected ICR mice and their survival rate was studied. Migration and killing activities of peritoneal macrophages, as well as phagocyte abilities of blood polymorphous-nuclear leukocytes (PMNs) and peritoneal macrophages were followed from days 3 to 21 post infection. Blood cultures and bacterial load of liver, spleen and small intestines were examined too. A marked protective effect of HLSOD on the survivability of mice after lethal YE infection was established. The mortality percent (MP) in the control group of infected animals without treatment was 100% and strongly diminished (up to 17%) after treatment by 40U HLSOD. Acceleration of bacterial clearance from blood was another effect of HLSOD treatment. Augmented values of macrophage number in healthy and infected animals were established after HLSOD application with a maximum at day 15th. Phagocyte indexes of macrophages from healthy animals increased under the action of enzyme treatment. The phagocyte number was 2 times increased in both groups of the infected animals. Macrophage killing indexes against *S. aureus* 209 were also increased in both groups of infected animals with a maximum at day 21st post infection. HLSOD restored the suppressed peroxide formation and killing ability of macrophages from infected animals against *E. coli* O₁₅₇. HLSOD produced a marked stimulation effect on the migration ability of macrophages, better expressed in healthy animals. Additionally, HLSOD induced an increase of PMNs phagocyte indexes from the infected animals and the number of the *in vitro* engulfed *S. aureus* Smith cells was 1.5-2.0 times higher in both infected groups during the whole period of observation. The established protective effect of HLSOD on survivability of YE infected mice can be explained by the participation of this enzyme in the host oxidant-antioxidant balance, resulting in a restoration of the suppressed phagocyte, killing and migration abilities of peritoneal macrophages and blood PMNs of the infected mice.

Key words: fungal superoxide dismutase, *Yersinia enterocolitica* infection, phagocyte cells

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ABBREVIATIONS USED IN THIS PAPER:

HLSOD - *Humicola lutea* superoxide dismutase, YE- *Yersinia enterocolitica*, PYV - *Yersinia* virulence plasmid, PMNs - polymorphous nuclear leukocytes, PI - phagocytic index, KI - killing index, YOPs - *Yersinia* outer membrane proteins

CORRESPONDING AUTHOR:

Assoc. Prof. E. Ivanova, MD, PhD
Institute of Microbiology, Bulgarian Academy of Sciences
26, Acad. G. Bonchev Str.
Sofia 11134, Bulgaria
E-mail: eivanova@microbio.bas.bg

INTRODUCTION

Yersinia enterocolitica (YE) is a common cause of enterocolitis, gastroenteritis, mesenteric lymphadenitis, terminal ileitis and other suppurative infections (7, 13, 16). Virulent strains of YE harbor a 70kB virulence plasmid (pYV), which bears genes that code for production of several secreted proteins, called Yops (17, 27) including the outer membrane protein Yad A (*Yersinia* adhesin) (6). Yad A and several Yops are involved in the inhibition of phagocytosis and killing of YE by human granulocytes (10, 25, 30).

Yops are synthesized at 37°C and translocated into mammalian cells upon contact (23). One of these proteins, Yop H dephosphorylates multiple tyrosine-phosphorylated proteins in eukaryotic cells (5). This protein is involved also in inhibition of the uptake of *Yersinia pseudotuberculosis* by cultured mouse macrophages (11) and by epithelial cells (22). Investigations of Visser *et al.* (30) showed that a pYV (+) strain of YP inhibited receptor-dependent or N-formyl-methionin-leucine-phenyl-alanine (f-MLP) inducible O₂⁻ production by human granulocytes, but did not affect the receptor independent (phorbol-myristat-acetate inducible) O₂⁻ production by granulocytes mediated from YE. The inhibition of f-MLP inducible O₂⁻ production by granulocytes depends on the secretion of Yops, but not on the expression of Yad A (30).

The formation of O₂⁻ radicals is catalysed by NADPH-oxidase, a membrane-bound enzymatic complex, which converts O₂ into O₂⁻ (9). NADPH-oxidase can be activated by receptor-dependent mechanisms, such as opsonized bacteria, C_{5a}, f-MLP and immune complexes, as well as by receptor-independent mechanisms, including long chain unsaturated fatty acids and phorbol-myristat-acetate (PMA). Activation of NADPH oxidase in human granulocytes is accompanied by protein tyrosine phosphorylation, suggesting that NADPH oxidase can be switched on by tyrosine kinases (3).

It was found that YE translocates virulence YOPs into a wide range of eukaryotic cells including neurons and insect cells. Yop E, H and T are cytotoxic for the adherent type cells. Invasin and/or Yad A are required for translocation of Yop into non-phagocyte cells, but not for translocation in macrophages (8). Yop E is an important virulence factor which penetrate into mammalian cells via a plasmid-encoded type III system in *Yersinia* spp. Yop E promotes the disruption of actin filaments, cell rounding and blockage of phagocytosis (4).

The effector Yops disrupt cellular defense functions such as TNF- α , O₂⁻ production and phagocytosis and allow *Yersinia* to grow (2). Yops are active proteins, which can suppress eukaryotic signal mechanism, for instance they can translocate the tyrosine phosphatase. Yop H dephosphorylates the focal adhesion proteins paxillin and 130C. The *Yersinia* effector Yop P is able to induce apoptosis in macrophages (2).

In the present work we aim to study the effect of Cu/Zn-superoxide dismutase isolated from *Humicola lutea* 103 fungal strain on the functions of phagocytic cells in mice infected by *Yersinia enterocolitica* 0:8 pYV (+).

MATERIALS AND METHODS

ANIMALS

ICR mice (130), 8 weeks old, from both sexes and weighing 18-20 g were used for the experiments. The animals were obtained from the animal house of the Institute of Microbiology, Bulgarian Academy of Sciences, Sofia. They were grown up and fed at standard conditions accepted by the Bulgarian Veterinary Health Service.

Microorganisms

Yersinia enterocolitica strain 8081, serotype O:8, pYV (+) (YE), was used for the induction of infection in mice. The strain was provided by Reference Laboratory, Institut Pasteur (Paris, France.). Bacteria were cultivated in Brain-heart infusion (BHI) (Difco) at 26°C for 18 h, then diluted 1:15 in fresh BHI supplemented by 0.02 M MgCl₂ and cultivated again for 4 h at 37°C. The bacterial cells were washed twice, suspended in saline and standardized by optical standard up to concentrations described elsewhere.

The referent strain *S. aureus* Smith was used for the phagocyte index tests of peritoneal macrophages and blood polymorphous-nuclear leukocytes (PMNs) of the experimental animals. The reference strains *S. aureus* 209 and *E. coli* O₁₂₄ K₇₂ H₃₀ were used to test microorganisms in the killing ability of peritoneal macrophages. All the 3 strains were maintained in the microbiological collection of the Department of Immunology, Bulgarian Academy of Sciences. For the present experiments the strains were cultivated on Bacto agar (Difco) (pH 7.2) at 37°C for 24 h. Bacterial cells were collected, washed and adjusted to the necessary concentrations in the respective tests.

Determination of lethal dose of YE for mice

The 50% lethal dose (LD₅₀) of YE for mice was determined according to the method of Reed and Muench (21). Four mice were injected from each dilution of bacterial suspensions. The pellets were resuspended in PBS in order to obtain a density 0.87 (at 650 nm) corresponding to log. CFU/ml=9.2.

Infections in mice

Sublethal and lethal YE infections were induced by i.p. inoculation of 1.056 x 10⁵ CFU (LD₅₀) or 2.112 x 10⁵ CFU (LD₁₀₀) respectively, per mice.

Humicola lutea Cu/Zn superoxide dismutase (HLSOD)

The fungal strain *Humicola lutea* 103 from the mycological collection of the Institute of Microbiology, Sofia, was used as a source for SOD production. The method of cultivation of this strain, the enzyme extraction and purification procedures are described elsewhere (1). The purified enzyme corresponds to a Cu/Zn SOD, homodimeric glycoprotein with a molecular mass of approximately 31 700 Da (1).

Protective effect of HLSOD on the mortality of mice with YE lethal infection

HLSOD was injected in mice intra-peritoneally at doses of 20U, 30U or 40 U (groups 1, 2 and 3 respectively) two times in a week, one week prior the lethal infection and 3 weeks during the infection. The mortality percent (MP) was followed till the day 27 compared to the infected untreated controls. The dose with the best protective effect on the mortality of the lethally infected mice was used for the next experiments on the immune-protective action of HLSOD on mice with a sublethal YE infection.

Protective effect of HLSOD on mice with YE sublethal infection

Experimental animals were separated in 4 groups: group 1 - sublethally YE infected mice treated with 40 U HLSOD; group 2 - sublethally YE infected mice without treatment; group 3 - uninfected mice treated by 40U HLSOD; group 4 - control - uninfected mice. Bacteriological and immunological parameters of the infected mice were examined at days 5th, 10th, 15th and 20th after infection.

Blood cultures

Blood samples obtained aseptically from the Plexus orbicularis of each animal, were inoculated in 5 ml volumes of BHI and cultivated at the above mentioned conditions. Afterwards the blood cultures were plated on Tryptic soy agar (TSA) (Difco) and cultivated again at 26°C for 24 h. Single colonies were tested for agglutination by rabbit polyclonal anti-YE antiserum (agglutination titer 1:16).

Bacterial clearance

Fragments from liver, spleen and small intestine (SI) of the infected animals were aseptically removed, weighed and homogenized separately in sterile saline. Than 0,1 volumes of serial 10 fold dilutions were plated on *Yersinia* selective medium (CIN agar, Oxoid). The plates were cultivated at 25°C for 48 h and colonies were confirmed by slide agglutination test with specific rabbit antiserum. The number of colonies were calculated for 1g weight.

Macrophage functional studies

Macrophage functions of mice with sublethal *Yersinia* infection, injected with the optimal protective dose of 40 U HLSOD per animal were followed.

a) Harvesting of peritoneal macrophages

Peritoneal macrophages were collected by washing procedure of peritoneal cavity by ice cold HBSS (pH 7.2) (10 ml per animal). After two washings the macrophage number was determined in a Burkens camera. Afterwards the cells were adjusted to the necessary concentration for the following examinations.

b) In vitro phagocytosis by peritoneal macrophages

Macrophages in concentration 1x10⁷ cells ml⁻¹ in RPMI 1640 medium (Sigma; Dieseldenhofen, Germany) without supplements were allowed to adhere on glass slides for 2 h at 37°C in humidified 5% CO₂ atmosphere. After washing with phosphate-buffered saline (PBS), fresh complete RPMI 1640 medium and suspension of killed *S. aureus* Smith cells was added to the macrophage layer (ratio 1:20 between macrophages and bacteria). The cells were allowed to interact 1 h under the same conditions, washed and stained according to the method of Pappenheim (19).

Phagocosis index (PI) (%) and phagocytic number (PN) were calculated according to the formula, proposed by Ossada *et al.* (18).

c) Killing effect of peritoneal macrophages

The killing effect of macrophages against the reference strains *S. aureus* 209 and *Escherichia coli* O₁₂₄ were estimated according to the method of Visser *et al.* (29). Briefly, macrophages at concentration 1 x 10⁶ ml⁻¹, suspended in RPMI 1640 medium, supplemented by 10% Fetal calf serum (Sigma; Dieseldenhofen, Germany), L-glutamine and antibiotics, were mixed with 5x10⁷ suspension of bacterial cells (*S. aureus* 209 or *E. coli* O₁₂₄) in saline. The ratio between macrophages and bacteria was 1:50, and the volume of samples was 0,5 ml. The cells were allowed to interact for 2 h or 24 h at 37°C in 5% CO₂ atmosphere. Bacterial cells and macrophages, at the same ratio, incubated at 4°C for 2 h were used as controls. After incubations the macrophage layers were washed and osmotically destructed by distilled water. The cell suspensions were then diluted 10-fold and cultivated on Bacto agar (Difco) at 37°C for 24 h. The number of colony forming units (CFU) from each sample was counted. The killing index (KI) was calculated as a ratio:

$$KI = \frac{\text{Initial CFU number (incubation at 4°C)}}{\text{CFU number after incubation at 37°C}}$$

Migration of peritoneal macrophages

Migration ability of peritoneal macrophages in complete RPMI 1640 medium was estimated by the method of Leu *et al.* (15). Macrophage suspension 6 x 10⁷ ml⁻¹ was introduced in glass capillaries covered by silicon (75/1 mm, for clinical laboratory use, DDR production). One side of the capillaries was stopped by bee wax. After centrifugation at 500 rpm for 5 min, the capillaries were cut at the levels of the visible macrophage cell pools and put into plastic plates containing complete RPMI 1640 medium. After cultivation at

37°C in 5% CO₂ thermostat the migration fields of macrophages were estimated by planimeter.

Hydrogen peroxide in peritoneal macrophages

Hydrogen peroxide (H₂O₂) production by peritoneal macrophages was determined according to the method of Pick and Mizel (20). Briefly, macrophage cells were collected from the peritoneal cavity, washed and suspended in Hanks' balanced salt solution (HBSS), containing 10% fetal bovine serum at a cell density of 4 × 10⁶ ml⁻¹. Cell suspensions were distributed to the wells of a 96-well plate (Falcon) (4 × 10⁵ per well). After incubation at 37°C, in 5% CO₂ thermostat for two hours the non-adherent cells were removed by washing with HBSS without phenol red. Reaction mixture was added (0.1 ml/well), containing 200 mg/ml phenol red and 50 mg/ml horseradish peroxidase type VI-A (Sigma) in HBSS without phenol red. After 45 min the reaction was stopped by addition of 10 ml/well 1N NaOH and the absorbance was read at 620 nm on an ELISA reader (Organon Teknika). For calculations, a standard curve with H₂O₂ concentrations from 5 to 50 mM was used.

Phagocytic indices of blood PMNs

The phagocyte indices (PI) of blood PMNs of the experimental animals were estimated according to the method of Vulchanov (31). Briefly, 0.1 ml blood obtained from Plexus orbicularis was immediately mixed with 0.05 ml 2% sterile sodium citrate solution. Then 0.05 ml 2 × 10⁶ suspension of killed cells of *S. aureus* Smith was added and the cells were allowed to interact at 37°C for 1 h. Smears from each sample were prepared and stained according to the method of Pappenheim (1911). Phagocytic indices of PMNs were calculated by formula of Ossada *et al.*, (18).

Statistical analysis

Differences between the results of experimental groups were analyzed by the Student's t-test. The data are presented as mean arithmetical values ± SD. P < 0.05 was accepted statistically significant. All experiments were performed in triplicate.

RESULTS

It was found that the enzyme Cu/Zn SOD isolated from the fungal strain *Humicola lutea* produced a protective effect on the survivability of mice with a lethal *Yersinia enterocolitica* O:8 infection. The most pronounced was the effect of 40U HL SOD, compared to the effects of the application of doses 20 U and 30 U HL SOD. The % mortality (MP) in the control group of the YE infected animals without treatment was 100% at day 5th, while the MP of the infected animals treated with 40 U of SOD was 17%. The mean survival time of animals in the group of mice treated with 40 U HL SOD was more than 27 days in 17% of animals. Mice from groups treated with lower doses HL SOD (20 U and 30 U) showed 100% mortality at day 17th of observation (Fig. 1). No mortality in the group of uninfected animals treated with HLSOD was observed (data not shown).

It was found that blood cultures isolated from both treated and untreated infected mice were positive in 50% until the day 7th of the observation. After day 14th until the end of investigation 25% positive blood cultures in HL SOD treated and infected group was found, when in the group of the infected mice without treatment positive blood cultures remained 50% until the day 14th. At the end of observation (day 21st) 25% blood cultures in both infected groups of animals were established (Table 1).

It was found that HLSOD treatment induced an increase of the peritoneal macrophage number both in healthy and infected animals with a maximum at day 15th (macrophages 20.2 × 10⁶ and 16.3 × 10⁶, healthy controls and infected being 13.2 × 10⁶ and 9.0 × 10⁶ respectively)(Fig. 2).

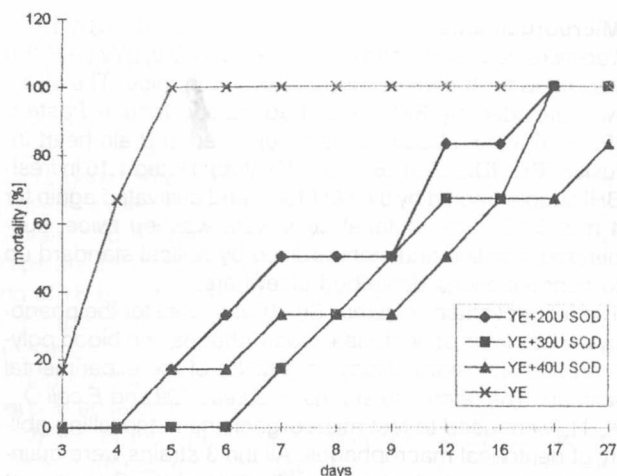


Figure 1. Mortality (%) of *Yersinia enterocolitica* O:8 pYV (+) infected mice, treated by Cu/Zn superoxide dismutase, isolated from the fungal strain *Humicola lutea* 103. Experimental groups: 1 - YE infected mice treated by 20 U HLSOD, one week prior the infection (2 injections) and 2 weeks after the day of infection (2 injections weekly); 2 - YE infected mice treated by 30 U HL SOD by the scheme of the animals in the group 1; 3 - YE infected mice treated by 40 U HL SOD according to the scheme of the animals in group 1; 4 - healthy mice without treatments.

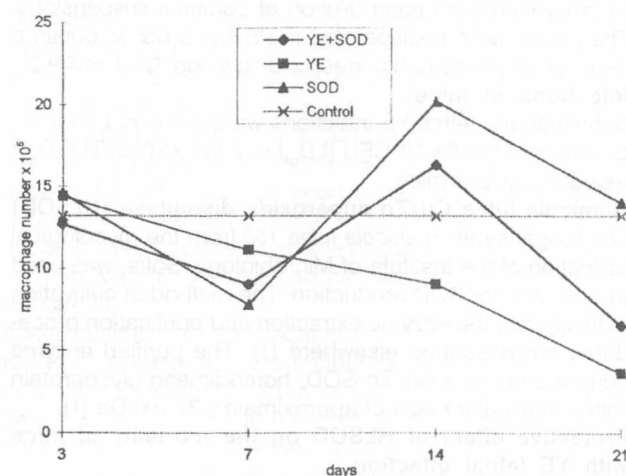


Figure 2. Number of the peritoneal macrophages mice treated by 40 U HL SOD. Experimental groups: 1 - YE infected mice, treated by 40 U HL SOD one week prior the infection (2 injections), and two weeks after the day of infection (2 injections weekly); 2 - YE infected mice without treatment; 3 - healthy mice treated by 40 U HL SOD according to the scheme of the animals in group 1; 4 - healthy mice without treatments.

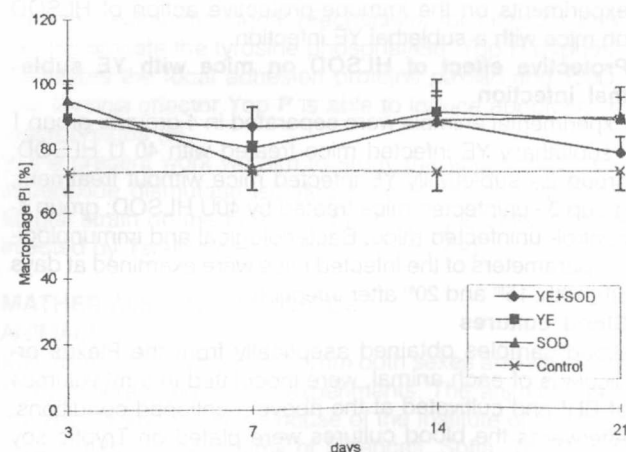


Figure 3. Phagocyte index (%) of the peritoneal macrophages of YE infected mice, treated by HL SOD. Experimental groups: as in Figure 2.

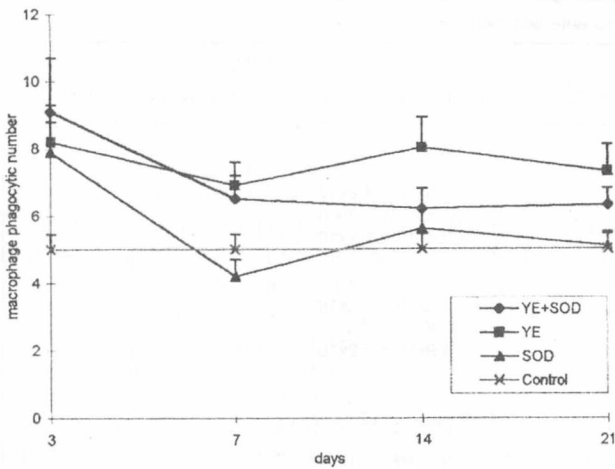


Figure 4. Phagocyte number (bacterial cells per one macrophage) of peritoneal macrophages from YE infected mice, treated by HL SOD. Experimental groups: as in Figure 2.

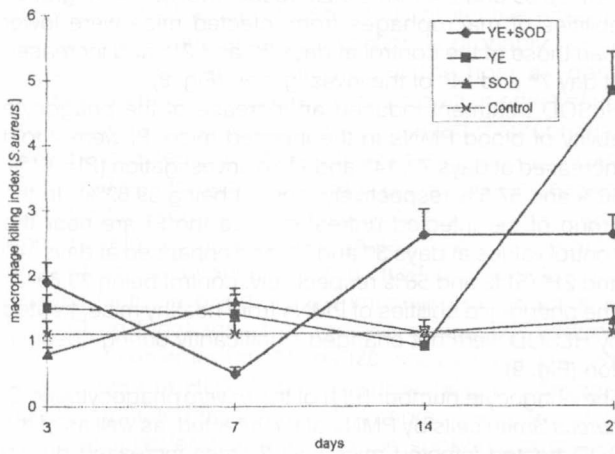


Figure 5. Killing index against *S. aureus* 209 of peritoneal macrophages from healthy and YE infected mice, treated by HL SOD. Experimental groups: as in Figure 2.

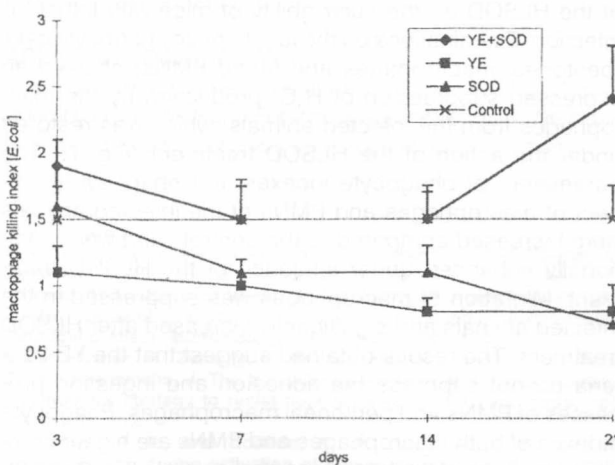


Figure 6. Killing index against *E. coli* O₁₂₄ of peritoneal macrophages from healthy and YE infected mice, treated by HL SOD. Experimental groups: as in Figure 2.

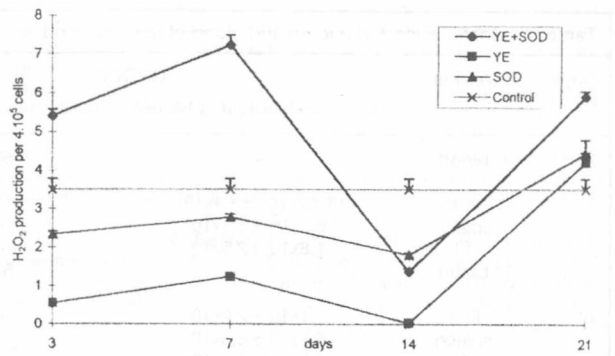


Figure 7. Spontaneous H₂O₂ production (mM per 4x10⁵ cells) by peritoneal macrophages from YE infected mice, treated by HL SOD. Experimental groups: as in Figure 2.

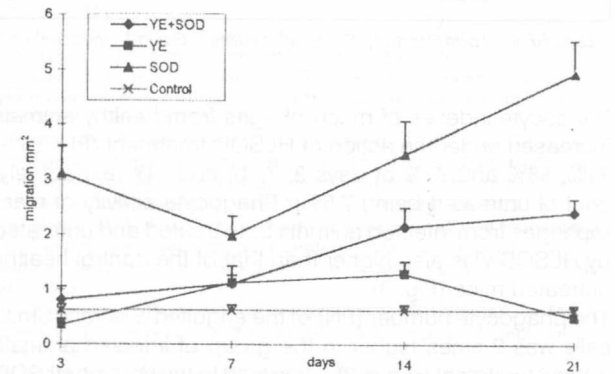


Figure 8. Migration (mm²) of peritoneal macrophages from YE infected mice treated by HL SOD. Experimental groups: as in Figure 2.

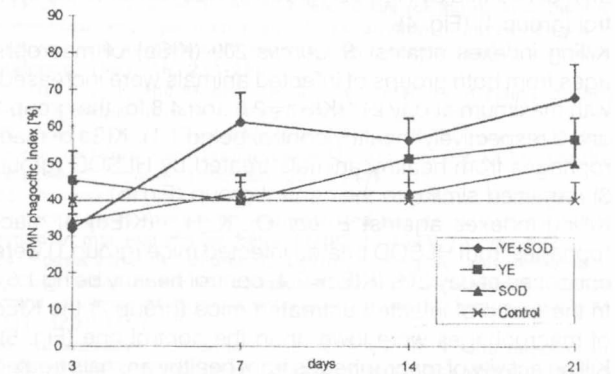


Figure 9. Phagocyte index (%) of blood PMNs from YE infected mice, treated by HL SOD. Experimental groups: as in Figure 2.

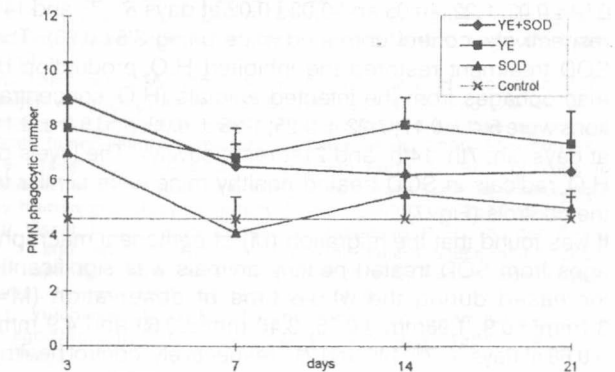


Figure 10. Phagocyte number (engulfed bacterial cells per one PMN cell) of PMNs from YE infected mice. Experimental groups: as in Figure 2.

Table 1. Bacterial load of viscera and blood of mice infected with *Yersinia enterocolitica* 0:8 strain

Days p.i.	Viscera	GROUP 1		GROUP 2	
		Bacterial load(cfu/g tissue)	Positive hemoculture (%)	Bacterial load(cfu/g tissue)	Positive hemoculture (%)
2	blood	-	50	-	50
7	liver	$1.02 \times 10^4 \pm 1.4 \times 10^5$	-	$3.6 \times 10^5 \pm 4.2 \times 10^4$	-
	spleen	$5.2 \times 10^5 \pm 3.6 \times 10^4$	-	$1.06 \times 10^5 \pm 1.8 \times 10^4$	-
	SI	$1.8 \times 10^2 \pm 2.5 \times 10^1$	-	$9.4 \times 10^1 \pm 1.2 \times 10^1$	-
	blood	-	50	-	50
14	liver	$1.1 \times 10^5 \pm 2.9 \times 10^4$	-	$5.4 \times 10^5 \pm 7.3 \times 10^4$	-
	spleen	$6.7 \times 10^5 \pm 8.3 \times 10^4$	-	$7.06 \times 10^5 \pm 1.1 \times 10^5$	-
	SI	$4.5 \times 10^4 \pm 6.1 \times 10^3$	-	$3.9 \times 10^5 \pm 5.2 \times 10^4$	-
	blood	-	25	-	50
21	liver	$6.3 \times 10^4 \pm 5.7 \times 10^3$	-	$1.08 \times 10^6 \pm 2.1 \times 10^5$	-
	spleen	0	-	0	-
	SI	$1.3 \times 10^5 \pm 2.5 \times 10^4$	-	$3.3 \times 10^5 \pm 4.4 \times 10^4$	-
	blood	-	50	-	25

Legend: p.i. - post infection; SI - small intestine; Group 1 - infection + SOD; Group 2 - infection.

Phagocyte indexes of macrophages from healthy animals increased under the action of HLSOD treatment (PI= 95%, 74%, 88% and 89% at days 3, 7, 14 and 21st respectively, control untreated being 7.5%). Phagocyte activity of macrophages from infected animals both treated and untreated by HLSOD was also higher than that of the control healthy untreated mice (Fig. 3).

The phagocyte number (PN) of the engulfed *S.aureus* Smith cells was 2 times higher in the group of infected animals without treatment (group 2) compared to the control. HLSOD treatment did not change the number of the engulfed bacterial cells (PN) in the macrophages of the infected animals (group 1). The PN of the macrophages from healthy animals treated by HLSOD (group 3) was similar to the control (group 4) (Fig. 4).

Killing indexes against *S. aureus* 209 (KISa) of macrophages from both groups of infected animals were increased, with maximum at day 21st (KISa=2.6 and 4.8 for the group 1 and 2 respectively; healthy control being 1.1). KISa of macrophages from healthy animals treated by HLSOD (group 3) remained similar to the control group (Fig. 5).

Killing indexes against *E. coli* O₁₂₄K₇₂H₃₀ (KIEc) of macrophages from HLSOD treated infected mice (group 1) were enhanced at day 21st (KIEc=2.4, control healthy being 1.5). In the group of infected untreated mice (group 2) the KIEc of macrophages were lower than the control one (Fig. 6). Killing activity of macrophages from healthy animals treated by HLSOD remained similar to the control (Fig. 6).

A suppression of H₂O₂ production by the macrophages of the infected animals up to day 15th of the investigation was observed (H₂O₂ concentrations in mM/ 4x10⁵ cells = 0.54±0.02, 1.22±0.05 and 0.02±0.02 at days 5th, 7th and 14th respectively, control untreated mice being 3.5±0.28). The SOD treatment restored the inhibited H₂O₂ production of macrophages from the infected animals (H₂O₂ concentrations were 5.4±0.17; 7.22±0.25; 1.36±0.08, and 5.9±0.14 at days 5th, 7th, 14th, and 21st respectively). The levels of H₂O₂ radicals in SOD treated healthy mice were similar to the controls (Fig. 7).

It was found that the migration (M) of peritoneal macrophages from SOD treated healthy animals was significantly increased during the whole time of observation (M= 3.1mm²±0.9, 1.98mm²±0.35, 3.46 mm²±0.69 and 4.9 mm²±0.68 at days 3rd, 7th, 14th and 21st respectively, control healthy being 0.62±0.17 mm²). The treatment by SOD enhanced also the migration ability of macrophages from the YE infected animals with maximums at days 14th and 21st (M=2.14

mm²±0.35 and 2.4 mm²±0.25, respectively). The migration abilities of macrophages from infected mice were lower than those of the control at days 3rd and 21st and increased at day 7th and 14th of the investigation (Fig. 8).

HLSOD treatment induced an increase of the phagocyte ability of blood PMNs in the infected mice. PI were 2 fold increased at days 7th, 14th and 21st of investigation (PI= 61%, 56% and 67.5% respectively, control being 39.69%). In the group of the infected untreated mice the PI are near the control values at days 3rd and 7th, and enhanced at days 14th and 21st (51% and 56% respectively, control being 39.69%). The phagocyte abilities of PMNs from healthy mice, treated by HLSOD were not changed significantly during observation (Fig. 9).

The phagocyte number (PN) of the *in vitro* phagocytosed *S. aureus* Smith cells by PMNs of the infected, as well as of the SOD treated infected mice was 2 times increased during observation. The PN of the healthy HLSOD treated mice was similar to the control (Fig. 10).

DISCUSSION

The results of the investigation show the protective effect of the HLSOD on the survivability of mice with lethal YE infection. Examinations on the functions of phagocyte cells (peritoneal macrophages and blood PMNs) showed an expressed suppression of H₂O₂ production by the macrophages from the infected animals, which was restored under the action of the HLSOD treatment (Fig. 7). The parameters of phagocyte indexes and phagocyte numbers of macrophages and PMNs of the infected animals were increased compared to the controls and were additionally enhanced under influence of the HLSOD treatment. Migration of macrophages was suppressed in the infected animals and significantly increased after HLSOD treatment. The results obtained suggest that the YE bacteria do not suppress the adhesion and ingestion processes of PMNs and peritoneal macrophages. Phagocyte indexes of both macrophages and PMNs are higher compared to those of the healthy control animals (Fig.3 and 9). The phagocyte number of the *in vitro* engulfed *S.aureus* Smith bacteria by both classes of phagocyte cells is also higher compared with the control one. It is known that the most important anti-phagocyte factor suppressing bactericidal activity of phagocyte cells in the YE infected animals are the Yops proteins. These proteins suppress also the host receptor-dependent superoxide anion production by human granulocytes (30).

Pathogenic *Yersinia* export 14 Yop proteins into cytosol of eukariotic cells, by type III pathway. Some of them, named effector Yops, are targeted into macrophages, thereby preventing phagocytosis and allowing bacterial replication into lymphoid tissue. Until now YopB and YopD were thought to promote the important of the effector Yops into eukariotic cytosol. Yop B and YopR are dispensable for the injection of toxins into eukariotic cells (14).

According to the investigations of Ruckdeschel *et al.* (26) interaction of YE with macrophages cell lines leads to macrophage cell death through apoptosis. Our previous experiments showed that *in vitro* and *in vivo* interactions of rat peritoneal macrophages with YE did not induce apoptosis and bacterial cells can be eliminated by both mouse and rat peritoneal macrophages (12).

Mn-cofactored superoxide dismutase (SOD-A) is an important factor of the virulence of YE (24). Resistance of YE to exogenous oxygen radicals, produced by phagocytes involves the SOD-A. The important role of SOD-A for pathogenicity of YE could be also due to detoxification of endogenous metabolically produced oxygen radicals, which are encountered by YE during the invasion of the host (24).

The established in the present study protective effect of SOD on mice with lethal YE infection is probably due to the participation of the enzyme in the host oxidant-antioxidant balance. As a result a restoration effect of the suppressed H_2O_2 production by peritoneal macrophages from the infected and HLSOD treated mice was achieved (Fig. 7). The increased peroxide production is responsible for the increased killing activities of macrophages from the infected and HLSOD treated animals. The presence of polysaccharide chain in HLSOD molecule (1) is another factor involved in the immunoprotective action of HLSOD on functions of phagocyte cells in YE infected mice. Our earlier investigations (28) showed similarly protective effect of HLSOD on the survivability of hamsters with transplanted myeloid Graffi tumors. The preparation exhibited a restoration effect on the suppressed phagocyte activities of peritoneal macrophages and blood PMNs, as well as on the *in vitro* proliferation abilities of the spleen B-lymphocytes (28).

The obtained results suggest the possible use of anti-oxidant enzymes in the therapy of some infections caused by phagocytosis-resistant bacteria.

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REFERENCES

- Angelova, M., Dolashka-Angelova, P., Ivanova, E., Serkedjieva, J., Slokoska, L., Pashova, S., Toshkova, R., Vassilev, S., Simeonov, I., Hartmann, H.-J., Stoeva, S., Weser, U., and Voelter, W. A novel glycosylated Cu/Zn - containing superoxide dismutase: production and potential therapeutic effect. *Microbiology*, 2001, 147: 1641-1650.
- Aepfelbaier, M., Zumbühl, R., Ruckdeschel, K., Jacobi, C. A., Barz, C., and Heesemann, J. The tranquilizing injection of *Yersinia* proteins: a pathogen's strategy to resist host defense. *Biol. Chem.* 1999, 380 (7-8): 795-802.
- Berkow, R. L., and Dodson, R. W. Tyrosine-specific protein phosphorylation during activation of human neutrophils. *Blood*, 1990, 75: 304-310.
- Black, D. S., and Bliska, J. B. The Rho GAP activity of the *Yersinia* pseudotuberculosis cytotoxin YopE is required for antiphagocyte function and virulence. *Mol. Microbiol.* 2000, 37(3): 515-527.
- Bliska, J. B., Guan, K. L., Dixon, J. E., and Falkov, S. Tyrosine phosphate hydrolysis of host proteins by essential *Yersinia* determinant. *Proc. Natl. Acad. Sci. (USA)*, 1991, 88: 1187-1191.
- Bolin, I. L., Norlander, L., and Wolf-Watz, H. Temperature-inducible outer-membrane protein of *Yersinia pseudotuberculosis* and *Yersinia enterocolitica* is associated with the virulence plasmid. *Infect. Immun.* 1982, 37: 506-512.
- Bottone, E. J. *Yersinia enterocolitica*: a panoramic view of charismatic microorganism. *Crit. Rev. Microbiol.* 1977, 5: 211-241.
- Boyd, A. P., Grosdent, N., Totenmeyer, S., Gemjen, C., Blever, S., Iriart, M., Lambermort, I., Octave, G. M., and Cornelis, G. R. *Yersinia enterocolitica* can deliver Yop proteins into a wide range of cell types: development of a delivery system for heterologous proteins. *Eur. J. Cell. Biol.*, 2000, 79(10): 659-671.
- Chanock, S. J., El Benna, J., Smith, R. M., and Babior, B. M. The respiratory burst oxidase. *J. Biol. Chem.*, 1994, 269: 24519-24522.
- China, B., Nguen, B. T., Debreyer, M., and Cornelis, G. R. Role of Yad A in resistance of *Yersinia enterocolitica* to phagocytosis by human polymorphonuclear leukocytes. *Infect. Immun.*, 1994, 62: 1275-1281.
- Fallman, M. K., Andersson, K., Hakanson, S., Magnusson, K. E., Stendahl, O., and Wolf-Watz, H. *Yersinia pseudotuberculosis* inhibits Fc-receptor-mediated phagocytosis in J774 cells. *Infect. Immun.*, 1995, 63: 3117-3124.
- Ivanova, E., Yanchev, I., Naidenski, H., Toshkova, R., Dimitrova, P., and Manov, V. 2000. Studies on the interactions of immunostimulated macrophages and *Yersinia enterocolitica* O:8. *Can. J. Microbiol.*, 2000, 46: 218-228.
- Kohl, S., Jacobson, A., and Nahmias, A. *Yersinia enterocolitica* infections in children. *J. Pediatr.*, 1977, 89:77-79.
- Lee, V. T., and Schneewind, O. Type III machines of pathogenic *Yersinia* secrete virulence factors into extra-cellular milieu. *Mol. Microbiol.*, 1999, 34(1): 196.
- Leu, W. R., Edeston, W. F., Hadden, J. W., and Good, R. A. Mechanism of action of migration inhibitory factor (MIF). Evidence for a receptor for MIF present on peritoneal macrophages but not on alveolar macrophages. *J. Exp. Med.*, 1974, 136:589-563.
- Marks, M. I., Pai, C. H., Lafleur, L., Lackman, L., and Hammerberg, O. *Yersinia enterocolitica* gastroenterocolitis. A prospective study of clinical, bacteriologic and epidemiologic features. *J. Pediatr.*, 1980, 96: 26-31.
- Michiels, T. P., Watian, P., Brasseur, R., Ruyschaert, J. M., and Cornelis, G. Secretion of Yop proteins by *Yersinia*. *Infect. Immun.*, 1990, 58: 2840-2849.
- Ossada, Y., Mitsuya, M., Ume, T., Matsumoto, K., Otani, T., Satoh, M., Ogawa, H., and Nomoto, K. Effect of L18 MDP (Ala), a synthetic derivative of muramyl-dipeptide on nonspecific resistance of mice to microbial infections. *Infect. Immun.*, 1982, 37: 292-300.
- Pappenheim, A. *Grundris der hamatologischen Diagnostik*. 1911, Leipzig.
- Pick, E., and Mizel, D. Rapid microassays for the measurement of superoxide and hydrogen peroxide production by macrophages in culture using an automatic enzyme immunoassay reader. *J. Immun. Methods*, 1981, 46: 211-226.
- Reed, L. J., and Muench, H. A simple method of estimating fifty percent endpoints. *Am. J. Hyg.*, 1938, 27: 493-497.
- Rosenshine, I., Duronio, V., and Finlay, B. B. Tyrosine protein kinase inhibitors block invasion-promoted bacterial uptake by epithelial cells. *Infect. Immun.*, 1992, 60: 221-2217.
- Rosqvist, R., Magnusson, K. E. and Wolf-Watz, H. Target cell contact triggers expression and polarized transfer of *Yersinia* Yop E cytotoxin into mammalian cells. *EMBO J.*, 1994, 13: 964-972.
- Rosenkamp, A., Bittner, T., Leitritz, I., Sing, A., and Heesemann, J. Contribution of the Mn-cofactored superoxide-dismutase (SOD-A) to the virulence of *Yersinia enterocolitica*. *Infect. Immun.*, 1997, 65 (11): 4705-4711.
- Ruckdeschel, K., Roggenkamp, A., Schubert, S., and Heesemann, J. Differential contribution of *Yersinia enterocolitica* virulence factors to evasion of microbicidal action of neutrophils. *Infect. Immun.*, 1996, 64: 724-733.
- Ruckdeschel, K., Roggenkamp, A., Lapont, V., Mangeat, P., Heesemann, J., and Rouot, B. Interaction of *Yersinia enterocolitica* with macrophages leads to macrophage cell death through apoptosis. *Infect. Immun.*, 1997, 65: 4813-4821.
- Starley, S. C., Skrzipek, E., Planí, G. V., and Bliska, J. B. Yops of *Yersinia* spp. pathogenic for humans. *Infect. Immun.*, 1994, 61: 3105-3110.
- Toshkova, R. A., Dimitrova, P. A., Ivanova, E. H., Dolashka, P. A., Angelova, M. B., Pashova, S. B., Slokoska, L. S., and Voelter, W. T. Immunoprotective effect of Cu/Zn superoxide dismutase on myeloid Graffi tumor-bearing hamsters. *Z. Naturforsch.*, 2000, 55c: 649-656.
- Visser, L. G., Anema, A., and van Furth, R. Role of Yops in inhibition of phagocytosis and killing of opsonized *Yersinia enterocolitica* by human granulocytes. *Infect. Immun.*, 1995, 63: 2570-2575.
- Visser, L. G., Siemonsbergen, E., Nibbering, P. H., Van den Broek, P. J., and van Furth, R. Yops of *Yersinia enterocolitica* inhibit receptor-dependent superoxide anion production by human granulocytes. *Infect. Immun.*, 1999, 67 (3): 1245-1250.
- Vulchanov, V. 1954. To the method of examination of the phagocytic activity of blood. *Investig. Biol. Inst.*, 1954, 5: 411-413 (in Bulg.).

COMPARISON OF THE SUSCEPTIBILITY TO 11 ANTIMICROBIAL AGENTS OF CLINICAL STRAINS BETA-HAEMOLYTIC GROUP A AND B STREPTOCOCCI ISOLATED FROM ADULTS AND REVIEW OF THE LITERATURE

B. Girgitzova¹, D.Chankova²

1. Department of Clinical Microbiology, Emergency Medical Institute
„N.I.Pirogov“

2. BB - NCIPD, Ltd

SUMMARY

Increasing antimicrobial resistance of group A and group B streptococci to some antibiotics has been observed during the last decade in Europe and worldwide. The aim of this study was to evaluate the susceptibility to antimicrobial agents of clinical strains group A and B streptococci isolated from adults and to compare our results to those from other parts of the world. Using the standard agar dilution method we studied the susceptibility to 11 antimicrobial agents of 156 *S.pyogenes* strains and 138 *S.agalactiae* strains isolated in 1999-2001. All isolates were susceptible to penicillin, cephalothin, cefuroxime and cefotaxime. The sensitivity to erythromycin in both groups was almost equal - about 96%, while clindamycin resistance (1,45%) was found only in group B streptococci. Resistance to chloramphenicol (3,62%), ciprofloxacin (1,45%), gentamicin (3,62%) and intermediate susceptibility to it (62,32%) was demonstrated again by group B streptococci. They also were resistant to tetracycline (92,76% against 9,62% for group A) and trimethoprim-sulfamethoxazole (100% and 30,13% respectively). Among comparable studies concerning other countries our results demonstrate a bit lower or near the average values reported from Europe.

Key words: group A streptococci, group B streptococci, antimicrobial susceptibility and resistance, MIC range.

INTRODUCTION

Group A (*GAS*; *S.pyogenes*) and B (*GBS*; *S.agalactiae*) streptococci cause many illnesses and clinical syndromes in neonates, children and adults and the monitoring of their susceptibility to antimicrobial agents is very important. Although the antimicrobial resistance among bacteria continues to increase and to be a clinical problem the beta-haemolytic streptococci remained remarkably susceptible to many antibiotics. Both group A and B streptococci are generally considered to be universally susceptible to the penicillins. As far as it is known there has never been isolated a single penicillin resistant group A streptococcal strain from a clinical source. Available data indicate that the minimal inhibitory concentrations (MICs) of penicillin for group A streptococci have not changed during the last 4 decades (15).

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ABBREVIATIONS USED IN THIS PAPER: GAS - Group A Streptococci, GBS - Group B Streptococci

CORRESPONDING AUTHOR:

Bojana Girgitzova Ph.D.

Department of Clinical Microbiology

Emergency Medical Institute "N.I.Pirogov"

21, Makedonia Blvd.

1606 Sofia, BULGARIA

Tel.: +359 2/ 5153 458

Fax: +359 2/ 986 73 76

E-mail: janico@hotmail.com

Although penicillin is the first choice for the treatment of pharyngeal and other infections due to those organisms, erythromycin (or someone of the newer macrolides) is the second-line drug of choice and is used in penicillin-hyper-sensitive patients (30). At the same time, the increased use of erythromycin and its derivatives has been related to the increased resistance of *S.pyogenes* and *S.agalactiae* to those antibiotics (6,11,13,17).

Addition of clindamycin to the therapy of serious streptococcal infections is recommended to inhibit protein synthesis and it is also a drug of choice for chronic, recurrent pharyngitis (30). Resistance to clindamycin, tetracyclines, aminoglycosides and sulfa drugs in this two streptococcal groups is also observed in many parts of the world and it becomes clinically important (12,15,16,17,29).

Many scientists considered that the antibiotic resistance rates varied with the geographic areas, as well as in different areas of the same country (6,11,15,30). Also there is an opinion that the susceptibility of the streptococci to antimicrobial agents depends on the age of the patients (15).

The aim of this study was to evaluate the susceptibility to antimicrobial agents of clinical strains group A and B beta-haemolytic streptococci isolated from adults and to compare our results to those from other parts of the world.

MATERIALS AND METHODS

Bacterial strains: The survey includes 156 beta-haemolytic group A and 138 beta-haemolytic group B streptococcal strains isolated from adults patients /over 16 years/ of Emergency Medical Institute "Pirogov" collected from January 1999 to June 2001. The isolates were identified on the basis of colony morphology, beta-haemolysis on blood agar, bacitracin susceptibility, commercial latex agglutination test (Oxoid, England) and confirmed by Api System bioMerieux, France. Most of the strains were stored at -70° C in Todd-Hewitt broth (DIFCO Laboratories, Detroit) with 15% glycerol until testing and some of them - at room temperature after lyophilization. The isolates were from different clinical sources: blood, pleural fluid, sputum, wound, throat, vaginal, urethral swabs, urine, sperm.

Control strains included: *S.pneumoniae* ATCC 49619 and *S.aureus* ATCC 25923.

Antimicrobial substances: penicillin G, cephalothin, cefuroxime, cefotaxime, erythromycin, clindamycin, chloramphenicol, tetracycline, gentamicin, ciprofloxacin, trimethoprim/sulphamethoxazol (TMP-SMZ). From the macrolide group only erythromycin was tested since the susceptibility to it can be used to predict susceptibility to clarithromycin and azithromycin. All antimicrobial substances were supplied as standard reference powders with defined activity. Stock solutions were prepared according to the recommendations of NCCLS (22) and the drugs were incorporated into the agar medium in serial twofold concentrations.

Determination of antibiotic susceptibility: The antibiotic susceptibility of the strains was determined by agar dilution method with Mueller-Hinton medium (DIFCO Lab, Detroit, Mich.) supplemented with 5% defibrinated sheep blood as it is recommended by NCCLS (22). The final inocula of ~ 1x10⁴ to 5x10⁴ cfu was applied by means of a multipoint inoculator and plates were incubated overnight at 37°C in an atmosphere of 5%CO₂. The MIC of the antibiotic was defined as the lowest concentration which inhibited the visible growth of the organism. NCCLS (22) breakpoints were used for all antimicrobial agents. All control strains gave MICs within two dilution steps of target values.

RESULTS

The susceptibility (in percent and MIC range) to 11 antimicrobial agents of the group A and B streptococci isolated from adults is given in table 1 and table 2.

As it was expected almost all streptococcal strains (group A - 99,36%, group B - 96,38%) were sensitive to penicillin with one and the same MIC range (0,008 - 0,06 mg/L) but there were many isolates with intermediate susceptibility to penicillin in GBS (3,62%) with MIC range 0,25-2 mg/L as in GAS (0,64%) with MIC range (0,25-0,5mg/L). No resistance to penicillin was observed. The susceptibility to erythromycin in both groups was almost equal (about 96%) but the upper limit of MIC values for the sensitive strains was twofold dilutions higher (0,25mg/L) in GBS. The resistance to this antibiotic was 2,57% in GAS with MICs up to 4mg/L against 2,17% resistance in GBS with MICs - 2mg/L.

Group A streptococci as a whole were susceptible to clindamycin (MIC range 0,015-0,06mg/L) while 1,45% of GBS were resistant to it (MIC \geq 2mg/L) and in addition in 2,17% of them intermediate sensitivity was observed. The two clindamycin resistant strains were resistant to erythromycin too. All *S.pyogenes* isolates expressed 100% susceptibility to cephalosporins (I, II and III generation) without a significant differences in their MIC values (0,008-0,06 mg/L), while in GBS the same sensitivity revealed only to cefotaxime (MIC range 0,008-0,06mg/L). A slightly higher MICs could be seen among cefuroxime sensitive (0,008-0,125mg/L) and cephalothin sensitive (0,015- 0,25mg/L) GBS strains. An intermediate susceptibility to cefuroxime (0,91%) and cephalothin (1,36%) was demonstrated only in this group. Chloramphenicol resistance was documented in 3,62% of group B streptococci while intermediate sensitivity was spread almost equally among the isolates of both groups. Although all streptococci tested had the same MIC range (0,5-64mg/L) for tetracycline there were significant differences in the susceptibility of the two groups. The prevalence of sensitive strains (88,46%) was well expressed in *S.pyogenes* while on the contrary 92,76% of GBS were resistant to this antibiotic.

The rates of susceptibility to gentamicin were nearly three times higher in group A (98,08%) than in GBS (36,96%). Characteristic for GBS was the availability of great amount of isolates (62,32%) with intermediate sensitivity.

TMP-SMZ MICs of *S.pyogenes* showed bimodal distribution - about three parts (69,87%) of the isolates were sensitive to this drug (MICs 0,06/1,2-2/38mg/L) while all GBS were resistant to it with very high MIC values (128/2432-256/4864mg/L). A resistance to ciprofloxacin was found only in two strains of GBS (1,45%).

DISCUSSION

Penicillin is the recommended drug of choice in the most cases of GAS pharyngitis in the prevention of acute rheumatic fever, in cases of GAS and GBS bacteremia and sepsis, deep tissue infections, osteomyelitis, meningitis, empyema due to this two groups streptococci. GAS are susceptible to penicillin independently of their isolation - neonates, adults, patients with invasive diseases or carriers. The amount (in percent) and MIC values of the susceptible to penicillin GAS obtained in this study agree with those reported by Awand (2), Cornaglia (6), De Azavedo (7), Kaplan (15), Orden (23) and York (30). *S.pyogenes* continues to be sensitive to this drug with MIC range (0,008 - 0,012mg/L). In spite of the demonstration of penicillin tolerance by some *S.pyogenes* strains (12,24) which is considered to cause treatment failures in pharyngitis, the consensus is that this phenomena does not have clinical relevance with respect to GAS (12,15,24).

As many authors report (3,8,9,15,16,17) GBS are also highly susceptible to penicillin (MIC 0,008-0,125mg/L), but intermediate sensitivity is more often met (2-8%) among those isolates (4,8,15). Our results confirm this fact - 3,62% GBS showed intermediate susceptibility (MIC 0,25-2mg/L). The penicillin tolerance is presented in GBS too (4,15) and in Spain it even reaches 17% (4), but as for *S.pyogenes* the clinical relevance is not convincing. In summary our data document the continued universal susceptibility of group A and B streptococci to penicillin which have not changed during the past four decades, therefore it remains the best antibiotic both for prophylaxis and treatment of infections caused by those microorganisms.

Despite of this fact bacteriologic failure occurs in up to 20% of the patients treated with penicillin, and half of those cases are also a clinical failure (24). Various theories have been offered to explain this phenomena: beta-lactamase-producing bacteria, bacterial interference, re-infection and penicillin tolerance (4,24).

However for patients sensitive to beta-lactam antibiotics and for the treatment of patients who failed penicillin therapy, macrolides are often the recommended drugs. Erythromycin has traditionally been regarded as an effective and very save drug for treatment of group A and B streptococcal infections, but his increased clinical use and the prescription of its macrolide derivates mostly for the upper respiratory tract infections has been associated with the increased resistance of GAS and GBS to this antibiotic (6,8,11,15,17,26). The incidence of erythromycin resistance in GAS still remains low in most part of the world - about 5%: Israel - 1% (29), Canada - 2,1% (7), USA - less than 5% (15), France - 6,2% (5), Chile - 7,2% (25). Widespread erythromycin resistance in *S.pyogenes* has so far been reported only in a limited number of countries - Germany 12,7% (2), Finland - 16-17% (26), Spain - 14,3% (23), 20%(11), Italy - 30-35% (6), Japan - 62% (21). With the 2,57% resistance to this antibiotic and 1,28% intermediate susceptibility our results confirm the conventional opinion about the generally low rates of erythromycin resistance (~ 5%) in this group.

York and co-workers (30) notified a statistically higher rate of erythromycin resistance (39%) among the strains GAS from serious infections compared to those from pharyngitis (13%) in San Francisco Country Hospital, but the cause of this resistance has not been determined. The authors had not identified an overuse of this drug in San Francisco but they supposed that the erythromycin resistance had an association with strains that were more invasive (30). The effect of different macrolides is well demonstrated in a Spanish multicentere study in 1998 where 23,5% of *S.pyogenes* isolates were resistant to erythromycin - a 14-membered ring macrolide and azithromycin - a 15-membered ring macrolide whereas only 1% of the isolates were resistant to miocamycin - a 16-membered ring macrolide (1).

The erythromycin resistance in GBS show also variations according to the geographic area. The resistance is notably high in Taiwan - 46% (13), France 18%,21,4% (10,19), USA - 8,5-32% (in different states (17), Canada - 18% (8), while Fernandez reports of 7,4% resistance (9) and Kaplan - 10% (15). The frequency of erythromycin resistance among our GBS is low (2,17%) and the same percent of the strains are with intermediate susceptibility.

As a whole both streptococcal groups studied demonstrated high susceptibility to erythromycin - over 96%. In fact the usage of this antibiotic in our country during the last two decades was limited because of its substitution with the modern cephalosporin groups. Also all new macrolides were not available in Bulgaria. Low resistance to erythromycin (only 3%) was also documented by scientists from Japan

Table 1. Susceptibility to 11 antimicrobial agents of 156 group A streptococcal strains

Antimicrobial agents	S (n/%) MIC range (mg/L)	I (n/%) MIC range (mg/L)	R (n/%) MIC range (mg/L)
Penicillin G	155 (99,36) 0,008-0,06	1 (0,64) 0,25-0,5	—
Cephalothin	156 (100) 0,015-0,06	—	—
Cefuroxime	156 (100) 0,008-0,03	—	—
Cefotaxime	156 (100) 0,008-0,03	—	—
Erythromycin	150 (96,15) 0,008-0,06	2 (1,28) 0,5	4 (2,57) 1-4
Clindamycin	156 (100) 0,015-0,06	—	—
Gentamicin	153 (98,08) 1-4	3 (1,92) 8	—
Tetracycline	138 (88,46) 0,5-2	3 (1,92) 4	15 (9,62) 8-64
Chloramphenicol	153 (98,08) 0,25-4	3 (1,92) 8	—
Ciprofloxacin	156 (100) 0,125-0,5	—	—
Trimethoprim/ Sulfamethoxazole	109 (69,87) 0,06/1,2-2/38	—	47 (30,13) 4/76-32/608

Note: S - sensitive, I - intermediate, R - resistant

for GBS isolated from pregnant women. There were no significant differences between the two studied periods (1985-1986 and 1999-2000) in the incidence of GBS resistance to 14 antibiotics (18). A statistically significant association was found between the local erythromycin consumption and the erythromycin resistance in GAS in Spain (11) and Finland (26) and this may be the explanation of the lower erythromycin resistance of our streptococcal strains. The routine determination of the macrolide resistance in group A and B streptococcal isolates on one hand and the confinement use of these antibiotics for prophylaxis and treatment of upper respiratory tract infections in outpatients on the other hand will preserve them for a successful applying to allergic to penicillin patients.

Some authors associated clindamycin resistance with the resistance to erythromycin (2,6,7). Our results support this thesis as the two clindamycin resistant GBS were resistant to erythromycin too. Data about clindamycin resistance in GAS and GBS show that it is more rarely met in comparison to erythromycin resistance. WHO collaborating center in Minnesota documents 5% resistance in *S. pyogenes* (15), in Italy it varies from 2,7% to 32,4% in different areas (6), in Germany - 11,3% (2), in Madrid - 1,4% (23), in Spain (multicenter study-1998) - only 0,8% (1). As for GBS a vast investigation including different institutes and centers in USA reports 12% rate of resistance; close to it is the resistance found in Canada - 8% (8), North Carolina - 9% (20); low is the resistance in Spain - 3,4% (9) and in Japan - 1% (18). The remarkably high sensitivity demonstrated by our isolates (100% in group A and 96,38% in group B) may be due to the fact that clindamycin is comparatively seldom used for the treatment of infections in Bulgaria. De Azavero et al (8) reported about a difference in the resistance to erythromycin and clindamycin in GBS isolated from neonatal blood cultures (8% and 4,5% respectively) and those isolated from vaginal swabs from adults (18% and 8%).

While our results do not indicate a notable differences in the susceptibility and MICs of *S. pyogenes* to the antibiotics from three cephalosporin generation studied (100% sensitivity and MICs - 0,008-0,06 mg/L), cefotaxime was the most effective against GBS. To our knowledge, there is no doubt

that cefotaxime is the preferable cephalosporin antibiotic for infections caused by group A and B streptococci with the highest rates of sensitivity - about 100% (2,9,15,16,17,25). It is important to mention that so far it is not observed any clinically significant resistance to the cephalosporins (15). The susceptibility to ciprofloxacin, a quinolone, which use is more frequent than is recognized, is well expressed - over 95% in both streptococcal groups (2,15,23). In our strains only 1,45% from GBS show a resistance to ciprofloxacin. Nevertheless Kaplan and his colleagues did not consider it as first line antibiotic for therapy of group A streptococci (15). Although the notable chloramphenicol sensitivity (about 100%) GAS and GBS demonstrated in the studies all over the world (7,17,23,29). Its application should be limited because of its toxicity and lack of bactericidal activity.

The average resistance of *S. pyogenes* to tetracycline is about 10% (15,23) but in Israel it reaches 41% (29). 9,62% of GAS in this study are resistant to this drug and 1,92% have intermediate sensitivity. It is known that the frequency of tetracycline resistance in GBS is remarkably higher in comparison to GAS - 95,8% (17), 97,4% (9), >80% (12). The same tendency can be seen in our results - 92,76% of those isolates were resistant to it. This fact is very important because tetracycline is still used by some primary care physicians for prophylaxis and therapy of pharyngitis. In general GAS are susceptible to gentamicin with MIC range 1-8mg/L (as our strains revealed too), while in GBS - a large cluster of strains (about 50%) are with intermediate sensitivity (9,16). Our GBS isolates from adults showed 62,38% intermediate resistance and 36,96% sensitivity.

The literature contains few and conflicting reports on the susceptibility of GAS and GBS to TMP-SMZ (14,27,29). Also high level resistance to TMP-SMZ (>1000-fold increase in MICs compared to MICs of sensitive strains) is observed among resistant strains, which more frequently cause a therapeutic failure than low level resistant. However, more clinical studies are needed to establish the role of both types of resistance to TMP-SMZ (14,27,29). So TMP-SMZ is not considered a drug for therapy of infections (pharyngitis, skin infections, pneumonia, meningitis, septicemia, etc) caused by GAS and GBS.

Table 2. Susceptibility to 12 antimicrobial agents of 138 group B streptococcal strains

Antimicrobial agents	S (n/%) MIC range (mg/L)	I (n/%) MIC range (mg/L)	R (n/%) MIC range (mg/L)
Penicillin G	133 (96,38) 0,008-0,06	5 (3,62) 0,2-2	—
Cephalothin	0,015-0,25	1	—
Cefuroxime	0,008-0,125	1	—
Cefotaxime	138 (100) 0,00-0,06	—	—
Erythromycin	132 (95,66) 0,008-0,25	3 (2,17) 0,5	3 (2,17) 2
Clindamycin	133 (96,38) 0,015-0,125	3 (2,17) 0,5	2 (1,45) 2-4
Gentamicin	51 (36,96) 2-4	86 (62,38) 8	5 (3,62) > 32
Tetracycline	7 (5,07) 0,5-2	3 (2,17) 4	128 (92,76) 8-64
Chloramphenicol	131 (94,93) 1-4	2 (1,45) 8	5 (3,62) 6 - >32
Ciprofloxacin	136 (98,55) 0,125-0,5	—	2 (1,45) 16 - >32
Trimethoprim/ Sulfamethoxazole	—	—	138 (100) 128/2432-256/4864

Note: S - sensitive, I - intermediate, R - resistant

In conclusion the results of this study showed that beta-lactams are the drugs of choice for the treatment of Group A and B streptococcal infections in adults. Comparing the two groups, GBS were less susceptible to beta-lactam antibiotics than GAS. The development of resistance to erythromycin is a clinically important problem in the management of the infections caused by these microorganisms. It also emphasizes the need for routine determination of the antimicrobial resistance in GAS and GBS in our country.

REFERENCES

- Alos JI, Aracil B, Oteo J et al. High prevalence of erythromycin-resistant, clindamycin/micamycin-susceptible (M phenotype) *Streptococcus pyogenes*: results of Spanish multicenter study in 1998. *J Antimicrob Chemother* 2000;45:605-609.
- Awand M, Hoeck H, Hahn H, Wagner J. Antimicrobial resistance in *Streptococcus pyogenes* isolates in Berlin. *J Antimicrob Chemother* 2000;46:621-623.
- Barile A, Kallen A, Wallace M. Fatal group B streptococcal meningitis in previously healthy young adult. *Clin Infect Dis* 1999;28:151.
- Bertriu C, Gomez M, Sanchez A, Cruceyra A, Romero J, Picazo JJ. Antibiotic resistance and penicillin tolerance in clinical isolates of group B streptococci. *Antimicrob Agents Chemother* 1994;38:2183-2186.
- Bingen E, Fitoussi F, Doit C, Cohen R, Tanna A, George R et al. Resistance to macrolides in *Streptococcus pyogenes* in France in pediatric patients. *Antimicrob Agents Chemother* 2000;44:1453-1457.
- Cornaglia G, Ligozzi M, Mazzariol A, Masala L, Casio G, Orefici G. Resistance of *Streptococcus pyogenes* to erythromycin and related antibiotics in Italy. *Clin Infect Dis* 1998;27(Suppl 1):S87-S92.
- De Azavedo J, Yeung R, Bast D, Duncan S, Borgia B, Low D. Prevalence and mechanisms of macrolide resistance in clinical isolates of group A streptococci from Ontario, Canada. *Antimicrob Agents Chemother* 1999;43:2144-2147.
- De Azavedo J, McGavin M, Duncan C, Low D, McGeer A. Prevalence and mechanisms of macrolide resistance in invasive and noninvasive group B isolates from Ontario, Canada. *Antimicrob Agents Chemother* 2001;45:3504-3508.
- Fernandez M, Hickman M, Baker CJ. Antimicrobial susceptibility of group B streptococci isolated between 1992 and 1996 from patients with bacteremia or meningitis. *Antimicrob Agents Chemother* 1998;42:1517-1519.
- Fitoussi F, Loukil Ch, Gros I, Clermont O, Mariani P, Bonacorsi S. et al. Mechanisms of macrolide resistance in clinical group B streptococci isolated in France. *Antimicrob Agents Chemother* 2001;45:1889-1891.
- Garcia-de-Lomas J et al. Antimicrobial susceptibilities of 1,684 *Streptococcus pneumoniae* and 2,039 *Streptococcus pyogenes* isolates and their ecological relationships: results of a 1-year (1998-1999) multicenter surveillance study in Spain. *Antimicrob Agents Chemother* 2001;45:3334-3340.
- Gerber MA. Antibiotic resistance in group A streptococci. *Pediatr Clin North Am* 1995;42:539-551.
- Hsueh P-R, Teng L-J, Lee L-N, Ho S-W, Yang P-C, Luh K-T. High incidence of erythromycin resistance among clinical isolates of *Streptococcus agalactiae* in Taiwan. *Antimicrob Agents Chemother* 2001;45:3205-3208.
- Huovinen P. Resistance to trimethoprim-sulfamethoxazole. *Clin Infect Dis* 2001;32:1608-1614.
- Kaplan EL. Recent evaluation of antimicrobial resistance of beta-hemolytic streptococci. *Clin Infect Dis* 1997; 24(Suppl 1):S89-S92.
- Liu JW, Wu JJ, Ko WC, Chuang YC. Clinical characteristics and antimicrobial susceptibility of invasive group B streptococcal infections in nonpregnant adults in Taiwan. *J Formos Med Assoc* 1997;96:628-633.
- Lin FY, Azimi PH, Weisman LE, Philips JB, Regan J, Clark P et al. Antibiotic susceptibility profiles for group B streptococci isolated from neonates, 1995-1998. *Clin Infect Dis* 2000;31:76-79.
- Matsubara K, Nishiyama Y, Katayama K et al. Change of antimicrobial susceptibility of group B streptococci over 15 years in Japan. *J Antimicrob Chemother* 2001;48:579-582.
- Mouy D, Cavallo J, Leclercq R, Fabre R. Antibiotic susceptibility and mechanisms of erythromycin resistance in clinical isolates of *Streptococcus agalactiae*: Franch multicenter study. *Antimicrob Agents Chemother* 2001;45:2400-2402.
- Murdoch D, Reller B. Antimicrobial susceptibilities of group B streptococci isolated from patients with invasive disease: 10 years perspective study. *Antimicrob Agents Chemother* 2001;45: 3623-3624.
- Nakae M, Kaneko Y, Mitsuhashi S. Drug resistance in *Streptococcus pyogenes* isolated in Japan. *Antimicrob Agents Chemother* 1977;12:427-428.
- National Committee for Clinical Laboratory Standards (1999). Performance standards for antimicrobial susceptibility testing. Ninth informational supplement, vol 19, N 1, M100-S9. NCCLS, Wayne, PA.
- Orden B, Perez-Trallero E, Montes E, Martinez R. Erythromycin resistance of *Streptococcus pyogenes* in Madrid. *Pediatr Infect Dis J* 1988;17:470-473.
- Orrling A, Stjernquist-Desalnik A, Schalen C, Kamme C. Treatment failure in streptococcal pharyngotonsillitis. An attempt to identify penicillin tolerant *Streptococcus pyogenes*. *Scand J Infect Dis* 1996;28:143-147.
- Palavecino E, Riedel I, Berrios X, Bajaksouzian S, Johnson D, Kaplan E, et al. Prevalence and mechanisms of macrolide resistance in *Streptococcus pyogenes* in Santiago, Chile. *Antimicrob Agents Chemother* 2001;45:339-341.
- Seppala H, Klaukka T, Lehtonen R, Nenonen E. Outpatient use of erythromycin: link to increased erythromycin resistance in group A streptococci. *Clin Infect Dis* 1995;21:1378-1385.
- Skold O. Sulfonamide resistance: mechanisms and trends. *Drug Resistance Updates* 2000;3:155-160.
- Tzelepi E, Kouppari G, Mavroidi A, Zaphiropoulou A, Tzouveleka L. Erythromycin resistance amongst group A beta-haemolytic streptococci isolated in a paediatric hospital in Athens, Greece. *J Antimicrob Chemother* 1999;43:745-746.
- Weiss I, Gorodnetzky Z, Korenman Z, Yagupsky P. Serotyping and susceptibility to macrolides and other antimicrobial drugs of *Streptococcus pyogenes* isolated from patients with invasive diseases in southern Israel. *Eur J Clin Microbiol Infect Dis* 1997;16:20-23.
- York M, Gibbs L, Perdreau-Remington F, et al. Characterization of antimicrobial resistance in *Streptococcus pyogenes* isolates from the San Francisco Bay Area of Northern California. *J Clin Microbiol* 1999;37:1727-1731.

ISOLATION OF LEGIONELLAE FROM HOTEL WATER SUPPLIES IN SOFIA

E. Tsvetkova¹, I. Tomova², B. Halova³, A. Tomov¹, V. Kassovsky¹, A. Milanova¹

1. Department of Microbiology, Military Medical Academy, Sofia, Bulgaria

2. Department of Microbiology, National Center of Infectious and Parasitic Diseases, Sofia, Bulgaria

3. Capital Hygiene Epidemiological Inspection, Sofia, Bulgaria

SUMMARY

Investigations for the presence of legionella bacteria in hotel water supplies in Sofia were performed. For this purpose one hundred and twenty five samples of potable water were collected from eleven hotels. A total of eighty-seven strains were isolated and determined as *Legionella* spp. using culture, biochemical, immunofluorescent (DFA) and gas-liquid chromatography methods. The legionella concentration ranged between 10^1 - 10^4 CFU/L. By the DFA technique the isolates were identified as *Legionella pneumophila*: Sg 1-2 strains, Sg 2- 5 strains, Sg3- 20 strains, Sg 4- 5 strains, Sg6-52 strains, Sg 8-2 strains, Sg 14- 1 strain. Twenty water samples from one of the hotels were detected for amoebae too. After cultivation and microscopic examinations eight strains of small free-living amoebae (FLA) were isolated. At the same hotel we traced out the water quality before and after decontamination by hyperchlorination. It was established that this procedure led to sharp decrease of the bacterial number or absence of culturable legionellae but after one-two months they appeared again. These data suggest that it is necessary to organize regular monitoring of hotel water supplies in Sofia for the presence and the concentration of legionellae.

Key words: *Legionella pneumophila*, free-living amoebae, potable water, hotels

INTRODUCTION

Monitoring for the presence of a pathogen in the environment was initially implemented for the determination of potable water quality with bacterial counts. The monitoring for the presence of fecal contamination in the potable water has been one of the major achievements of public health programs. In recent years, it was found that other pathogens are also present in potable water supplies and that some of them are responsible for severe and even life threatening infections. Legionellae are among these bacteria. Legionellae first commanded attention in 1976-77, when *L. pneumophila* was identified as etiopathological agent in a massive outbreak of pneumonia in a hotel in Philadelphia [1,2]. The intensive survey on the problems of legionellosis found out that inhalation of contaminated with legionellae aerosols is the basic mechanism for infection [3,4,5]. The role of protozoa, especially free-living amoebae (FLA) for the persistence and multiplication of legionellae in the hostile aquatic environment has been confirmed by many investigators [6,7,8,9,10]. In this respect hot and cold water supplies are of a great health importance. There have

been reported many epidemic and also sporadic cases of legionellosis (Legionnaires' disease, Pontiac fever) associated with hotels, leisure complexes and cruise ships [1,10,11,12,13,14] which designate the significance of this problem for tourists business.

The aim of the present study is to shed a light on the presence of legionella bacteria in the plumbing systems of big hotels in Sofia.

MATERIALS AND METHODS

Hotels and water samples. One hundred and twenty five samples of potable water were collected from eleven hotels of residence in Sofia. The frequency of examination for ten hotels was one to ten samples and in one hotel (X) - five samples monthly for one year. For each sample an amount of 200 ml water was collected in a sterile glass container mostly from showerheads and also from taps and tanks. Processing for isolation of legionellae. An amount of 200mL from the sample was centrifuged at 5000 rpm for 30 min. The supernatant was removed, leaving approximately 1% of the original volume in which the sediment was resuspended. A volume of 0.1 ml from the suspended deposit was seeded on Petri dish with Buffered Charcoal Yeast Extract agar- BCYE agar (GIBCO) with PAV selective supplements. spread with a glass spreading rod and after that a second plate with BCYE agar was inoculated with the same glass spreading rod. For 25 samples we used additional 2 plates with BCYE agar (OXOID) and MWY selective supplements which were inoculated by the same manner. All seedings were incubated at 37°C in a humidified atmosphere containing 2.5% CO₂ for 14 days and were examined daily from the third day. Colonies that morphologically resembled legionellae were liable to further identification using culture methods, DFA technique (using antisera for 37 species) and gas liquid chromatography analysis of the fatty acid profile. Processing for isolation of FLA, 0.05mL from the bottom of the glass container were inoculated in the center of a plate with non-nutrient agar (NNA) seeded in advance with inactivated (at 56°C) *Emerobacter aerogenes*. Cultivation was carried out at 25°C for 7 days and regular observation on stereo microscope. FLA containing pieces NNA were transfer on bi-phasic media (NNA, Page's amoeba saline and inactivated *E. aerogenes*) for further observation by phase contrast microscopy.

RESULTS

A total of 125 water samples from 11 hotels in Sofia were investigated for the presence of legionellae. From them 41 samples were positive [tabl.1]. The number of legionella colonies on a plate varied from 1 to 42 colonies on BCYE agar (GIBCO) and from 1 to 120 colonies (or more in some cases- data not shown) on BCYE agar (OXOID) [fig. 1]. The concentration of legionellae in the water samples was mostly between 10^2 - 10^4 cfu/L. From the positive samples a total of 87 strains was isolated and underwent further identification. They showed typical for legionellae culture (morphology of the colonies, absence of growth on 5% blood agar) and biochemical characteristics, as well as production of brown pigment on FG agar.

Species and serogroup identification was carried out by the DFA test. All isolates stained very well with work dilution of the antisera, demonstrating bright yellow-green fluorescence of the periphery and relatively darker center of the cells. All they were rod shaped with length polymorphism in some cases. We found out that the isolates were from seven serogroups of *L. pneumophila* as follows: L.p. Sg6- 52 strains, L.p. Sg3- 20, L.p. Sg2- 5, L.p. Sg4- 5, L.p. Sg1- 2, L.p. Sg8-2/L/J.Sg14-1 [tabl.2].

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ABBREVIATIONS USED IN THIS PAPER:

FLA - free living amoebae

CORRESPONDING AUTHOR:

I. Tomova

Dept. of Microbiology, NCIPD

26, Yanko Sakazov Blvd.

1504 Sofia, Bulgaria

itomova@excite.com

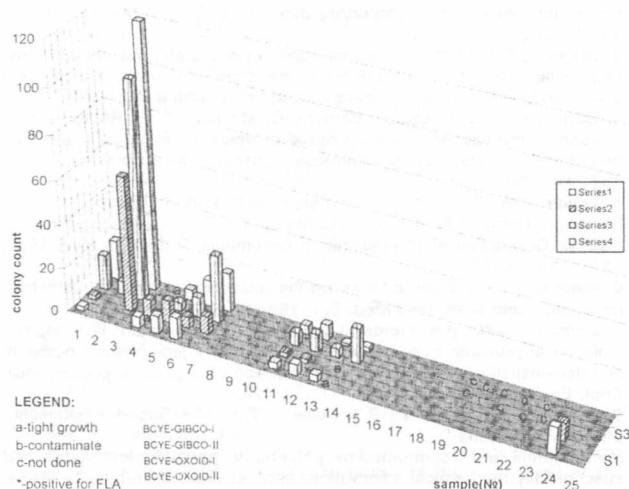


Figure 1.

Presence of *Legionellae* and FLA in water samples from hotel "X"

The gas-liquid chromatography of the isolated strains (compared to the corresponding referent strains) revealed predomination of branched-chain fatty acids with big amount of $iC_{16:0}$ and $iC_{17:0}$ which are typical for genus *Legionella*. [fig.2].

Because of the big number of colonies in sample N3 [fig.1] we performed parallel serotyping of ten colonies, with antisera for thirteen Sg of *L. pneumophila*. Eight colonies react with work dilution of fluorescent antibodies for *L.p.* Sg6 [fig.3] and two colonies - for *L.p.* Sg4 [fig.4].

In the investigation of 20 water samples from hotel "X" we were able to isolate FLA in eight of them [fig.1]. Their identification is forthcoming but some preliminary data (presence of typical acanthopodia) gave grounds to assume that most of them are from *Acanthamoeba* spp, [fig.5]. Six samples positive for FLA were positive for legionellae two.

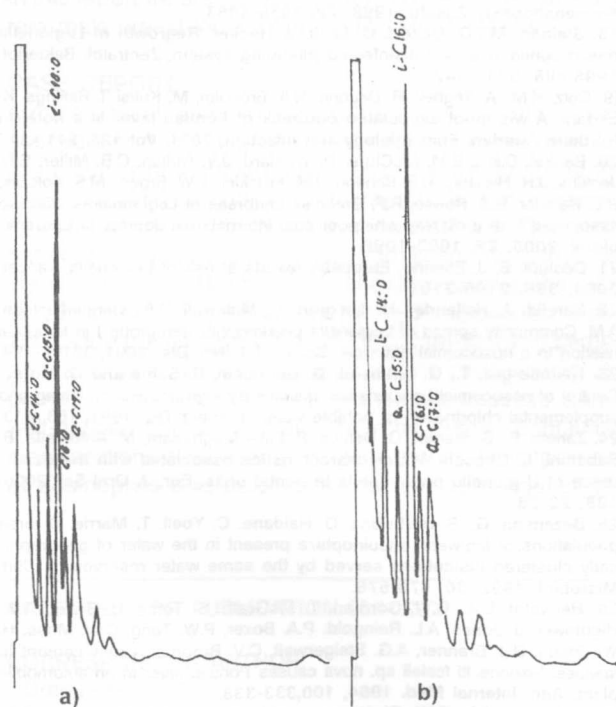


Figure 2.

Gas liquid chromatography of fatty acids of *L. pneumophila* Sg 6:
a) referent strain Chicago-2 (ATCC 33215)
b) strain SH-VIII from water sample N3

In the same hotel ("X") the presence of *Legionella* spp. was traced out monthly for one year and legionellae were found to be common isolates. Because of this a mechanical cleaning with following hyperchlorination of the plumbing system was performed. It was established that this procedure led to sharp decrease of the bacterial number or absence of culturable legionellae but after one-two months they appeared.

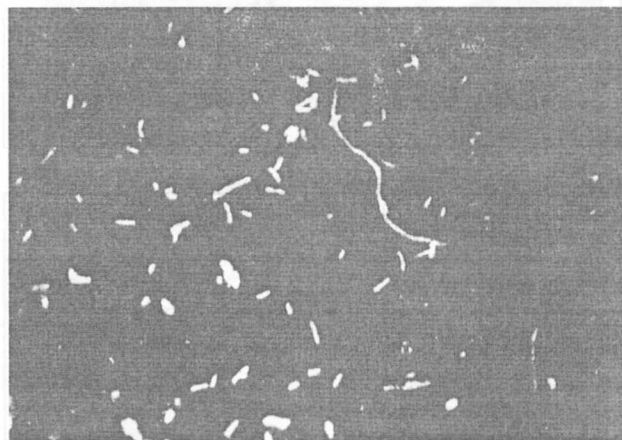


Figure 3.

L. pneumophila Sg 6 strain SH VIII isolated from sample N 3 DFA staining

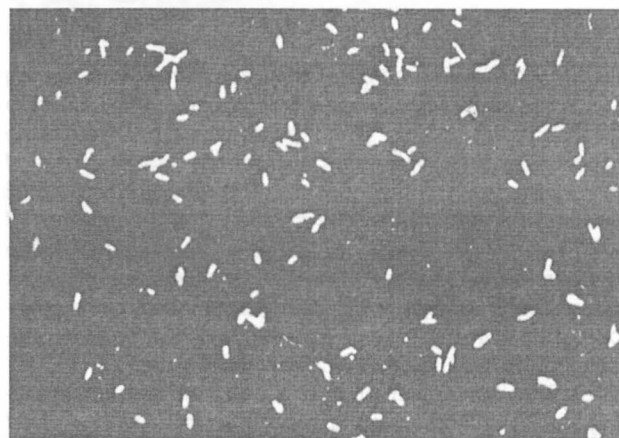


Figure 4.

L. pneumophila Sg 3 strain SH 340d isolated from sample N 3
DFA staining

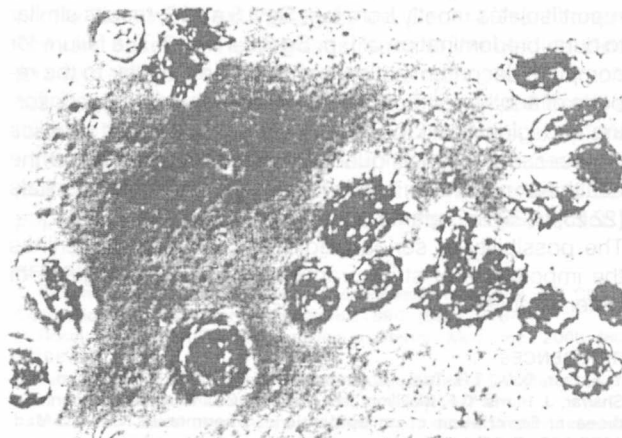


Figure 5.

FLA isolated from sample N 3. Presence of typical acanthopodia.
Phase contrast microscopy

Table 1. Presence of legionellae in water samples from eleven hotels

Hotels	Investigated samples	Positive Samples
I	2	-
II	3	-
III	1	-
IV	5	-
V	5	1
VI	3	-
VII	3	2
VIII	5	2
IX	3	-
X	85	35
XI	10	1
Total	125	41

Table 2. Results from the identification of 87 legionella strains by the dfa method

Species and serogroup	Number of strains
<i>Legionella pneumophila</i> Sg 1	2
<i>Legionella pneumophila</i> Sg 2	5
<i>Legionella pneumophila</i> Sg 3	20
<i>Legionella pneumophila</i> Sg 4	5
<i>Legionella pneumophila</i> Sg 6	52
<i>Legionella pneumophila</i> Sg 8	2
<i>Legionella pneumophila</i> Sg 14	1
Total	87

DISCUSSION

In the investigation of water samples from the plumbing system of eleven hotels in Sofia legionellae were found to be a common isolates. When the investigations were done monthly with five samples per month the isolation of legionellae was more successful. The results from parallel seeding on selective media showed that the number of colonies on BCYE agar (OXOID) was more than twice greater compare to these from the same water sample but grown on BCYE agar (GIBCO). These findings are in accordance with the data of Fallown and Rowbotham concerning the isolation of *L. micdadei* from water [11].

The mixed contamination in one water sample is interesting finding which shows the necessity for detailed study of as much as possible colonies from a plate for complete picture of the *Legionella* population in a plumbing system. The presence of FLA and legionellae in 6 (from 20) samples confirm that FLA are basic hosts of legionella bacteria in the environment, found by another authors [15,16,17]. They report isolates mostly from L.p. Sg 3,5 and 6, results similar to ours- predominance of L.p. Sg 6,3,4 and 2. The failure for complete decontamination is observation similar to the reports of another authors [18]. It shows that regular monitoring for legionellae (and FLA) in man made water supplies is necessary for their quantity control and to reduce the health risk not only in hotels [19,20,21] but also in hospitals [22,23,24,25] and plants [26,27].

The possibility of serious legionella infection underlines the importance of strict practices to maintain hygiene of water supplies.

REFERENCES

- Fraser, D.W., T.R. Tsai, W. Orenstein, W.E. Parkin, J. Beecham, R.G. Sharrar, J. Harris, G.F. Mallison, J.E. McDade, C.C. Shepard. Legionnaires' disease. Description of an epidemic of pneumonia. N.Engl.J.Med. 1977,297, 1189-1197.
- McDade, J.E., C.C. Shepard, D.W. Fraser, T.R. Tsai, M.A. Redus, W.R. Dowdle. Legionnaires' disease. Isolation of a bacterium and demonstra-

tion of its role in other respiratory disease. N. Engl.J. Med. 1977,297, 1197-1203.

- Breiman, R.F. Modes of transmission in epidemic and nonepidemic *Legionella* infection: directions for further study. In: *Legionella- current status and emerging perspectives*. ASM, Washington DC. 1993.30-35.
- Click, T.H., M.B. Gregg, B. Berman, G. Mallison, W.W. Rhodes, Jr., I. Kassanoff. Pontiac fever: An epidemic of unknown etiology in a health department. Clinical and epidemiologic aspects. Am. J. Epidemiol. 1978, 107, 149-160.
- Fraser, D.W., T.R. Tsai, W. Orenstein, W.E. Parkin, J. Beecham, R.G. Sharrar, J. Harris, G.F. Mallison, J.E. McDade, C.C. Shepard. Legionnaires' disease. Description of an epidemic of pneumonia. N. Engl. J. Med. 1977, 297, 1189-1197.
- Rowbotham, T.J. Current views on the relationships between amoebae, legionellae and man. Isr.J.Med. Sci. 1986,22,678-689.
- Barbaree, J.M., B.S. Fields, J.C. Feeley, G.W. Gorman, W.T. Martin. Isolation of protozoa from water associated with a legionellosis outbreak and demonstration of intracellular multiplication of *Legionella pneumophila*. Appl. Environ. Microbiol. 1986, 51,422-424.
- Barker, J., M.R.W. Brown, P.Y. Collier, I. Farrell, P. Gilbert. Relationship between *Legionella pneumophila* and *Acanthamoeba polyphaga*: physiological status and susceptibility to chemical inactivation. Appl. Environ. Microbiol. 1992, 58, 2420-2425.
- Anand, C.M., A.R. Skinner, A. Malic, J.B. Kurt/. Interaction of *Legionella pneumophila* and a free living amoeba (*Acanthamoeba palestinensis*). J. Hyg. Camb. 1983, 91: 167-178.
- Fields, B.S., J.M. Barbaree, G.N. Sanden, W.E. Morrill. Virulence of a *Legionella anisa* strain associated with Pontiac fever: an evaluation using protozoan, cell culture and guinea pig models. Infect. Immun. 1990, 58, 3139-3142.
- Fallon, R.J., T.J. Rowbotham. Microbiological investigations into an outbreak of Pontiac fever due to *Legionella micdadei* associated with use of a whirlpool. J. Clin. Pathol. 1990, 43, 479-483.
- Bornstein, N., D. Marmet, M. Surgot, M. Nowicki, H. Meugnier, J. Fleurette, E. Ageron, F. Grimont, P.A. Grimont, W.L. Tliackier. *Legionella* *grubiana* sp. nova isolated from french spa water. Res. Microbiol. 1989, 140, 541-552.
- Centers for Disease Control and Prevention (CDC). Update: outbreak of Legionnaires disease associated with a cruise ship. MMWR. 1994.43, 574-575.
- Benin, A.L., R.F. Benson, K.E. Arnold, A.E. Fiore, P.G. Cook, L. Keoki Williams, B. Fields, R.E. Besser. An outbreak of travel-associated Legionnaires' disease and Pontiac fever; the need for enhanced surveillance of travel-associated legionellosis in the United States. J.Infect.Dis. 2002,185,237-243.
- Fields, B.S. *Legionella* and protozoa: interaction of a pathogen and its natural host. In: *Legionella: Current status and emerging perspectives*. ASM. Washington, D. C.1993, 129-136.
- Henke, M., K.M. Seidel. Association between *Legionella pneumophila* and amoebae in water. Isr. J. Med. Sci. 1986, 22, 690-695.
- Kuroki, T., S. Sata, S. Yamai, K. Yagita, Y. Katsube, T. Endo. [Occurrence of free-living amoebae and *Legionella* in whirlpool bathes]. Kansenshogaku. Zasshi. 1998, 72, 1056-1063.
- Steinert, M., G. Ockert, C. Luck, J. Hacker. Regrowth of *Legionella pneumophila* in a heat-disinfected plumbing system. Zentralbl. Bakteriologie. 1998,288, 331-342.
- Gotz, H.M., A. Tegnell, B. DeJong, K.A. Broholm, M. Kutisi, I. Railings, K. Ekdahl. A whirlpool associated outbreak of Pontiac fever at a hotel in Northern Sweden. Epidemiology and Infection, 2001. Vol 126, 241-247.
- Benkel, D.H., E.M. McClure, D. Woolard, J.V. Rullan, C.B. Miller, S.R. Jenkins, J.H. Hershey, R.F. Benson, J.M. Pruckler, E.W. Brown, M.S. Kolczak, R.L. Hackler, B.S. Rouse, R.F. Breiman. Outbreak of Legionnaires' disease associated with a display whirlpool spa. International Journal of Epidemiology. 2000, 29, 1092-1098.
- Declut, B., J. Etienne. European resorts at risk of *Legionella*. Lancet, 2000, 356, 2100-2101.
- Darelid, J., Hallander, H., Lotgren, S., Malmvall, B.E., Olinder-Nielsen, A.M. Community spread of *Legionella pneumophila* serogroup 1 in temporal relation to a nosocomial outbreak. Scand. J. Infect. Dis., 2001, 33194-199.
- Heimberger, T., G. Birkhead, D. Bornstein, S. Same and D. Morse. Control of nosocomial Legionnaires' disease through hot water flushing and supplemental chlorination of potable water. J. Infect. Dis. 1991, 163, 413.
- Zanetti, F., S. Stampi, G. Deluca, P. Fateh Moghadam, M. Antonietta, B. Sabatini, L. Checchi. Water characteristics associated with the occurrence of *U-gionella pneumophila* in dental units. Eur. J. Oral Sci. 2000, 108, 22-28.
- Bezanson, G., S. Burbridge, D. Haldane, C. Yoell, T. Marrie. Diverse populations of *Legionella pneumophila* present in the water of geographically clustered institutions served by the same water reservoir. J. Clin. Microbiol. 1992, 30, 570-576.
- Herwaldt, L.A., G.W. Gorman, T. McGrath, S. Toma, B. Brake, A.W. Hightower, J. Jones, A.L. Reingold, P.A. Boxer, P.W. Tang, C.W. Moss, H. Wilkinson, D.J. Brenner, A.G. Steigervalt, C.V. Broome. A new *Legionella* species, *Legionella* *lancefieldii* sp. nova causes Pontiac fever in an automobile plant. Ann. Internal Med. 1984, 100,333-338.
- Solomon, J.A., S.W. Christensen, R.L. Tyndal, C.B. Pliermans, S.B. Gouch. Distribution of *U-gionella pneumophila* in power plant environmental. In: *Legionella: Proceedings of the 2nd International Symposium*. ASM. Washington, DC, 1984. 309-311.

CASE OF DIROFILARIASIS WITH MALE BRAEST LOCALIZATION CAUSED BY DIROFILARIA REPENS

I. Drandarska¹, D. Vuchev², M. Chergova³

1. Laboratory of Immunomorphology, National Center of Infectious and Parasitic Diseases, Sofia, Bulgaria

2. Department of Parasitological and Tropical Medicine, National Center of Infectious and Parasitic Diseases, Sofia, Bulgaria

3. HEI, Department of medical parasitology, Shumen, Bulgaria

SUMMARY

The rare case of dirofilariasis with male breast localization of *Dirofilaria repens* has been described. The parasitological diagnosis has been set up after diagnostically and therapeutically extirpation and histopathological examination of the tissue material.

Key words: dirofilariasis, pathohistology, *Dirofilaria repens*.

INTRODUCTION

Sporadic cases due to *Dirofilaria repens* have been rarely diagnosed in Bulgarian population. The invasions of nematode *Dirofilaria repens* that predominate in South Europe are mainly described (2). The females are usually longer than males and reach a length of 15 cm. The parasite's body is threadlike with diameter 0,5 mm. *Dirofilaria repens* is distinguished with longitudinal ridges cuticle, in contradistinction from *Dirofilaria immitis*, which cuticle is smooth (1). The human is accidental unspecific host and he is not source of infection, because in him the parasite doesn't develop to sexual maturity. The main final host is dog, the cat is possible too, and the transmissible vector is different types of mosquitoes (3). The parasite is localized in the subcutaneous tissues over all parts of the human body where microphilaria is inoculated by the mosquitoes (4). In half of the all clinical cases *Dirofilaria repens* is located in the conjunctive of the eye (5).

In this report we are going to describe briefly one case with rare male breast localization by *Dirofilaria repens*.

CASE REPORT

A male patient, 28-y-old, during the present summer, has noticed a nodule in his left breast near the areola area. On this place is determined a painless, nodular tumor at the size 2 cm. With diagnostic and therapeutic aim an operative removal of the tumor has been carried out. The initial patient's diagnosis was: Obs. Fibroadenoma glandulae mammae sinistri. But histological examination of the extirpated "in toto" tissue material showed unexpected result. The tissue material was fixed in 10 % formalin and carried by the routine histological method. The microscopic appearance of the nodule is a granuloma abscess with a well-defined fibrous wall and necrotic center, containing many sections of parasites *Dirofilaria repens*. Note the mass of eosinophils and polymorphonuclear cells against the

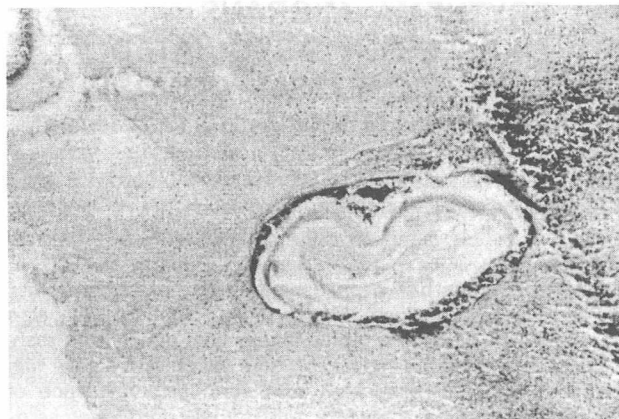


Figure 1.
Dirofilaria repens hematoxylin and eosin stain. Low power view of granuloma with necrotic center, containing of parasite and a lot of polymorphonuclear cells (40 x 2,5)

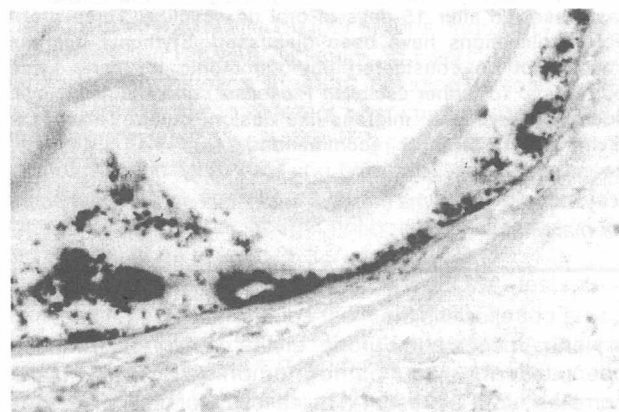


Figure 2.
Dirofilaria repens - high magnification showing the thick layer cuticle, the internal thickening and the longitudinal ridges (10 x 2,5)

worm's cuticle making the morphology rather vague (Fig.1). High magnification of cross section of *Dirofilaria repens* shows the thick layer cuticle, the internal thickening and the longitudinal ridges (Fig.2).

DISCUSSION

The described pathologic-histological finding shows only one possible unusual localization of *Dirofilaria repens*, because the parasite usually is found subcutaneous or subconjunctive. In that case the exact parasitological diagnosis has become possible after histological examination of the tissue material. Usually etiological diagnosis of dirofilariasis is not set initially by the General Practitioners. The rare cases of the diseases reach to the Medical Doctors with different specialties, who are surprised of parasitological diagnosis. It is necessary for General Practitioners to foresee the disease in the differential diagnostic plans. In the last years the cases of dirofilariasis in humans are more often. Probably because of the uncontrollable dog population- the final host and the source of the infection.

REFERENCES

1. Gutierrez, Y. Diagnostic Pathology of Parasitic Infections with Clinical Correlations. Lea & Febiger, Philadelphia. 1990, (532), 316-20.
2. Roccurt, C. La dirofilariose humaine en France. Med. trop., 2000, (60), 3, 308-9.
3. Tarello, W. Un cas de dirofilariose sous-cutanee a *Dirofilaria repens* chez un chat.-Rev. med.vet., 2000, (151), 10, 969-71.
4. Vuchev, D. Clinical observations on dirofilariasis due to *Dirofilaria repens*.-Infactology, XXXV, 1998, 1, 35-6.
5. Vuchev, D., At. Coruev. Cases of dirofilariasis with optimistic localization caused by *Dirofilaria repens*. -Problems of Infectious and Parasitic Diseases, 2001, v. 29, 2, 39- 40.

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CORRESPONDING AUTHOR:

I. Drandarska, MD

Laboratory of Immunomorphology

National Center of Infectious and Parasitic Diseases

26, Yanko Sakazov Blvd.

1504, Sofia, Bulgaria

ERYTHEMA MIGRANS - LIKE LESION ASSOCIATED WITH RICKETTSIA CONORI

R. Komitova¹, M. Murdjeva¹, E. Aleksandrov²

1. Higher Medical Institute, Dept. Infect Diseases, Plovdiv, Bulgaria
2. Military Medical Academy, Sofia, Bulgaria

SUMMARY

Erythema migrans has been described as "pathognomonic" for early Lyme borreliosis. Mediterranean spotted fever, also transmitted by ticks, is a febrile illness with generalized rash, although few cases are "spotless". Here we present a case with Erythema migrans-like lesion with serological evidence of *Rickettsia conori* and transitory anti *Borrelia burgdorferi* seropositivity. A 58-year-old woman was admitted to the hospital in March 2000 with a 2-day history of low-grade fever and rash on her popliteal fossa. An engorged tick was removed from the same place, later identified as *Dermacentor marginatus*. The rash resolved after 15 days of oral doxycycline. Three probable explanations have been discussed. Erythema migrans should not be considered pathognomonic for early Lyme borreliosis. To further establish *Rickettsia conori* as aetiologic agent of Erythema migrans-like lesion culture and PCR technique are strongly recommended.

Key words: *Erythema migrans*, *Rickettsia conori*, *Dermacentor marginatus*

Lyme borreliosis is the most common tick-borne infection in North America and Europe. Erythema migrans (EM) has been described as "pathognomonic" for early Lyme borreliosis and is present in at least 90% of objective cases of Lyme disease. Although once considered classic finding for Lyme disease, EM may be confused with other rashes identical in appearance. These include a hypersensitivity reaction to tick or other insect bites and a recently described rash linked to the bite of an *Amblyomma* tick. This infection likely is caused by another, as yet uncultivable *Borrelia* (2). Mediterranean spotted fever, also transmitted by ticks, is a febrile illness with generalized rash, although few cases are "spotless" (1). The aim of this study is to present a case with Erythema migrans-like lesion with serological evidence of *Rickettsia conori*.

Case report: In March 2000 a 65-year old woman was admitted to the hospital with a 2-day history of low-grade fever and localized rash. Twelve days earlier she removed a tick from his popliteal fossa; subsequently a circular red rash developed at the site of the tick attachment. The tick was kept and later it was identified as *Dermacentor marginatus*, a species found across Europe. At presentation the woman was afebrile and her physical examination was unremarkable apart from a fading erythematous circular rash in his popliteal fossa measuring 25/30 cm. Laboratory tests were within normal range. A diagnosis of

Lyme borreliosis was suggested and treatment with oral doxycycline 100 mg twice daily for 15 days was prescribed. She recovered uneventfully and remained well 6 months later. The acute-phase serum sample, taken on day sixth, was positive by immunoenzyme assay (DAKO) for IgG antibodies to *Borrelia burgdorferi* but the convalescent serum sample taken 20 days later was negative. The patient seroconverted against *Rickettsia conori* by immunofluorescence assay at a titer of 1:80.

Discussion: Our patient presented with fever and an erythematous rash around the site of a tick bite that resembled EM, and she had serological evidence of infection with *R. conorii*. It might be that the case is Mediterranean "spotless" fever with a cutaneous reaction to the bite. However the 6-day incubation period makes this unlikely, although rarely, delayed reaction sometimes occur.

Another explanation would be a dual infection with *Rickettsia conori* and *Borrelia burgdorferi*, which to our knowledge has not been previously described. Yet transitory anti *Borrelia burgdorferi* IgG seropositivity is difficult to explain, as IgM and IgG responses to *Borrelia burgdorferi* not rarely may persist for years even after successful treatment.

Finally, it is probable, that there may be an unidentified rickettsial tick-borne pathogen, causing the clinical picture in our case, that cross reacts with *Rickettsia conorii*. Recently two new rickettsia species, pathogenic for humans, have been described in Europe - *Rickettsia helvetica* and *Rickettsia slovaca* (3,5). Interestingly, they have been reported in patients with no rash, which is the characteristic clinical feature of other rickettsial diseases. The main symptoms of infection with *Rickettsia slovaca* are lymphadenopathy and inoculation skin lesion, varied from crusted scalp lesion (eschar) to EM-like lesion (5). What's more, cases with *Rickettsia slovaca* occur during early spring mainly in March and April and are associated with *Dermacentor marginatus* tick, as was the case with our patient.

Our case highlights difficulties that might come out when evaluating patient with EM-like lesion in early spring. To further establish *Rickettsia conori* as aetiologic agent of Erythema migrans-like lesion culture and PCR technique are strongly recommended.

REFERENCES

1. Brouqui Petal. Spotless Boutonneuse fever. Clin Infect. Dis. 1992; 14:114-6
2. James A et al. *Borrelia lonestari* after a bite by an *Amblyomma americanum* tick. J Infect Dis 2001; 183:1610-1614
3. Nilsson K, Lindquist O, Pahlson C. Association of *Rickettsia helvetica* with chronic perimyocarditis in sudden cardiac death. Lancet 1999;354:1169-7399
4. Raoult D et al. Spotless rickettsiosis caused by *Rickettsia slovaca* and associated with *Dermacentor* ticks. Clin Infect Dis 2002;34:1331-6
5. Raoult D, Brouqui P, Roux V. A new tick-transmitted disease due to *Rickettsia slovaca*. Lancet 1997;350:112-3.

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ABBREVIATIONS USED IN THIS PAPER:

EM - Erythema migrans

CORRESPONDING AUTHOR:

Dr. R. Komitova
Dept. Inf. Diseases; Higher Medical Institute
15A, V. Aprilov Blvd.
4002 Plovdiv, Bulgaria

APPLICATION OF RUBELLA IgG TOTAL ANTIBODY AVIDITY MEASUREMENT IN THE DIAGNOSIS OF RUBELLA INFECTION

Z. Mihneva

Lab. "Measles, mumps, rubella", Department of Virology, NCIPD

SUMMARY

OBJECTIVE: The measurement of total IgG antibody avidity in human sera is a useful technique in confirming the differentiation of an infection. **MATERIALS AND METHODS:** Sera samples (n=79) from rubella immune patients (n=54), classified in four groups: I (group) - including cases (n=12) with clinically and serologically confirmed primary infection; II (group) - including persons (n=25) with MMR vaccine in different post-immunization period; III (group) - including women during pregnancy and women in vitro inseminate (n=15) and IV (group) - including cases (n=2) with rubella embryopathy (CRI) were investigated. ELISA test (commercial kit, firm Bio-Rad) - Platelia Rubella IgG was used. 8 molar urea in wash buffer as a protein denaturing agent was used too. **RESULTS:** Low avidity - antibody index (AI) below 30% (cut-off value) in a range between 10.80% to 25.54% was proved among cases with primary infection, immunized persons in the first three months following vaccination and in one of the rubella embryopathy cases. High avidity - antibody index (AI) above 30% was proved in convalescent sera (previous infection and immunization) - 68.90% (mean value). **CONCLUSION:** The priorities of the IgG total antibody avidity measurement test as good diagnostic marker were discussed.

Key words: IgG total antibody - avidity measurement, urea index (ratio), primary infection, reinfection, rubella embryopathy

Primary rubella infection is usually diagnosed by the detection of specific IgM antibodies in addition to the clinical information. When the blood sample is taken during the first one to three weeks after onset of symptoms the result must be confirmed in paired sera by routine hemagglutination inhibition test (HIT), radial hemolysis test (RHT) and enzyme-immunoassay (EIA) test for detection of specific IgG antibodies.

Sometimes the diagnosis may be difficult to interpret by these sensitive methods, for example in cases when a specific IgM antibodies can persist longer than 8 to 12 weeks and it is difficult to decide if a low positive IgM results is due to recent rubella infection or is caused by rubella reinfection. The last is observed following previous vaccination and is mostly detected following rubella contact.

The HI antibodies rise and often reacted at very high titer with or without presence of specific IgM antibodies.

The avidity or affinity of IgG is initially low after primary antigenic challenge, and it matures slowly with weeks and months (3, 9). The conventional affinity assays of basic immunology for example the globulin precipitation tests are not suitable for the measurement of the polyclonal an-

tibody responses. Decade before, novel techniques have been developed for the serodiagnosis of rubella infection which measure the avidity of specific IgG. These tests are based on the finding that the avidity of antibodies for their antigenic target increase with time, reflecting maturation of the immune response. In the quantitative test, urea and another chemical substances (diethylamine, thiocyanate, guanidine-hydrochloride) is used as a protein denaturing agent (7) to elute IgG from immobilised rubella virus antigen. This results in almost complete dissociation of low-avidity IgG in conditions where high-avidity IgG antibodies mostly remain antigen-bound (6).

Here we examine the ability of the IgG avidity assay, under routine diagnostic laboratory conditions by using a collection of sera previously examined for different evidence of rubella immunity status.

MATERIALS AND METHODS

1. Materials: Sera samples (n=79) from rubella immune patients (n=54), classified in four groups: I (group) - including cases (n=12) with clinically and serologically confirmed primary infection; II (group), including persons (n=25) with MMR vaccine in different post-immunization period; III (group) - including women during pregnancy and women in vitro inseminate (n=15) and IV (group) - including cases (n=2) with rubella embryopathy (CRI) were investigated. The concentration of specific IgG antibodies in the urea studied sera must be at least 25 IU/ml.

2. Methods: standard HI test, indirect ELISA tests, Bio-Rad, (Platelia Rubella IgG - Lot.Ch.B.:1G0073 and Platelia Rubella IgM - Lot.Ch.B.:1H0064, commercial kits) were used. 8 molar urea in wash buffer as a protein denaturing agent was used too. Each test included controls for low and high avidity antibodies (pool from acute and convalescent sera). In case of equivocal result, one more serum was tested.

RESULTS AND DISCUSSION

The maturation of specific IgG antibodies after clinically and laboratory confirmed rubella infection was followed in 34 sera from 12 patients (the I-st group). Sera samples were taken in the first to six and above months after onset of symptoms. Initially, all sera were tested with standard HI test, indirect ELISA IgG and IgM tests for presence of antibody activity. Then a standard ELISA IgG test by urea-index (ratio) procedure was applied. An avidity index is calculated in per cent for each specimen as follows: optical density (OD)-assay with urea / optical density (OD)-assay without urea x 100. The results are shown in Table 1.

After the first week and up to the fourth week following rubella the IgG ratios increased steadily and were in range 5% and 36%.

The development of IgG ratios in 31 sera from 25 vaccinees (the II-nd group) was also studied. In 11 sera from 7 persons, age 1 - 2 years we have observed post-vaccination rubella infection (the sera were taken from 17 days to one month) with IgG ratios below 30% in range between 18.50% and 27.10%. In all seven sera IgM antibodies were also positive. In 20 sera taken between 8 months and 10 years post-vaccination the ratios increased to 47% - 77%.

The investigated sera from the III-rd group show IgG ratios in range between 32% and 79%. Three of them had serological data probably for re-infection with very high HI titer above 1:512 but no evidence for IgM antibodies. It worth to remark that antibody-index of the present IgG antibodies (OD of the sample before urea treatment was 2.199) in one serum was only 22% (28 years after rubella infection).

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ABBREVIATIONS USED IN THIS PAPER:

HIT - hemagglutination inhibition test, ELISA test - enzyme-linked immunoassay test, MMR vaccine - measles mumps rubella vaccine CRI - congenital rubella infection, AI - antibody index

CORRESPONDING AUTHOR:

Z. Mihneva MD, PhD
NCIPD, Department of Virology
26, Yanko Sakasov Blvd
1504 Sofia, Bulgaria

Table 1. Serological investigation and development of IgG antibody-avidity index in 12 cases with recent primary rubella infection

Patient No	Sera	Onset of symptoms (day)	HI test	ELISA IgM test	ELISA IgG test IU/ml	Ratio (AI) %
1	I	3	256	+	< 15	-
	II	15	1024	+	60	18.50
	III	240	64	-	200	73.30
2	I	2	16	+	< 15	-
	II	12	64	+	120	25
	III	210	16	-	190	51
3	I	2	< 8	-	< 15	-
	II	10	256	+	90	26
	III	210	64	-	150	62
4	I	1	< 8	-	< 15	-
	II	7	256	+	130	24.30
	III	180	32	-	230	56
5	I	3	32	+	< 15	-
	II	30	128	-	160	24
	III	170	128	-	290	64
6	I	5	1024	+	200	5
	II	27	1024	+	205	32
	III	210	128	-	216	67
7	I	4	256	+	80	15
	II	30	256	+	105	25
	III	240	128	-	250	65
8	I	8	128	+	178	14
	II	29	128	+	170	34
	III	210	32	-	150	64
9	I	9	512	+	140	25
	II	20	256	+	145	30
	III	180	64	-	175	57
10	I	17	128	+	108	21
	II	36	256	+	178	36
	III	240	128	-	252	59
11	I	32	32	+	187	31
	II	200	16	-	230	88
12	I	2	8	+	<15	-
	II	23	1024	+	55	35.50

Our data are in accordance with Hedman et al. (6) and Enders et al. (4) who have discussed this diagnostical method.

We have studied only two cases (*the IV-th group*) with rubella embryopathy (CRI) whose mother had been infected in early pregnancy. Information on the maturation of avidity of specific IgG after intrauterine infection is not available, however. Low avidity of specific IgG antibodies (ratios - 5%, 15%, 16%, 25% and 29.50%) was demonstrated in consecutive sera (20 days, 6 months, one year, 1 1/2 years and two years of age) from one of these infant. All sera with exception of the last one (2 years after birth) gave positive Ig M values. The present HI rubella antibodies in birth's sera were: baby's - 1:128 and mother's - 1:256. The next sera from the child show results: 1:32/64, 1:32/64, 1:64 and 1:32 as well as a specific IgG - 350 IU/ml, 180 IU/ml, 180 IU/ml, 200 IU/ml and 150 IU/ml. The mother's IgG antibody-

avidity was 61.50% at birth. The child has emphatic clinical manifestations: low birth weight, hepatosplenomegaly, microcephaly, rubelliform rash, rubella pneumonitis, inguinal hernia and mental retardation.

The second child had a thrombocytopenic purpura (Thr, 32000-38000) only as a clinical manifestation after delivery. The present HI rubella antibodies at birth were: baby's serum-1:128, mother's serum 1:128; specific IgG of baby's serum were 109 IU/ml and spesific IgM - positive. Spesific IgG urea-ratio was 58%. The mother's antibody-avidity was 56.80% at birth.

The two mothers have been had a rash in the first trimester of their pregnancy, estimated as allergical. We have studied the other four sera (IgM antibodies negative) from children at birth or shortly after delivery (14, 25 and 28 days and 6 months) as controls. They have also maternal antibodies and the urea-ratios were 15%, 11%, 48.30% and 75.40% respectively. Our data are in accordance with Thomas et al. (8) who have studied the persistence of specific IgG and IgM antibodies and immuno-pathological problems in CRI. Maturation of the immune response is abnormally slow in congenital infection with regard to both isotype switching and the development of high-avidity IgG (8).

A difference in avidity in sera from mother and child at, or shortly after delivery would be diagnostic of an early infection in utero. These results may also be of relevance in immunopathology observed in CRI. The last may result from the formation in certain tissues of antigen-antibody complexes containing low avidity antibody that are not readily cleared by the immune system (2, 8). Rubella spesific immune complexes, not observed in sera from cases of remote rubella, have been demonstred in sera from CRI cases, although the avidity of the specific IgG antibodies was not ascertained (1, 8).

In conclusion, it is necessary a large number investigations with sera in different stage of rubella infection. The results show that the IgG total antibody avidity measurement have priorities as useful marker in the diagnosis of rubella infection.

REFERENCES

1. Coyle, P., J. Wolinsky, E. Buimovici-Klein, R. Moucha, L. Cooper Rubella-specific immune complexes after congenital infection and vaccination. *Infect. Immun.*, 36, 1982, 498-503
2. Devey, M. The biological and pathological significance of antibody affinity. In French MAH (ed), Reeves WG (series ed): "Immunoglobulins in Health and Disease", Immunology and medicine series, Lancaster: MTP Press, 1988
3. Eisen, H., Q. Siskind Variations of affinities of antibodies during the immune response. *Biochemistry*, 3 1964, 996-1008
4. Enders, G., F. Knotek Rubella IgG Total Antibody Avidity and IgG Subclass-specific Antibody Avidity Assay and their Role in Differentiation between Primary Rubella and Rubella Reinfection. *Infection*, 17, 1989, 4, 218-226
5. Hedman, K., I. Seppala Resent Rubella Virus Infection Indicated by a Low Avidity of Specific IgG
6. Hedman, K., S. Rousseau Measurement of Avidity of Specific IgG for Verification of Recent Primary Rubella
7. Kamoun, P. Denaturation of globular proteins by urea: Break-down of hydgen or hydrophobic bonds? *Trends in Biochemical Sciences*, 13, 1988, 424-425
8. Thomas, H., P. Morgan-Capner, J. Craddock-Watson, G. Enders, J. Best, S. O'Shea Slow Maturation of IgG1 Avidity and Persistence of Specific IgM in Congenital Rubella: Implications for Diagnosis and Immunopathology. *J. Med. Virology*, 41, 1993, 196-200
9. Webrin, T., Y. Kim, Q. Siskind Studies in the control of antibody synthesis. III. Changes in the heterogeneity of antibody affinity during the course of the immune response. *Immunology*, 24 1973, 477-492

CLINICAL EXPERIENCE IN THE TREATMENT OF FLU PATIENTS WITH OSELTAMIVIR DURING THE INTERPANDEMIC PERIOD

V. Markovski, Z. Milenkovic, M. Dimzova, B. Tosevski

Clinic for Infectious Diseases, Clinical Center, Skopje, Republic of Macedonia

SUMMARY

The influenza epidemics are very important health and economics issues. That explain why defining the methods for proper treatment and prophylaxis of influenza has become so important over the past few years. The study included people with present influenza symptoms throughout the last influenza season / December/2001-April/2002 / confirmed by IIF, ELISA-IgM, isolation of the virus and with clear clinical symptoms. The goal of the study was on the basis randomized, double-blind study to evaluate the therapeutic effects of oseltamivir concerning the duration of the illness and symptoms. The study shows that the therapy with oseltamivir resulted in reduction of symptoms in febrile as well as in non febrile patients, smaller administration of symptomatic therapy, less complications as a secondary bacterial infections, lower administration of antibiotics. The drug oseltamivir is very well tolerated and only 5,9% of the treated patients noted intolerance with vomiting. From the present study the authors conclude that treatment of influenza with oseltamivir causes significant shortening of illness period as well as of clinical expression and symptoms of influenza.

Key words: influenza epidemics, oseltamivir, influenza virus A and B

In regions having a temperate climate, influenza epidemics occur annually. They occur predominately during the winter months, and are usually associated with increased mortality among the higher risk population (namely, the elderly and persons with chronic diseases). Likewise, the general population's morbidity rate also increases, very dramatically in some years. The effect of influenza epidemics is here attested by significant work or school absenteeism, and decreased level of worker productivity. These effects are prolonged, in many cases, as symptoms of the illness can last for up to seven days. For illustration, in 1997 influenza and pneumonia were ranked sixth among the leading causes of mortality in USA. Over the past 25 years, an average of 20,000 Americans a year have died from these disease. It is estimated that the annual economic loss due to influenza stands at \$3-5 billion. These factors explain why defining the methods for proper treatment and prophylaxis of influenza has become so important over the past few years. While vaccination remains the standard form of prophylaxis, an increasing number of studies have begun to investigate the prophylactic potential of certain chemotherapeutics. Despite everything the illness continues to appear, especially among the children and young adults, the groups with the highest risk for morbidity. Antiviral agents remain a complementary treatment, and in no way substitute for extensive and efficient vaccine programs. Antiviral agents administered early during the infection can reduce the duration of the illness and lessen the degree of symptom expression (intensiveness, severity), and with all that also to reduce the general impact of influenza.

The first antiviral agents used to treat influenza infections were amantadine and rimantadine, but their use was limited only to a few

regions worldwide. With time, questions were raised regarding their true role in the treatment of influenza, their side effects, and most importantly, their potential in selection of resistant viral strains.

Previously, a consensus had been reached regarding the need for prompt treatment of influenza. The rationale was that even the most uncomplicated bout of influenza is a long-lasting illness which significantly limits daily activities. However, this consensus changed after several additional considerations were taken into account. The risk of side effects, especially during the amantadine therapy, the possibility for appearance of resistant types, and the need for reassurance that the illness treated is in fact caused by the Influenza A virus. The efficiency of these drugs is documented only against influenza virus type A. Therefore, their administration becomes insufficient in years when mixed influenza epidemics occur. Thus the need for an efficient antiviral agent with broader activity spectra and with better characteristics regarding the safety profile. First in the new class of antiviral agents which actively inhibit the influenza-neuraminidase, the enzyme which is essential for viral replication in vitro, was zanamavir. Its efficiency, proven in vitro on experiment model with animals and young adults, covers both influenza A and B infections. The clinical effectiveness of locally administered (through oral inhalation) zanamavir against natural influenza infection in humans has been documented in large placebo-controlled studies. But the very use of the inhaler for its application means that patients need to be taught how to use an inhaler. In patients with asthma or chronic obstructive pulmonary disease (COPD), Zanamavir can induce bronchospasm and reduced pulmonary function- such patients, therefore, should avoid using zanamavir. In addition, zanamavir has certain, albeit rare, side effects; most prominent are irritation of the nose and throat, coughing and headaches. Unlike zanamavir, the newer neuraminidase inhibitor, oseltamivir, is administered orally (75 mg tablets) and is a "prodrug". This means it must be converted, with the activity of hepatic esterase, into its active form, oseltamivir carboxylate. Oseltamivir carboxylate is not further metabolized and as such is completely eliminated through renal excretion. Its only registered side effects are nausea and vomiting, which can be lessened by taking the drug together with food. Compared with other antiviral agents, this drug has a very small potential for selection of resistant strains/variants of the virus. With all of these considerations in mind, we sought to define the clinical efficiency and safety of oseltamivir on the general population, with our own material. Furthermore, this study involved persons at high risk for developing complications associated with influenza infection. This randomized, double-blind study evaluated the therapeutic effects concerning the duration of the illness and symptoms relief in function of defining the indicators for the severity of the illness.

MATERIAL AND METHODS

This study included people aged 18 or older with present influenza symptoms, of a duration of 48 hours or less throughout the last influenza season (December/2001-April/2002). Primarily, these were patients treated during the last two months of the season. Influenza symptoms are defined by fever, chills and the presence of two or more of the following: myalgia, headache, cough or sore throat. The probability of proving influenza infection in the patients that were included was high, considering the fact that this study was undertaken during the period in which there was high percent of 85% of serologically proven (by the IIF method) influenza cases. In addition, this study also included patients over 65 years of age, as well as patients with chronic diseases (cardiovascular, respiratory, endocrinal, diseases of the metabolism, excluding those patients having an unstable disease); this category of patients is marked as a high risk group for developing complications or more severe or prolonged forms of the illness. In the study, oseltamivir was administered in an oral dosage of 2x75 mg/daily, for a period of 5 days. The study was completed with successive gathering of the planned 50 (20 with, and 30 without oseltamivir) patients, based on the principle of coincidence and according to their daily arrival at the Clinic for Infectious Diseases in Skopje. We randomly analyzed the patients in a 1:1/1:2 relation to those patients who did not

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ABBREVIATIONS USED IN THIS PAPER:

COPD - chronic obstructive pulmonary disease

CORRESPONDING AUTHOR:

V. Markovski

Clinic for Infectious Diseases, Clinical Center
Skopje, Republic of Macedonia

take Oseltamivir. All of them were outpatients and they were given a questionnaire in which, on a twice daily basis, morning and evening, over the next ten days, they marked their temperature and the severity of symptoms (clinical manifestation). The clinical manifestation of symptoms we marked on a 6 degree scale, on which 0 corresponds with a total absence of symptoms, and 5 with the highest intensity of presence of symptoms. On the same questionnaire the patients noted any sleeping disorders, their capability for completing daily normal activities, and their overall health condition. The further analysis of the study included only patients with confirmed influenza infection (IIF, ELISA-IgM, isolation of the virus from samples taken from the upper respiratory tract, on the very first day the patients arrived at the Clinic). Of 41 patients (82%, 41/50), it was found later that 17 (85%, 17/20) had taken oseltamivir, and 24 (80%, 24/30) had not. Our primary clinical goal was to determine the duration of the illness until any lessening could be noted in clinically significant symptoms, defined with the absence of chills, fever of lower than 37.5°C, and a mark "0" (absence) or "1" (mild) for any other major symptoms (for example headache, myalgia, sore throat or cough) remaining during the next 24 hours. The time needed for relief of symptoms was determined on the basis on half day from starting of the treatment (day one), with the morning parameters from day one of the treatment that correspond with "0" day. The time period from the morning measurements until the evening ones represents half day. In addition, we sought to determine the middle score of symptoms, sleep disruption, duration of symptoms before normalization of daily activities, as well as the usage of acetaminophen and cough-relieving drugs. The degree of clinical expression of the five symptoms (chills, headache, myalgia, cough, and sore throat) was summarized for each patient by calculating the middle score of symptoms, used as a measure for the overall severity of clinical symptoms. The middle score of the symptoms is determined with varians analysis.

RESULTS AND DISCUSSION

In the group of patients treated with oseltamivir, the final point, alleviation of major symptoms, was achieved around the fourth day. In the group of patients who were not treated with oseltamivir, alleviation occurred only on the sixth day. This difference was more obvious in patients who arrived at the Clinic and began their treatment during the first 24 hours of symptoms appearance, (for 2-3 days). The difference was also most notable among those patients who were febrile or belonged to the high risk group at the time when put under therapy. In patients who had their treatment started within 24 hours of symptomatic appearance, therapy with oseltamivir shortened the illness period to 0.5-1 day. Therapy with oseltamivir resulted in a reduction of symptomatic duration in febrile, as well as non febrile patients (at the time of inclusion in the study), though the effects were more evident in the former group. The middle duration of symptoms in the high risk group of patients was longer compared with the entire treated group (6.56 contrary to 6.0 days), which documents the trend for faster relief of symptoms in patients with high risk. Therapy with oseltamivir also resulted with smaller administration of symptomatic therapy, e.g. the same was associated with reduction of the usage of acetaminophen and antitussives. During the first three days, administration of acetaminophen was decreased for 38%, and for cough-relieving drugs 29%. In this study we also analyzed the incidence of secondary diseases (otitis media, pneumonia, etc), as well as the incidence of hospitalizations. Complications were rare in the group treated with oseltamivir, meaning, a reduction of patients with otitis media as a complication has been achieved from 12.5% (3/24) to 5.9% (1/17) as well as with pneumonia from 20.8% (5/24) to 11.8% (2/17), and generally speaking with all this a dramatic reduction of hospitalizations has been managed, from 29.2% (7/24) to 11.8% (2/17). Administration of antibiotics was significantly lower in the group of patients treated with oseltamivir. Patients treated with oseltamivir tolerated the medicine very well, and only in one patient is noted intolerance with vomiting (5.9%). It took many years for consensus on use of amantadine and rimantadine to be reached in North America. This was partly due to the fact that the studies, especially

those involving treatment, were small and differed in design, making assessment of the precise therapeutic benefit difficult. The development of neuraminidase inhibitors provides an alternative antiviral therapy for the management of influenza infection. This agents have convincing efficacy against influenza caused by different types of influenza viruses, as is shown in many studies. In contrast to older antiviral drugs, studies with the neuraminidase inhibitors have used the same well-defined end point and similar design. The benefits of them appear to be greatest where the illness is most severe, as would be expected from the natural history of influenza infection, and when treatment is given early. There are also benefits on patient quality of life and functioning. The results of all studies with oseltamivir indicate that it is well tolerated, with a reported side-effect profile comparable to placebo. In this study this was evident in both otherwise healthy persons and in those at high risk of complications. The fact that oseltamivir can be used to treat all strains of influenza A and B viruses and that resistance has not been detected during acute therapy, also suggests that it has definite advantages over the older antiviral agents. From the presented results we can conclude that treatment of influenza with oseltamivir causes significant shortening of illness period as well as of clinical expression and symptoms of influenza. Thus, there seems to be a clear role for oseltamivir in the treatment of type A and B influenza infections.

REFERENCES

- Centers for Disease Control and Prevention: Prevention and control of influenza. Part II. Antiviral agents - recommendations of the Advisory Committee on Immunization Practices, MMWR, 43:1-10, 1994.
- Couch, R. B.: Prevention and treatment of influenza, N.Engl. J. Med., 343(24):1778-1787, 2000.
- Couch, R.B.: Prevention and treatment of influenza, N.Engl.J.Med., 343(24):1778-1787, 2000.
- Cox, N.J., Hughes, J.M.: New options for the prevention of influenza, N.Engl.J.Med., 341(18):1387-1388, 1999.
- Feder Jr., H.M.: Zanamivir to prevent influenza, N.Engl.J.Med., 344(7):528-530, 2001.
- Govorkova, E.A., Leneva, I.A., Goloubeva, O.G., Bush, K., Webster, R.G.: Comparison of efficacies of RWJ-270201, Zanamivir, and Oseltamivir against H5N1, H9N2, and other avian influenza viruses, Antimicrob.Agents Chemother., 45(10):2723-2732, 2001.
- Gubareva, L.V., Matrosovich, M.N., Brenner, M.K., Bethell, R.C., Webster, R.G.: Evidence for zanamivir resistance in an immunocompromised child infected with influenza B virus, J.Infect.Dis., 178:1257-62, 1998.
- Gubareva, L.V.: Molecular mechanisms of influenza virus resistance to neuraminidase inhibitors, 1st Europ.Influen.Confer., Malta, 20-23.X., Abstracts, abstr. W4-1, p.14, 2002.
- Hayden, F.G., Osterhaus, A.D.M.E., Treanor, J.J.: Efficacy and safety of the neuraminidase inhibitor zanamivir in the treatment of influenza virus infections, N.Engl.J.Med., 337:874-80, 1997.
- Hayden, F.G., Gubareva, L.V., Monto, A.S., Klein, T.C., Elliot, M.J., Hammond, J.M., Harp, S.J., Ossi, M.J.: Inhaled zanamivir for the prevention of influenza in families, N.Engl.J.Med., 343(18):1282-1289, 2000.
- Hayden, F.G., Gubareva, L.V., Monto, A.S.: Postexposure prophylaxis with inhaled zanamivir was efficacious for household contacts of people with influenza, Evidence Based Med., 6:140, 2001.
- Hayden, F.G.: Perspectives on antivirals for influenza, 1st Europ.Influen.Confer., Malta, 20-23.X., Abstracts, abstr. S4-3, p.18, 2002.
- Matrosovich, M.: A laboratory cell line for testing influenza virus sensitivity to neuraminidase inhibitors, 1st Europ.Influen.Confer., Malta, 20-23.X., Abstracts, abstr. W4-2, p.14, 2002.
- Neuzil, K.M., Reed, G.W., Mitchel, E.F., Simonsen, L., Griffin, M.R.: Impact on influenza on acute cardiopulmonary hospitalizations in pregnant women, Am.J.Epidemiol., 148(11):1094-1102, 1998.
- Nguyen-Van-Tam, J.S.: Zanamivir for influenza: A public health perspective, Brit.Med.J., 319: 655-656, 1999.
- Nikolic, S.: Nove mogucnosti u terapiji i profilaksi influence-inhibitori neuraminidaze, Acta Infect. Yugoslavica 5:161-167, 2000.
- Suzuki, Y., Suzuki, T., Miyamoto, D., Hidary, K., Guo, C., Ito, T. and oth.: Approach to develop new anti-influenza drugs targeted to the function of hemagglutinin and neuraminidase, 1st Europ.Influen.Confer., Malta, 20-23.X., Abstracts, abstr. S4-4, p.15, 2002.
- The MIST Study Group: Randomised trial of efficacy and safety of inhaled zanamivir in the treatment of influenza A and B virus infections, Lancet, 352:1877-81, 1998.
- Wiselka, M.: Influenza: diagnosis, management, and prophylaxis, Brit.Med.J., 308:1341-1345, 1994.
- Zambon, M., Carr, J., Ives, J., Roberts, N.: Influenza virus neuraminidase mutations selected by oseltamivir phosphate in clinical use do not impact immune recognition, 1st Europ.Influen.Confer., Malta, 20-23.X., Abstracts, abstr. P-W4-2, p.62, 2002.

INTRADERMAL TESTS WITH ALLERGENS *S. AUREUS* AND *C. ALBICANS* IN PATIENTS WITH ATOPIC ECZEMA/ DERMATITIS SYNDROME

J. Kochetova², M. Mokronosova², B. Petrunov¹

1. National Center of Infectious and Parasitic Diseases, Sofia, Bulgaria.

2. Mechnikov's Research Institute for Vaccines and Sera, Moscow, Russia.

Atopic eczema/dermatitis syndrome (AEDS) - is a chronic allergic skin disease. There is increasing evidence that infectious agents are important triggering factors in the pathogenesis of AEDS [23].

S. aureus has been studied as possible trigger factor. It is well known fact that the skin of 70-90% patients with AEDS is colonized with *S. aureus* in extremely high density (10^7 colony - forming units/cm²) [3,20]. In contrast only 5-10% of healthy individuals without any skin diseases are carriers *S. aureus* in low density of colonization [2,27]. *S. aureus* is known as the producer of proteins with superantigenic properties: staphylococcal enterotoxin A (SEA), staphylococcal enterotoxin B (SEB), staphylococcal enterotoxin C (SEC), toxic shock syndrome toxin - 1 (TSST-1), staphylococcal exfoliative toxin (SET). At first, they can bind to the MHC II molecules of monocytes and dendritic cells and release a number of proinflammatory molecules such as interleukin-1 (IL-1) and tumor necrosis factor - alpha; at the second, they can stimulate the proliferation of T-cells with V β chain expression [11,15,18]. Moreover staphylococcal enterotoxins are considered as a new group of allergens. Specific IgE-antibodies to SEA, SEB, SEC and TSST-1 have been detected in sera from 57% of patients [4,16,17,26]. Basophils of the patients with AEDS were found to release histamine upon incubation with these toxins [9].

The relationship between the severity of AEDS and the density of *S. aureus* colonization in the skin was revealed. The moderate and severe courses of AEDS are characterized by the high density of staphylococcal colonization. Recently a number of studies have published that the severity of AEDS depends on presence of staphylococcal enterotoxins in the skin secreting by *S. aureus* [1,3].

The lipophilic yeast *Malassezia spp* is not only part of the normal human skin flora but also an opportunistic pathogen. Various recent reports have discussed *Malassezia spp* as a pathogenic factor in AEDS [7,22]. *C. albicans*, unlike *Malassezia spp*, does not normally colonize the skin, but rather the gastrointestinal tract and the nasopharynx. A pathogenic role for allergy of *C. albicans* has for long been proposed, particularly in asthma and rhinitis, and a high rate of positive immediate skin test to *C. albicans* has been observed in atopic patients [24]. Allergic cross reactivity

between *Malassezia spp* and *C. albicans* has been investigated and found to be significant [6,12,14].

The allergenic properties of *Malassezia spp* and *C. albicans* are well known and few recombinant allergens have been obtained and characterized [28]. The allergens have been recognized as mannoproteins. It was shown that predominantly sensitization to *Malassezia spp* has been occurred in patients with head and neck skin lesions [19]. *C. albicans* as allergen, together with other antigens (PPD, tetanic toxin, trichophytin), is as the marker of cell immunity and reveals delayed type hypersensitivity reactions *in vivo* [5,8]. The aim of the study was to investigate immediate and delayed type skin reactivity by means of intradermal testing by allergens from *S. aureus* and *C. albicans* in patients with AEDS.

MATERIALS AND METHODS

Patients and diagnostic criteria. 28 patients from 14 till 40 years (age - $23,9 \pm 4,6$ years) with different severity of AEDS were observed. The following participants were recruited: AEDS was diagnosed according the criteria Hanifin and Rajka [10] and the revised nomenclature for allergy was used [13]. The severity of AEDS was scored according SCORAD system [25]. The severity of AEDS was mild in 16 patients, moderate - in 12, severe - in 7 cases. All patients did not receive antihistaminic, cytostatic and corticosteroid drugs for 2 weeks before testing.

28 healthy nonatopic people in the age of from 15 till 38 years (mean - 18,1 years) made the control group.

The local ethics committee approved the protocol and we obtained written informed consent from all patients.

Intradermal tests with allergens. Allergenic extracts of *S. aureus* and *C. albicans* provided by the National Center of Infectious and Parasitic Diseases (Sofia, Bulgaria) were used. Testing was performed by means of intradermal injections of the 0,05 ml allergen solution in skin of an internal surface of a forearm. The estimation of immediate reaction was carried out after 20 minutes and delayed reactions after 72 hours. The following parameters were taking into account: the size of wheal in 20 minutes and the size of papula in 72 hours. Measuring of the papula/wheals sizes after 20 minutes 24, 48 and 72 hours was provided to estimate the dynamics of the skin reactivity in patients with AEDS.

Statistics: P - value of less than 0,05 was considered to be statistically significant. Chi - squared tests and t - tests were used for categorical and continuous variables, respectively. The statistical package spss 9,0 for Windows was used for the analysis.

RESULTS

Intradermal testing with allergen *S. aureus* and *C. albicans* was provided in 28 patients with AEDS and in 28 non atopic healthy persons. There were not registered side effects during intradermal testing by allergens *S. aureus* and *C. albicans*.

The results of intradermal testing with allergen *S. aureus* with an estimation of the sizes of skin reactions after 20 minutes are presented on fig.1. Apparently the sizes of wheal in the group of patients with AEDS have made on the average 3,9 mm, against 1,8 mm in a control group, i.e. in 2 times more. However there was no statistically significant correlation between these data in both groups ($P=0,5$). At the same time, the wheal after introduction of allergen *S. aureus* was absent in 15 patients with AEDS and in 19 - from group of comparison. Thus immediate reactions to *S. aureus* are revealed in 46,4 % of atopic patients and in 32,1 % of nonatopic healthy people. The severity of AEDS (according to SCORAD) in the patients with positive immediate re-

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CORRESPONDING AUTHOR:

Dr. J. Kochetova
Mechnikov's Research Institute for Vaccines and Sera
Malij Kezennijpereluk 5a
103064 Moscow, Russia

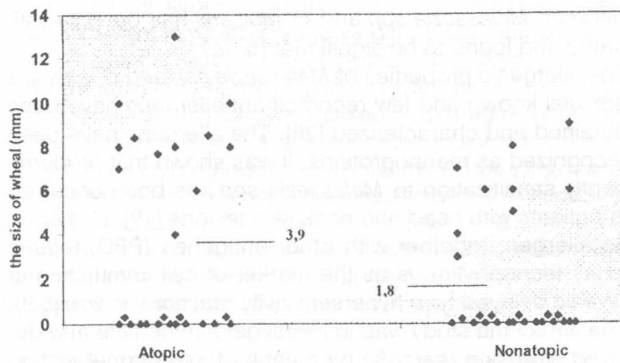


Figure 1.
Immediate hypersensitivity type reactions
at intradermal testing with allergen *S. aureus*
in atopic and nonatopic patients

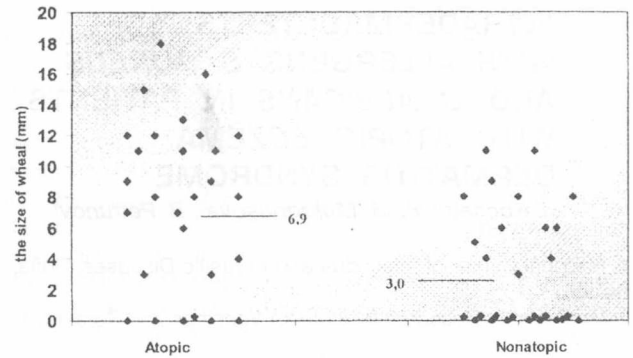


Figure 2.
Immediate hypersensitivity type reactions
at intradermal testing with allergen *C. albicans*
in atopic and nonatopic patients

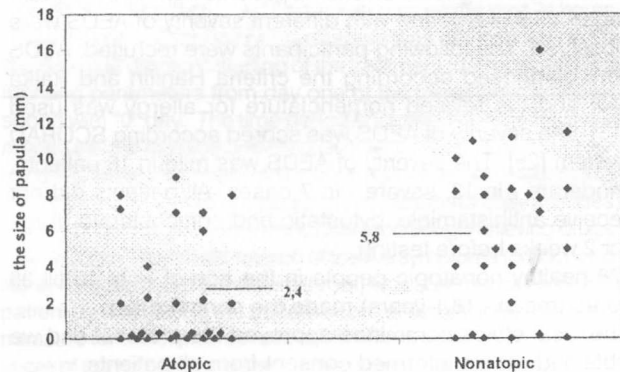


Figure 3.
Delayed hypersensitivity type reaction
at intradermal testing with allergen *C. albicans*
in atopic and nonatopic patients

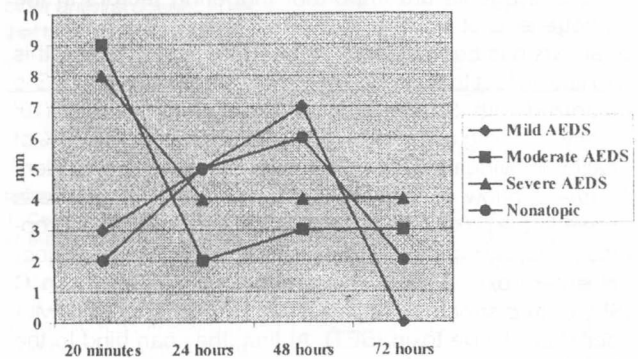


Figure 4.
Dynamics of medians' sizes of wheal/papula
at intradermal testing with allergen *C. albicans*
in patients with various severity of AD

actions to *S. aureus* was regarded as moderate and severe in 8 of 28 (28,6 %) patients.

The comparative analysis of papula's sizes detecting delayed type reactivity after testing *S. aureus* has not revealed significant differences between these parameters in both groups (0,8 mm and 1,0 mm, accordingly).

The results reflecting immediate reactions to allergen *C. albicans* are shown on pic.2. There was absence of any reactions in 7 patients with AEDS against of 15 nonatopic persons after intradermal introduction of allergen *C. albicans*. Frequency of positive immediate reactions to allergen *C. albicans* in group with AEDS has made 75 % against 46,4 % in a control group. The average size of wheal in patients with AEDS has made 6,9 mm and 3,0 mm in nonatopic people. These data differed statistically significant according to *chi*-test, but $P=0,5$.

The results demonstrating delayed reactions to yeasts essentially differed from those in testing with allergen *S. aureus*. Obvious oppression of cellular infiltration in place of intradermal injections of allergen *C. albicans* in patients with AEDS is observed in fig.3. The skin reaction to *C. albicans* in 22 (78,6 %) patients with AEDS either was absent or did not exceed 5 mm against 10 (35,7 %) in nonatopic persons. Accordingly the mean size of papula also significantly differed in 2 times in both groups (in patients with AD - 5,8 mm, in patients from control group - 2,4 mm, $P=0,01$). Absence of delayed type reactions to *C. albicans* was observed in the same 8 patients with moderate and severe AEDS and positive immediate reactions to *S. aureus* that were revealed earlier.

The intradermal testing by allergen *C. albicans* was provided for investigation of the correlation between wheal/papula size and severity of the disease in 28 patients with various severity of AEDS. The estimation of skin reactivity was carried out after with 20 minutes, 24, 48 and 72 hours. Dynamics of medians' sizes of wheal/papula at intradermal testing with allergen *C. albicans* in patients with mild, moderate, severe AEDS and in nonatopic persons is shown on fig. 4. The lines of changes of skin reactions' sizes practically coincide in patients with mild AEDS and nonatopic people. The threefold increase of the wheal's sizes reflecting immediate skin reactivity on the background of decrease of delayed skin reactivity is marked in patients with moderate and severe AEDS.

DISCUSSION

The AEDS is multifactorial disease, in pathogenesis of which both infectious and noninfectious agents are participating. It is necessary to establish individual spectrum of sensitized factors for adequate therapy and elimination measures.

The publications of Leung and coauthors proved that the staphylococcal superantigens influence on the severity of the course of AEDS [16,17,18]. But using of such expensive methods of diagnostics in clinical practice is unacceptable.

In contrast the results of the present and published before papers [21] in studying of immediate and delayed reactions to infectious agents *S. aureus* and *C. albicans* allow using intradermal testing to verifying a sensitization of pa-

tients to infectious allergens and an objective estimation of severity of AEDS. It is obvious that the deepest infringements of immune answer are characteristic in patients with positive immediate reactions to *S. aureus* and *C. albicans*. Positive immediate reactions and the absence of delayed reactions to allergens *S. aureus* and *C. albicans* in a control group testify, most likely, the presence of infection and decrease of cellular immunity [5,8].

The data to be obtained suggest that the changes of skin reactivity to *S. aureus* and *C. albicans*, the increase of the frequency and the sizes of immediate reactions and the decrease of the sizes of delayed reactions are typical features in patients with AEDS. Besides these changes are especially characteristic for severe AEDS.

The use of intradermal test with allergens *S. aureus* and *C. albicans* can serve as the objective marker of the severity of AEDS and prognostic factor of chronization of AEDS.

REFERENCES

1. Akiyama H., Toi Y., Kanzaki H. et al. Prevalence of producers of enterotoxins and toxic shock syndrome toxin-1 among *Staphylococcus aureus* strains isolated from atopic dermatitis lesions.//Arch. Dermatol. Res. - 1996. - Vol. 288 (7). - P.418 - 420.
2. Aly R. Microflora of atopic dermatitis.//Arch.Dermatol. - 1977. - Vol. 113 (6). - P.780-782.
3. Breuer K., Wittmann M., Bosche B., et al. Severe atopic dermatitis is associated with sensitization to staphylococcal enterotoxin B (SEB).//Allergy. - 2000. - Vol. 55. - P.551-555.
4. Bunikowski R., Mielke M., Skarabis H. et al. Prevalence and role of serum IgE antibodies to the *Staphylococcus aureus* - derived superantigens SEA and SEB in children with atopic dermatitis.//J. Allergy Clin. Immunol. - 1999. - Vol. 103. - P.119-124.
5. Cainzos M., Culebras J.M., Lozano F., et al. A study of the delayed hypersensitivity response in healthy people in Spain: Spanish National Tables.- National Surgical Infection Committee of the Association of Spanish Surgeons.// J. Parenter. Enteral. Nutr. - 1993. - Vol. 17(5). - P.85-89.
6. Doekes G., Vanleperenvandijk A.G. Allergens of *Pityrosporum ovale* and *Candida albicans*. Cross reactivity of IgE - binding components.//Allergy. - 1993. - Vol. 48 (6). - P.394 - 400.
7. Duff A.L., Wilson B.B., Nolte H.K. et al. Hypersensitivity to *Pityrosporum orbiculare* in atopic dermatitis.//J. Allergy Clin. Immunol. - 1993. - Vol. 91 (1). - P.245-249.
8. Esch R.E., Buckley C.E.- A novel *Candida albicans* skin test antigen: efficacy and safety in man.//J. Biol. Stand. - 1988. - Vol. 16. - P.33-43.
9. Espersen T., Jarlov J.O. *Staphylococcus aureus* peptidoglycan induced histamine release from basophil human leukocytes in vitro.//Infect. Immun. - 1984. - Vol. 46. - P.710-714.
10. Hanifin J., Rajka G. Diagnostic features of atopic dermatitis//Acta Derm. Venerol. (Stockh). - 1980. - Vol. 114 (Suppl). - P. 44-47.
11. Herman A., Kappler J.W., Marrack P. et al. Superantigens: mechanism of T-cell stimulation and role in immune responses.//Ann. Rev. Immunol. - 1991. - Vol. 9. - P.745-772.
12. Huang X., Johansson S., Zargari A., Nordvall S.L.//Allergen cross reactivity between *Pityrosporum orbiculare* and *Candida albicans*.//Allergy. - 1995. - Vol. 50. -P.648-656.
13. Johansson S.O., Hourihain J.B., Bousquet J. et al. Arevised nomenclature for allergy. An EAACI position statement from the EAACI nomenclature task force.//Allergy. - 2001. - Vol. 56. - P.813-824.
14. Koivikko A., Kalimo K., Nieminen E. et al. Allergenic cross-reactivity of yeasts.//Allergy. - 1988. - Vol. 43. - P.192-200.
15. Konig B., Neuber K., Konig W. Responsiveness of peripheral - blood mononuclear cells from normal and atopic donors to microbial superantigens.//Intern. Arch. Allergy Immunol. - 1995. - Vol. 106 (2). - P.124-133.
16. Leung D., Reiser R.F., Harbeck R. et al. *Staphylococcus aureus* grown from the skin of atopic dermatitis and psoriasis patients secrete toxins with superantigenic properties.//Clinical. Research. - 1991. - Vol. 39 (2). - P.422.
17. Leung D., Harbeck R., Bina P. et al. Presence of IgE antibodies to staphylococcal exotoxins on the skin of patients with atopic dermatitis - Evidence for a New Group of Allergens.//J. Clin. Invest. - 1993. - Vol. 92 (3). - P.1374-1380.
18. Leung D., Travers J.B., Norris D.A. The role of superantigens in skin disease.//J. Invest. Dermatology. - 1995. - Vol. 105 (1). - P.37-42.
19. Lintu P., Savolainen J., Kalimo K. IgE antibodies to protein and mannan antigens of *Pityrosporum ovale* in atopic dermatitis patients.//Clin. Exp. Allergy. - 1997. - Vol. 27 (1). - P.87-95.
20. Mokronosova M., Maksimova A., Batur A. The significance of skin colonization with *S.aureus* for differential diagnostic of atopic and seborrheic dermatitis.//Vestnik Venerol. Dermatol. - 1997. - Vol. 5. - P.37-40.
21. Mokronosova M., Lyapova T., Kochetova Ju. Skin reactivity to *Candida albicans* in the patients with atopic dermatitis.//Immunologia (Rus). - 2002 - Vol. 3. - P.161-164.
22. Nordvall S.L., Johansson S. IgE antibodies to *Malassezia spp.* in children with atopic diseases.//Acta Paediatr. Scan. - 1990. - Vol. 79. - P.343-348.
23. Ring J., Abeck D., Neuber K. Atopic Eczema - Role of microorganisms on the skin surface source.//Allergy. - 1992. - Vol. 47 (4). - P.265 -269.
24. Savolainen J., Lamintausta K., Kalimo N. et al. *Candida albicans* and atopic dermatitis. Clin. Exp. Allergy. - 1993. - Vol. 23. - P.332-339.
25. Severity scoring of atopic dermatitis: The SCORAD Index (Consensus report of the European Task Force on atopic dermatitis)///Dermatology. - 1993. - Vol. 186. - P.23-31.
26. Skov L., Baadsgaard O. Superantigens - Do they have a role in skin diseases.//Arch. Dermatology. - 1995. - Vol. 131 (7). - P.829-832.
27. Williams R., Gibson A.G., Aitchison T.C. et al. Assessment of contact-plate sampling technique and subsequent quantitative bacterial studies in atopic dermatitis.//Br. J. Dermatol. - 1990. - Vol. 123. - P.493-501.
28. Zargari A., Harfast B., Johansson S. et al. Identification of allergen components of the opportunistic yeast *Pityrosporum orbiculare* by monoclonal antibodies.//Allergy. - 1994. - Vol. 49 (1). - P.50-56.

SEASONAL DYNAMIC OF MITE ALLERGENS EXPOSURE (DER 1, DER 2) AND HOUSE DUST MITES IN MOSCOW'S APARTMENTS

T. Zheltikova, V. Gervajieva, M. Mokronosova

Mechnikov Research Institute for Vaccines and Sera

SUMMARY

It is well known that the number of pyroglyphid mites and, therefore, exposure to mite allergens varies during the year. Seasonal dynamic of mites has its specific geographical features, which plays a causal role in the development of bronchial hyperreactivity in patients with asthma. In this connection the monitoring of house dust mites and mite allergen levels in patients' homes should be considered as the strategy for the treatment of bronchial asthma. Our aim was the study of the seasonal dynamic house dust mites and mite allergens exposure (Der 1, Der 2) in some asthma patients' apartments in Moscow. In 1994-1996 the number of pyroglyphid mites and exposition to mite allergens of groups 1 and 2 were monitored in the apartments of asthma patients sensitized to house dust mites. The study was carried out in 5 apartments, random chosen as models, by means of the acarological analysis and ELISA with the use of monoclonal antibodies to mite allergens. The data obtained in this study made it possible to determine the periods of rises and falls in the concentration of mite allergens in the dust of apartments in Moscow. The seasonal dynamics of exposition to mite allergens of groups 1 and 2 had a regional character. One period of a rise in the concentration of mite allergens ($>2 \mu\text{g/g}$ of dust), common to all model apartments under study in Moscow, was established: the end of August - the beginning of October. All other periods of rises in the concentration of allergens were specific for each individual apartment. Thus, the prognostication of the periods of rises in the concentration of allergens in a patient's environment proved to be rather difficult. Therefore, prior to the prescription of specific immunotherapy to an individual patient exposition to mite allergens should be established and, with such concentration exceeding $2 \mu\text{g/g}$ of dust (100 mites/g of dust), the intensive sanitary and hygienic treatment of the apartment should be carried out.

Key words: mite allergens exposure, house dust mites, seasonal dynamic

Epidemiological studies, carried out all over the world, indicate the presence of dose-dependent relationship between sensitization and exposition to house-dust mite allergens (Der 1, Der 2) (1-3). It is well known that the number of house dust mites and, therefore, exposure to mite allergens varies during the year. Seasonal dynamic of mites has its specific geographical features, which plays a causal role in the development of bronchial hyperreactivity in patients with asthma. In this connection the monitoring of house dust mites and mite allergen levels in patients' homes should be considered as the strategy for the treatment of bronchial asthma (8). Our aim was the study of the seasonal dynamic house dust mites and exposure to mite allergens (Der 1, Der 2) in some asthma patients' apartments in Moscow.

MATERIAL AND METHODS

The study was carried out in 5 Moscow apartments, inhabited by asthma patients with pronounced sensitization to house dust mites and random chosen as models, during the period of 1994-1996. The apartments were located on

the lower stories (1-3) of multistory houses with central heating. The houses were built of bricks or concrete blocks. The area of the apartments varied from 30 to 65 sq. m, the number of inhabitants was 3-4 persons. The apartments were furnished with standard modern furniture.

The dust was collected from beds and bedding, by means of a household vacuum cleaner with the use of textile filters. The mites were detected by means of a binocular microscope (model МБС-3) in the water suspension of dust. The mite preparations were obtained with the use of 40% lactic acid. Sifted house dust samples were extracted in ammonium bicarbonate buffer solution (pH 8.2) (w/v 1:40) for 24 hours. After filtration the supernatant was stored at a temperature of -20°C .

The detection of Der p 1, Der p 2, Der f 1, Der f 2 was carried out by biotin-avidin ELISA method of Chapman M.D., described earlier (9).

RESULTS

In apartments Nos. 1-3 the presence of *D. farinae* population was established.

Apartment No 1. The number of mites varied from 20 to 1267 mites/g of dust. The concentration of allergens in the dust was $1.0-21.0 \mu\text{g/g}$ of dust and $0.1-20.0 \mu\text{g/g}$ of dust for Der f 1 and Der f 2 respectively. The number of mites below 100 mites/g of dust was registered from February to the end of April and in November 1995. A considerable rise in the number of mites (exceeding 500 mites/g of dust) was observed from the end of November to December 1994 and in September 1995. The mite allergens level exceeding $10 \mu\text{g/g}$ of dust was registered at the end of December 1994, the end of February and in September 1995 for Der f 1, at the end of November 1994 and the end of September 1995 for Der f 2 (Fig. 1A).

Apartment No 2. The number of *D. farinae* varied from 1 to 319 mites/g of dust. The concentration of Der f 1 and Der f 2 was $0.02-17.0 \mu\text{g/g}$ of dust and $0.02-11.0 \mu\text{g/g}$ of dust respectively. Out of 15 months of observation, during 11 months the number of mites was below 100 mites/g of dust. Four peaks in the number of *D. farinae* were noted: the beginning of May, the end of June, the beginning of October 1995 and the beginning of January 1996. Still the number of mites never reached 500 mites/g of dust during the whole term of observation. The level of Der f 1 exceeded $2 \mu\text{g/g}$ of dust at the end of June and the beginning of January 1996. During the remaining two periods of rises in the number of mites the concentration of allergens exceeded $10 \mu\text{g/g}$ of dust (at the beginning of May and October). At the same time the concentration of Der f 2 was equal and exceeded $2 \mu\text{g/g}$ of dust during 3 periods (the end of June, the beginning of October 1995 and the beginning of January 1996), and only at the beginning of May 1995 it exceeded $10 \mu\text{g/g}$ of dust (Fig. 1B).

Apartment No 3. The number of *D. farinae* varied from 20 to 1320 mites/g of dust. The mite allergens concentration varied from 0.5 to $84.0 \mu\text{g/g}$ of dust for Der f 1 and from 0.5 to $30.0 \mu\text{g/g}$ of dust for Der f 2. The periods when the number of mites was below 100 mites/g of dust were registered in April and December 1995, as well as in February-March 1996. At all other periods the number of mites never fell below 600 mites/g of dust. Changes in mite allergens exposure, both Der f 1 and Der f 2, showed practically complete correlation with the number of mites (Fig. 1C).

In apartments Nos. 4 and 5 the presence of coexisting *D. pteronyssinus* and *D. farinae* populations was established.

Apartment No 4. The number of *D. pteronyssinus* and *D. farinae* varied from 10 to 690 mites/g of dust and from 15 to 380 mites/g of dust respectively. At the same time the maximum number of *D. pteronyssinus* was almost twice greater

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CORRESPONDING AUTHOR:

T. Zheltikova
Mechnikov Research Institute
for Vaccines and Sera
Malij Kazennij pereulok 5a,
103064, Moscow, Russia
Fax: (095)917-49-00

than those of *D. farinae*. Still the average number of *D. farinae* exceeded that of *D. pteronyssinus* and reached 225 mites/g of dust in comparison with 159 mites/g of dust. The concentration of allergens varied, respectively, from 0.1 to 9.0 $\mu\text{g/g}$ of dust and from 0.1 to 10.0 $\mu\text{g/g}$ of dust for Der p 1 and Der p 2, from 1.7 to 10.3 $\mu\text{g/g}$ of dust and from 0.3 to 8.0 $\mu\text{g/g}$ of dust for Der f 1 and Der f 2. For *D. pteronyssinus* population only one peak of rise in its number was noted in June 1995 when the number of mites exceeded 500 mites/g of dust and exposure to Der p 1 and Der p 2 was 9.0 and 10.0 $\mu\text{g/g}$ of dust respectively. This peak was preceded by the period of rise, April-May 1995, when the number of mites exceeded 100 mites/g of dust and the concentration of allergens exceeded 2 $\mu\text{g/g}$ of dust. Such period of rise took place also in September 1995. At all other periods of observation the number of mites was below 100 mites/g of dust, and the concentration of allergens was below 2 $\mu\text{g/g}$ of dust (Fig. 2A).

The peak of density of *D. farinae* occurred at the end of May and September when the levels allergens Der f 1 exceeded 10 $\mu\text{g/g}$ of dust, though the density of these mites did not reach 500 mites/g of dust. Only in March 1995 the number of mites was below 100 mites/g of dust with exposure to Der p 2 exceeding 2 $\mu\text{g/g}$ of dust (Fig. 2B).

Apartment No 5. The number of *D. pteronyssinus* and *D. farinae* varied from 5 to 250 mites/g of dust and from 5 to 90 mites/g of dust respectively. At the same time the maximum number of *D. pteronyssinus* was almost 3 times and their average number was 2 times greater than those of *D. farinae*. The number of mites in this apartment did not reach 500 mites/g of dust during the whole period of observation. For *D. pteronyssinus*, three periods of rises were noted (the end of May, July-August and November 1995), when the number of mites was higher than or approximated to 100 mites/g of dust and the concentration of allergens exceeded 2 $\mu\text{g/g}$ of dust. The number of *D. farinae* did not reach 100 mites/g of dust for the whole period of observation (Figs. 2C, 2D).

The positive correlation between the number of mites (*D. pteronyssinus*, *D. farinae*) and the mite allergens exposure (Der p 1, Der p 2, Der f 1, Der f 2) was established: the coefficient of correlation (r) varied from 0.6647 to 0.9712 ($P < 0.01$).

DISCUSSION

Our data indicate the specific regional of the mite seasonal dynamics (*D. farinae* and *D. pteronyssinus*) and mite allergens exposure (Der 1 and Der 2) in Moscow' apartments, linked with the climate and mainly with the specific social and economic conditions of country life: the typical structure of dwellings (houses built of bricks and prefabricated sections) with central heating and hot water supply, standard arrangement of apartments furnished with standard furniture, bedding, toys, etc. We observed one period of the increasing of the mites populations and mite allergens (>2 $\mu\text{g/g}$ of dust) - the end of August - the beginning of October - common for the all Moscow's model apartments. The analysis of literature confirms the presence of specific regional features of house dust acarofauna and mite allergens exposure in different geographical regions (10-18).

The monitored of the seasonal dynamics of pyroglyphid mites and exposure to mite allergens of groups 1 and 2 in Moscow apartments, random chosen as models, showed that each individual apartment had a number of characteristic specific features, reflected in the average number of mite, the amplitude of density and at the periods of rises and falls in the mite populations. The microclimate of the apartment was found to play an even more important role in the seasonal dynamics of house dust mites than the geographical climate of the region (17, 19, 20). Some au-

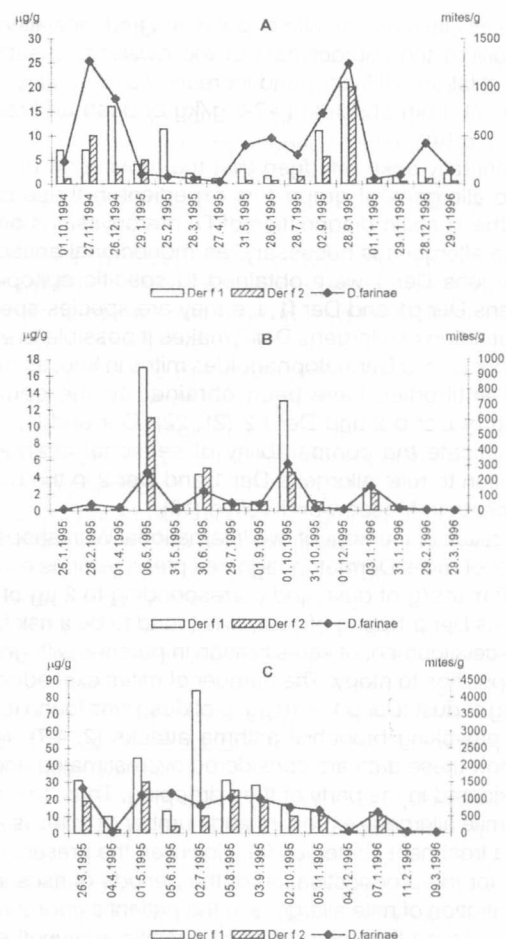


Figure 1. Seasonal dynamic of exposure to mite allergens (Der f 1, Der f 2) and number of *D. farinae* in dust of: A - apt. No 1; B - apt. No 2; C - No 3.

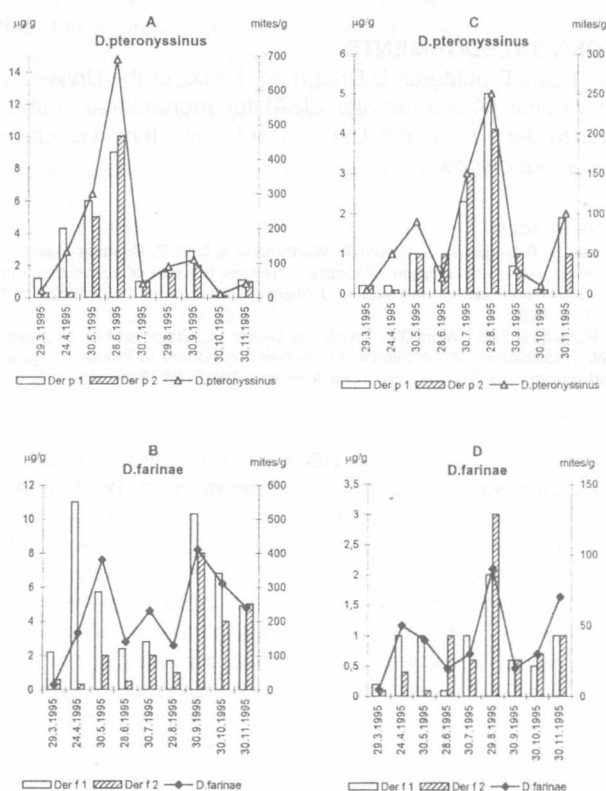


Figure 2. Seasonal dynamic of exposure to mite allergens (Der 1, Der 2) and number of *D. pteronyssinus* and *D. farinae* in dust of: A, B - apt. No 4; C, D - apt. No 5.

thors attribute a rise of mite populations and mite allergens exposure to the microclimate of the dwelling: unsatisfactory ventilation (<0.5 ach) and increased air humidity in the apartment, both absolute ($>7-9$ g/kg of dust) and relative (45%) (4, 6, 10).

It is generally acknowledged that the monitoring of exposure to allergens of group 1 is expedient in those cases when the specific diagnostics of *D. pteronyssinus* and *D. fari-nae* allergens is necessary, as monoclonal antibodies to allergens Der 1 were obtained to specific epitopes of allergens Der p1 and Der f1, i. e. they are species-specific. The detection of allergens Der 2 makes it possible to evaluate exposition to *Dermatophagoides* mites in toto, as monoclonal antibodies have been obtained to the common epitope of Der p 2 and Der f 2 (21, 22). Our and literature data indicate the comparability of seasonal changes in exposition to mite allergens Der 1 and Der 2 in the dust of apartments in Moscow and Tokyo (18).

As the result of the work of two International Workshops, the number of mites *Dermatophagoides pteronyssinus* exceeding 100 mites/g of dust, and corresponding to 2 μ g of mite allergens Der p 1 per g of dust, was found to be a risk factor for the development of sensitization in persons with genetic predisposition to atopy. The number of mites exceeding 500 mites/g of dust (Der p 1 = 10 μ g/g of dust) was found to be a factor provoking bronchial asthma attacks (2, 4-7). Moreover, now these data are considered overestimated and are reconsidered in the party of their dropping. Thus, the monitoring mite allergens exposure and number of mites is anew asthma treatment strategies (8). However, the presents difficulties for the prognostication of the periods of rises in the concentration of mite allergens in the patient's environment. For this reason the prescription of specific immunotherapy to an individual patient must be preceded by the determination of exposition to mite allergens. All these facts indicate that it is necessary to monitor the number of mites and exposition to mite allergens in different regions of the world.

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REFERENCES

1. Lau S, Falkenhurst G, Weber A, Werthmann I, Lint P, Buettner-Goetz P, Wahn U. High mite-allergen exposure increases the risk of sensitization in atopic children and young adults. *J Allergy Clin Immunol* 1989; 84: 718-725.
2. Platts-Mills TAE, Ward GW, Sporik RB, Gelber LE, Chapman MD, Heymann PW. Epidemiology of the relationship between exposure to indoor allergens and asthma. *Int Arch Allergy Appl Immunol* 1991; 94: 339-345.
3. Peat JK, Tovey E, Toelle BG, Haby MM, Gray EJ, Mahmic A, Woolcock AJ. House dust mite allergens - a major risk factor for childhood asthma in Australia. *American J Respiratory and Critical Care Medicine* 1996; 153, iss 1; 141-146.
4. Platts-Mills TAE, Sporik RB, Chapman MD, Heymann PW. The role of indoor allergens in asthma. *Allergy* 1995; 50, suppl 22: 5-12.
5. Kuehr J, Frischer T, Meinert R, Barth R, Foster J, Schraub S, Urbanek R, Karmaus W. Mite allergen exposure is a risk for the incidence of specific sensitization. *J Allergy Clin Immunol* 1994; 94: 44-52.
6. Munir AKM, Bjorksten B, Einarsson R, Ekstrand-Tobin A, Moller C, Warner A, Kjellman N-IM. Mite allergens in relation to home conditions and sensitization of asthmatic children from three climatic regions. *Allergy* 1995; 50: 55-64.
7. Pollart S, Platts-Mills TAE. Mites and mite allergy as risk factors for asthma. *Ann Allergy* 1989; 63: 364-365.
8. Chapman M.D., Tsay A., Vailes L.D. Home allergen monitoring and control -improving clinical practice and patient benefits // *Allergy*. 2001. V.56. P.604-610.
9. Chapman MD, Heymann PW, Wilkins SR, Brown MJ, Platts-Mills TAE. Monoclonal immunoassays for major dust mite (*Dermatophagoides*) allergens, Der p 1 and Der f 1, and quantitative analysis of the allergen content of mite and house dust extracts. *J Allergy Clin Immunol* 1987; 80: 184-94.
10. Platts-Mills T.A.E., Hayden M.L., Chapman M.D., Wilkins S.R. Seasonal variation in dust mite and grass-pollen allergens in dust from the houses of patients with asthma. *J Allergy Clin Immunol* 1987; 79: 781-791.
11. Petrova AD, Zheltikova TM. Seasonal dynamic of house dust mites population (*Acariformes*, *Pyroglyphidae*) in Moscow's apartments. *Biol Sci* 1990; 10: 37-45.
12. Colloff MJ. Practical and theoretical aspects of the ecology of house dust mites (*Acar: Pyroglyphidae*) in relation to the study of mite-mediated allergy. *Rev Med Veterinary Entomology* 1991; 79, 11/12: 611-630.
13. Heide S, Monchy JGR, Vries K, Bruggink TM, Kauffman HF. Seasonal variation in airway hyperresponsiveness and natural exposure to house dust mite allergens in patients with asthma. *J Allergy Clin Immunol* 1994; 93: 470-475.
14. Zheltikova TM, Ovsyannikova IG, Gervazieva VB, Platts-Mills TAE, Chapman MD, Petrova-Nikitina AD, Stepanova GN. Comparative detection of mite allergens in house dust of homes in Moscow by enzyme-linked immunosorbent assay and acarofogic analysis. *Allergy* 1994; 49: 816-819.
15. Echechipia S, Ventas P, Audicana M, Urrutia, Gastaminza G, Polo F, Fernandez de Cones L. Quantitation of major allergens in dust samples from urban populations collected different season in two climatic areas of the Basque region (Spain). *Allergy* 1995; 50: 478-482.
16. Matsuoka H, Maeda N, Atsuta Y, Ando K, Chinzei Y. Seasonal fluctuations of *Dermatophagoides* mite population in house dust. *Japanese J Med Sci & Biology* 1995; 48: 103-115.
17. Chanyung M, Becker A, Lam J, Dimichward H, Ferguson A, Warren P, Simons E, Manfreda J. House-dust mite allergen levels in 2 cities in Canada - effects of season, humidity, city and home characteristics. *Clinical Exp Allergy* 1995; 25: 240-246.
18. Miyazawa H, Sakaguchi M., Inouye S., Ikeda K., Honbo Y., Yasueda H., Shida T. Seasonal changes in mite allergen (Der 1 and Der 2) concentrations in Japanese homes. *Annals of Allergy Asthma and Immunology* 1996; 76, iss 2: 170-174.
19. Linter TJ, Brame KA. The effects of season, climate, and air-conditioning on the prevalence of *Dermatophagoides* mite allergens in household dust. *J Allergy Clin Immunol* 1993; 91: 862-867.
20. Chih-Shan Li, Gwo-Hwa Wan, Kue-Hsiung Hsieh et al. Seasonal variation of house dust mite allergen (Der p 1) in subtropical climate. *J Allergy Clin Immunol* 1994; 94, 1: 131-134.
21. Heymann PW, Chapman MD, Aalberse RC, Fox JW, Platts-Mills TAE. Antigenic and structural analysis of group 2 allergens (Der f 1 and Der p 2) from house dust mites (*Dermatophagoides* spp). *J Allergy Clin Immunol* 1989; 83: 1055-1067.
22. Lombardero M, Heymann PW, Platts-Mills TAE, Fox JW, Chapman MD. Conformational stability of B cell epitopes on group 1 and group 2 *Dermatophagoides* spp. allergens. *J Immunology* 1990; 144: 1353-1360.

STUDY OF THE FC FUNCTION OF THE IMMUNOGLOBULIN IN IMMUNOVENIN-INTACT

J. Natcheva*, Z. Mihneva**, A. Mihailov**, I. Bineva*

* BB-NCIPD Ltd.

** NCIPD

SUMMARY

Immunovenin-intact (immunoglobulin for intravenous use) contains immunoglobulin in its native form and is presented as ready to use liquid. The recommendation of the European Pharmacopoeia to prove the nativity of the IgG molecules (their Fc function) by the assessment of the level of rubella antibodies was followed. The measurements of these antibodies during the shelf life of the preparation showed that their levels were preserved and this gave assurance of the efficacy and stability of Immunovenin-intact. The method of rubella radial haemolysis was applied to prove that the Fc function of the immunoglobulin in this preparation was not impaired.

Key words: Fc function, immunoglobulin, Immunovenin-intact, rubella antibodies, radial haemolysis

Immunovenin-intact is a plasma derivative that belongs to the group of life-saving preparations. The product meets the requirements of the European Pharmacopoeia, 2002. The preparation is well characterized for protein concentration, pH, purity, sterility, pyrogenicity, viral safety, etc. The following key markers on the nativity of the immunoglobulin are regularly recorded: 1) anticomplementary activity; 2) molecular size distribution; 3) prekallikrein activator activity. The fourth key marker is the Fc function of the immunoglobulin in the product. The regular assessment of the Fc function of the immunoglobulin in Immunovenin-intact would give better assurance of its clinical efficacy. This is important because the indications for immunoglobulin therapy are numerous and their scope is increasing. Now the indications may be grouped into three main categories: adjuvant therapy in severe life-threatening infections; replacement therapy in primary and secondary antibody deficiency states; immunomodulation in immunoinflammatory conditions and autoimmune diseases. The European Pharmacopoeia describes a method for the testing of Fc function, which requires a rubella antigen of very high activity (above 256 haemagglutination units). That reagent was not available at the time of the study. The study was directed to the application of the method of single radial haemolysis (as approved by the British Pharmacopoeia) for the testing of the Fc function of immunoglobulin in Immunovenin-intact and to the comparison of the results with these obtained by two other methods for determination of rubella antibodies (haemagglutination inhibition test and indirect ELISA-IgG antibodies).

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ABBREVIATIONS USED IN THIS PAPER:

SRH - single radial haemolysis

HIT - haemagglutination inhibition test

CORRESPONDING AUTHOR:

I. Bineva

BB-NCIPD Ltd.,

26, Yanko Sakazov Blvd.

1504 Sofia, Bulgaria

MATERIALS AND METHODS

1. 29 lots of Immunovenin-intact (5% IgG) with expiry dates from November 2001 to March 2004, produced by BB-NCIPD Ltd were tested. The samples were heat treated for 20 minutes at 56 °C to remove any unspecific inhibitors. They were diluted 1:2 with phenobarbitone buffer.

2. International standard of EDQM - Human Immunoglobulin BRP

3. Rubella antigen with biological activity 1:128 haemagglutination units determined by the reaction of inhibition of haemagglutination (produced by NCIPD)

4. Positive and negative control sera

5. Reaction of single radial haemolysis (1, 2, 3, 4).

The following reagents were used:

- red blood cells of pigeon (8% suspension in phenobarbitone buffer, pH = 7,2) or red blood cells of sheep (5% suspension in phenobarbitone buffer, pH = 7,2)

- freshly prepared complement of guinea pig (1:73 titre)

- petri dishes with 1,5% agarose (Merck) in phenobarbitone buffer, pH = 7,2 with 0,05% sodium azide.

The method was performed as follows:

a) the red blood cells were sensitized with rubella antigen by mixing them 1:1 with the antigen and leaving them at +4 °C for 1 hour; then the cells were centrifuged at 1500 rpm/min for 5-10 min and washed twice with phenobarbitone buffer; the washed cell were suspended in phenobarbitone buffer to their initial volume

b) the agar plates (test and control) were prepared:

- 0,6 ml sensitized red blood cell suspension was mixed with 3,5 ml 1,5% agarose (warmed to 43 °C), and poured into a sterile petri dish; it was allowed to set and labeled "test"

- the control plate was prepared in the same way but the red blood cells were not sensitized

- wells were punched out; 10 µl samples of the studied Immunovenin-intact and the controls were filled in them; the samples were doubled

- the plates were left in a moist chamber till the next day; then the agarose was covered with the fresh complement (diluted 1:2 with phenobarbitone buffer) and the plates were incubated at 37 °C for 4-5 hours. The haemolysis was visualized and the diameters of the zones of red cell lysis were measured in mm

- zones with diameter less or equal to 5 mm were read negative.

6. Haemagglutination inhibition test (HIT)- the generally accepted method for the detection of rubella antibodies was applied

7. Indirect ELISA test kits for rubella IgG antibodies of ETI-Rubek plus (dua Sorin, Italy), ImmunoComb II Rubella IgG (Organics, Israel) or Platelia Rubella IgG (Bio Rad) were used

RESULTS AND DISCUSSION

The method of single radial haemolysis was performed as described above. It was based on the lysis of sensitized red blood cells with rubella antigen with the complex antigen-antibody formed in the presence of the studied product and with the addition of complement. The sensitized red blood cells were incorporated in the agarose gel and the area of the lysis was related to the amount of rubella antibodies (immunoglobulin with preserved Fc function). The method was applied to demonstrate that it could be used for the regular evaluation of the Fc function of immunoglobulin in Immunovenin-intact instead of the method published in the European Pharmacopoeia, 2002 (the method in the Pharmacopoeia required an antigen of very high activity that was not available at the time of the study). The use of pigeon or sheep red blood cells did not have signifi-

cant impact on the results. It seemed important to use fresh complement.

The expiry term of Immunovenin-intact is two years. The samples of the lots were divided in groups according to their state of expiry. The results of their testing by radial haemolysis (RH) were compared to the results by reaction of inhibition of haemagglutination (RIH) and presented in the following Table:

No of lots	Expiry term	SRH - diameter of haemolysed zone	HIT - titre
2	expired	8-10 mm	1:256/512
7	end of expiry term	10-14 mm	1:512/1024
4	1 1/2 year after release	7-10 mm	1:16/1024
6	1 year after release	10-14 mm	1:256/1024
6	1/2 year after release	10-13 mm	1:256/1024
4	at release	10-13 mm	1:512/1024
Immunoglobulin standard BRP		10-12 mm	1:128
(+) control serum	7-8 mm	1:16/32	
(-) control serum			

In all samples of Immunovenin-intact, tested by ELISA, the antirubella IgG content was above the highest standard of each kit.

The diameters of the haemolysed zones of 26 samples were equal or above to the results obtained with the immunoglobulin standard. The diameters were 10-14mm. Three samples (one - expired lot, two - 6 months before expiry) showed haemolysis at 7-8 mm. The agglutination inhibition titre of the one expired lot was 1:256 that was comparable to the results with the lots during their term of use. The smaller haemolysis diameters of the other two lots could be related to the lower HIT titres (1:16 and 1:128).

On the basis of these lower results, it may be concluded in this case that the Fc function in the product was impaired. All other samples showed HIT titre above the titre of the immunoglobulin standard and their results with the single radial haemolysis test were in concurrence. Beside the practically equal results in radial haemolysis for 26 of the 29 samples, the titres in RIH for 27 samples were 1:256 and above (the immunoglobulin standard gave 1:128 titre) that reflect the high level of rubella antibodies in the studied product and give another proof of its stability in the liquid form. The high rubella antibody content was confirmed by the ELISA test also.

The method of single radial haemolysis could be applied in the screening of Immunovenin-intact lots to evaluate the Fc function of the immunoglobulin in the product and this was an additional assurance of its efficacy and stability in the liquid form.

CONCLUSION

The state of intactness of the immunoglobulin in Immunovenin-intact was studied by the evaluation of its Fc function with the method of single radial haemolysis for the detection of rubella antibodies. The results obtained by this method were compared with these obtained by the haemagglutination inhibition test and ELISA. They were in good correlation.

The measurements of these antibodies during the shelf life of the preparation showed that their levels were preserved and this gave assurance of the efficacy and stability of Immunovenin-intact. The method of rubella radial haemolysis gave proof that the Fc function of the immunoglobulin in this product was not impaired.

REFERENCES

1. Appleton, P., A.Macrae. Comparison of radial haemolysis with haemagglutination inhibition in estimating rubella antibody. *J.Clin. Pathology*, 1978, 31, 479-482
2. Gee, B., B.Jordan, P.Mortimer. An assessment of radial haemolysis in the detection of rubella antibody. *J.Clin.Pathology*, 1978, 31, 35-38
3. Morgan-Capner, P., H.Puleen, J.Pattison, D.Bidwell, A.Bartlett, A.Voller. A comparison of three tests for rubella antibody screening. *J.Clin.Pathology*, 1979, 32, 542-545
4. Russell, S., S.Benjamin, M.Briggs, M.Jenkins, P.Mortimer, S.Payne. Evaluation of single radial diffusion technique for rubella antibody measurement. *J.Clin.Pathology*, 1978, 31, 521-526
5. British Pharmacopoeia, 2000
6. European Pharmacopoeia, 2002

COMPARATIVE CHARACTERIZATION OF ANTIGENS ISOLATED FROM *TOXOCARA CANIS*

I. Rainova

National Center of Infectious and Parasitic Diseases

SUMMARY

Toxocarosis is parasitic disease in humans, caused by the larval form of dog nematode *Toxocara canis*. For diagnosis of the disease different serological tests are used which demonstrate the presence of specific antibodies in serum specimens of suspected patients. In the development of the immunological methods two kinds of antigens are used mainly - somatic from adult forms and excretory-secretory (metabolic) from parasite larvae cultivated in vitro. The aim of the present work is to make comparative characterization of these two antigens using SDS-electrophoresis and to study their specificity in enzyme-linked immunosorbent assay (ELISA). The results showed that by electrophoretic separation in SDS-PAGE the two kinds of antigens differ in number of isolated fractions and their molecule mass in kDa. The testing in ELISA with sera from patients, reacted negative for toxocarosis in another tests and from patients with other parasitic diseases, showed that E/S antigen isolated from *T.canis* larvae is more useful and possess enough specificity as a diagnostic reagent.

Key words: *Toxocara canis*, somatic antigen, E/S antigen, SDS-PAGE, ELISA

Toxocarosis is a parasitic disease in humans, caused by the larval form of dog nematode *Toxocara canis*. The clinical signs of the disease induced by larval migration in organs and tissues are united in complex larva migrans. The specific laboratory diagnosis of this parasitosis is difficult, because *T.canis* larvae don't complete their life cycle in humans. For the practice the accurately diagnosis is connected with *Toxocara* IgG antibodies detection in sera samples of suspect for toxocarosis patients. The search of species antigens and their applying in contemporary immunological tests as ELISA and Western blotting is an object of many investigations. Up to now the serological diagnosis of toxocarosis is connected with the application of two antigens mainly - total extract from adult worms and excretory-secretory (E/S) products obtained from *in vitro* cultivated larvae. The aim of this study is to compare the both mentioned kinds of antigens isolated from *Toxocara canis* by SDS-PAGE and determine their specificity in ELISA.

MATERIALS AND METHODS

I. ANTIGENS

1. Total extract from adult *T.canis*:

Mature *T.canis* were harvested from naturally infected puppies (till 6 months) after anthelmintic treatment. Parasites were killed by freezing and then partially grinded in a Potter tissue grinder in saline. This crude material was centrifuged for 30 min. at 12000 rpm. The supernate was collected and kept at 4°C. The pellet was homogenized again by sonication for 10 min. and centrifuged under the previous conditions. The second supernate was collected, mixed with the supernate obtained after the first centrifugation and used as a total antigen from adult *T.canis*.

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CORRESPONDING AUTHOR:

I. Rainova, MD
NCIPD, Dept. of Parasitology
26, Yanko Sakazov blvd.
1504 Sofia, Bulgaria

2. Excretory/secretory (E/S) antigen from *T.canis* larvae:

Excretory/Secretory (E/S) products were obtained from *T.canis* larvae according to the method described by Fairbairn (2). In vitro hatched live larvae were cultured in Eagle's minimal essential medium (MEM) as described by Savigny (6). The culture was carried out at 37°C and the medium was changed weekly. Collected fluid containing E/S products was centrifuged, dialysed against water and concentrated by ultrafiltration (Amicon PM10). This concentrated fluid was used as E/S antigen for SDS-PAGE and ELISA.

Protein content was determined by absorbance at 280 nm.

3. Sera samples. Followed groups of sera were used in the study:

- A. From patients, negative for toxocarosis in another tests (Ridascreen *Toxocara* IgG) - 6 sera samples
- B. From patients with other parasitic diseases - 4 sera samples

II. METHODS

1. Polyacrylamide gel electrophoresis (SDS-PAGE):

SDS - electrophoresis was carried out on MiniProtein II (BioRad) apparatus according to the manufacturer's instructions under reduced conditions. Apparent molecular weights of tested *Toxocara* antigens were estimated by mobility relative to standard marker proteins (Sigma) between 14 000 - 70 000 Da.

2. Enzyme linked immunosorbent assay (ELISA):

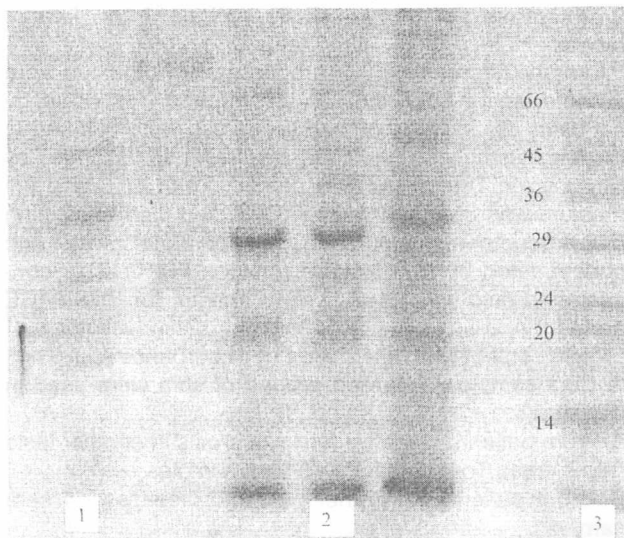
Indirect ELISA was performed according to van Knapen (7). Polystyrene strips were coated with solutions prepared from both kinds of antigens and contained 2 mg protein per ml carbonate buffer (pH 9.6). The initial sera dilution was 1:200 and the dilution of the antihuman goat IgG conjugated with horse radish peroxidase (HRP) was 1:4000 in PBS BSA Tween 20. The optical density was measured by ELISA Reader Uniscan (Labsystems) at 492 nm. Extinction value greater with 0.150 then extinction value of negative control serum was accepted as a cut off.

RESULTS AND DISCUSSION

In this study a comparative characterization by SDS-PAGE and ELISA of two kinds of antigens from parasite nematode *Toxocara canis* was accomplished.

SDS-PAGE analysis revealed the composition of tested antigens. Figure 1 and Table 1 show that the total antigen from adult worms is made up of more than 10 major components ranging in molecular weight from 108 000 to 15 000 kDa. Bands between 37 000 and 20 000 kDa are the brightest and cut clearly. Our results support the observations of other authors (1,3) detected in total *toxocara* extract using the same method till 30 fractions ranging from 330 000 to 18 000 Da. The differences between number of isolated fraction in the antigen, characterized by us and reported by other authors probably due to a some variety of the application of polyacrylamide electrophoresis as well as to standard marker proteins have been used.

When E/S antigen by SDS-PAGE was separated a considerably reduction of the number of isolated fractions was observed. Figure 1 and Table 1 show 6 components in this kind of antigen with molecular weight 63 000, 52 000, 40 000, 35 000, 29 000, 20 000 kDa. According to the previous studies (4) E/S antigen contains fractions with molecular weight 26 000-32 000, 55 000-70 000, 120 000 and 400 000 kDa. Antigens with molecular weight 32 000 kDa and 70 000 kDa are secreted in the most quantity and that with molecular weight 400 000 kDa can be detected with Schiff reagent for polysaccharides. Unfortunately till now we could not isolate in our



SDS-PAGE of *Toxocara canis* antigens

LEGEND : 1. E/S antigen from in vitro cultivated larvae; 2. Somatic antigen from adult forms; 3. Molecular weight standards (sigma)

antigen high molecular weight fractions, but as for some new investigations (5) specific for toxocarosis are fractions with lower molecular weight ranging between 24 000 and 35 000 kDa.

The second aim of the study is to test in ELISA the specificity of the two isolated *toxocara* antigens. The results obtained are performed on Table 2. There were tested ten sera samples divided into three groups:

- 6 sera samples from patients serologically negative for toxocarosis
- 1 serum sample from patient with proved ascaridosis
- 3 sera samples from patients with trichinellosis (clinical and serological proved).

Sera samples negative for toxocarosis tested with total antigen showed extinction values equal or higher than accepted cut off value. Sera samples from patients with ascaridosis and trichinellosis showed positive reaction. When sera samples were tested in ELISA with E/S antigen the extinction values showed negative results for more of them. False-positive reaction was observed only in one patient with trichinellosis. Our observations confirm those from other authors, who have also founded cross - reactions between E/S antigen from *T.canis* larvae and sera from patients with trichinellosis (8). Perhaps these cross-reactions occurred because of some common epitopes between E/S antigens from *Trichinella spiralis* and *Toxocara canis* larvae. Further investigations by ELISA in large number of patients will give more reliable results about the cross-reactivity observed.

The results obtained in ELISA reveal that total antigen from adult *T.canis* is not useful as a diagnostic reagent for

Table 1. SDS-PAGE analysis of *toxocara canis* antigens

N ^o Antigen of fractions	Number in kDa	Molecular weight
1. Somatic from adult	10	108, 100, 94, 88, 62, 37, 25, 23, 20, 15
2. E/S from in vitro cultivated larvae	6	63, 52, 40, 35, 29, 20

Table 2. Results in ELISA testing of the two *toxocara* antigens

Antigen	Patients sera samples	Number of sera samples	ELISA optical density at 492 nm	Result	
1. Somatic from adult forms	negative for toxocarosis	6	1.253	-	
			0.512	-	
			0.604	±	
			1.179	+	
			1.314	+	
				1.513	+
	ascaridosis	1	1.369	+	
	trichinellosis	3	1.121	+	
			0.550	±	
			1.097	+	
(+) c. - 1.91; (-) c. - 0.39					
2. E/S from in vitro cultivated larvae	negative for toxocarosis	6	0.333	-	
			0.313	-	
			0.219	-	
			0.244	-	
			0.204	-	
				0.187	-
	ascaridosis	1	0.305	-	
	trichinellosis	3	0.534	+	
			0.270	-	
			0.315	-	
(+) κ. - 1.65; (-) κ. - 0.215					

toxocarosis because the cross - reactions with sera samples from patients with another nematodoses. Our observations indicate that E/S antigens possess enough specificity for application in serological tests for diagnosis of human toxocarosis.

REFERENCES

1. Желева, Р.Ц. Медицинская паразитология и паразитарные болезни. 1975, 44, 293-298.
2. Fairbairn. Canadian Journal of Zoology. 1961, 39, 153-162.
3. Haque, M.R. et al. Helminthology, 1989, 27, 55-63.
4. Maizels, R.M. et al. Parasite Immunol. 1984, 6, 23-37.
5. Piarro, R. et al. VIII Europ. Multicollloquium of Parasitol. Poznan, 10-14 IX 2000.
6. Savigny, D.H. de. J.Parasit. 1975, 61(4), 781.
7. van Knapen, F et al. Z.Parasitenkd. 1983, 69, 113-118.
8. Zarnowska, H. and Jastrzebska, M. Acta Parasitologica, 1994, 39(1), 41-45.

EXSTRADIGESTIVE DISEASES ASSOCIATED WITH *H. PYLORI* INFECTION

I. Mladenova-Hristova

Dept. of Epidemiology, Medical Faculty, Stara Zagora

SUMMARY

H. pylori gastric infection is a chronic process lasting for decades. The persistent infection induces a chronic inflammatory and immune response able to induce lesions both locally and remote to the primary site of infection. *H. pylori* infection may be a risk factor for the development of coronary heart disease. In a recent cross-sectional study Mendall et al, confirmed that CHD is associated with C reactive protein and with high serum levels of IL-6 and TNF- α demonstrated predictors of myocardial infarction and stroke. These cytokines are associated with high levels of fibrinogen, sialic acid, triglycerides and reduced HDL-cholesterol. Following *H. pylori* infection, a link, between increased levels of cytokines and an increased level of C reactive protein, leading to CHD, can be hypothesized. A significant amelioration of both - Raynaud phenomenon and migraine symptoms has been described after *H. pylori* eradication. The prevalence of CagA(+) *H. pylori* strain was higher in infected subjects with autoimmune thyroiditis than in infected controls. An association with chronic idiopathic urticaria has been proposed with the pathogenetic mechanism related to an increase in gastric vascular permeability during *H. pylori* infection, resulting in a greater exposure of the host to alimentary allergens.

Key words: *H. pylori* infection, extradigestive diseases, coronary heart diseases

H. pylori is the main aetiological factor for gastritis and peptic ulcer disease. Its eradication is associated to healing of these diseases and significant reduction of ulcer recurrence and rebleeding. The characteristics of the infecting strain may influence the clinical outcome of the infection. The postulated role of *H. pylori* in the pathogenesis of extradigestive manifestations is based on the facts that: 1) local inflammation has systemic effects; 2) *H. pylori* gastric infection is a chronic process lasting for decades and 3) persistent infection induces a chronic inflammatory and immune response able to induce lesions both locally and remote to the primary site of infection.

Coronary heart disease

A number of studies have been published recently on the association between *H. pylori* infection and CHD. *H. pylori* infection, which is accompanied by a persistent inflammatory response, may be a risk factor for the development of coronary heart disease. If such is the case, it could at least partially explain the link observed between childhood poverty and adult CHD(2); on the other hand, older age, low socioeconomic status and being part of specific ethnic groups are all associated with both *H. pylori* infection and CHD and thus would be expected to confound the association. Furthermore, some studies have not supported this

hypothesis(3, 4). However, if *H. pylori* itself is responsible for the association, then this would be of great potential importance as the infection is treatable with a single course of antibiotics, and reinfection rarely occurs. In a recent literature review on chronic infections and CHD, Danesh et al.(5) pointed out the lack of homogeneity in *H. pylori* studies: few studies with a powerful design, heterogeneity in the definition of CHD, biased control groups, large variation in the adjustment of potential confounding factors, different sampled population.

An interesting aspect of this work was the method used to score the way counfounders were controlled in these studies. Using Danesh et al's method and updating with recent publications, the aim of our (18) meta-analysis on the association between *H. pylori* infection and CHD was to evaluate the degree of heterogeneity between studies and how the adjustment for confounding factors influenced the pooled odds ratio. The pooled odds ratio for the studies adjusted for control population score was 1.99 (95% CI [1.67-2.36]) when the control population was not appropriate and 1.25 (95% CI [1.06-1.47]) when the control population was randomly chosen in the general population. The studies with no appropriate control population were very heterogeneous ($p < 0.001$) while studies with an appropriate population control were less heterogeneous with a p value borderline for significance (0.053).

The stomach is a large organ and billions of *H. pylori* are present at any moment. The inflammation induced is intense as is the humoral response. It is, therefore, plausible that this host response has consequences outside of the target organ. Different but not necessarily alternative mechanisms are hypothesized to be involved. Tumor Necrosis Factor α (TNF α) which is elevated in blood following *H. pylori* infection(12) is a good candidate. Neri et al. also found an increased level of soluble TNF α receptor I in sera of *H. pylori* patients suffering from CHD(11). Patel et al. demonstrated that *H. pylori* infection was accompanied by acute phase reactants such as fibrinogen, white cell count and sialic acid, which are predictors of CHD(13, 14). However, such findings were not confirmed by other works(8,15). In a recent cross-sectional study Mendall et al.(16), confirmed that CHD is associated with C reactive protein and with high serum levels of IL-6 and TNF- α demonstrated predictors of myocardial infarction and stroke(17). These cytokines are associated with high levels of fibrinogen, sialic acid, triglycerides and reduced HDL-cholesterol. Following *H. pylori* infection, a link, between increased levels of cytokines and an increased level of C reactive protein, leading to CHD, can be hypothesized.

Raynaud phenomenon and prymary headache

H. pylori infection has been linked with functional vascular disorders (19,20). A direct or indirect release of vasoactive substances has been proposed as the pathogenic mechanism. The persistent immunological and inflammatory response to *H. pylori* infection may influence peripheral vessels through an immune-mediated release of vasoactive substances such as cytokines (interleukin, interferon- γ , TNF- α), prostaglandins, oxyradicals, C- reactive protein or fibrinogen. Most of these factors are persistently enhanced during *H. pylori* infection.

Prymary headache has recently been implicated as related to *H. pylori* infection. In some investigation clinical attacks complitely has been disapeared in the successefully treated patients or the intensity, duration and frequency of attacks has been reduced. The actuale question should be whether headache is relieved following cure of *H. pylori* infection and thus a low prevalence of *H. pylori* does not

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ABBREVIATIONS USED IN THIS PAPER:

CHD - Coronary Heart Diseases

CORRESPONDING AUTHOR:

Dr. I. Mladenova - Hristova, MD, PhD
Dept. of Epidemiology
Medical Faculty
17, Atanas Iliev Str.
6000, Stara Zagora, Bulgaria

exclude a role for *H. pylori*. The treatment trials are incomplete in that there are other infections besides *H. pylori* that can cause headache (sinus infection).

A significant amelioration of both - Raynaud phenomenon and migraine symptoms has been described after *H. pylori* eradication (21).

Autoimmune diseases and *H. pylori*

Autoimmune thyroiditis, Schonlein-Henoch purpura etc.

The prevalence of CagA(+) *H. pylori* strain was higher in infected subjects with autoimmune thyroiditis than in infected controls (22). The pathogenic mechanisms relating *H. pylori* to any of these immunological diseases are unknown. An alternative hypothesis is the production of autoantibodies induced by the bacteria through cross-reactions mechanism. *H. pylori* have similar epitopes to those found on gastric epithelium, salivary gland ducts and endometrium.

Skin diseases

An association with chronic idiopathic urticaria has been proposed with the pathogenetic mechanism related to an increase in gastric vascular permeability during *H. pylori* infection resulting in a greater exposure of the host to alimentary allergens. The possibility that patients with urticaria develop specific IgE against *H. pylori* is an attractive explanation that requires confirmation. Although a few trials of chronic urticaria have reported improvements in several patients following *H. pylori* eradication (23, 24) these studies were small, non-randomized and lacked any comparison groups. *H. pylori* infection was reported in 84% of 31 patients with rosacea. Disappearance of the clinical manifestations after *H. pylori* eradication has also been described, but rosacea was already known to be an antibiotic-responsive skin condition and the link is therefore tenuous (25). A possible association between *H. pylori* infection and idiopathic sideropenic anemia has been suggested. The resolution of anemia has been observed in some cases after cure of the infection. Several case reports have suggested reversal of iron deficiency anaemia refractory to iron replacement after *H. pylori* eradication (26,27) as has one trial of antibiotics in 30 *H. pylori* positive patients with persistent iron deficiency refractory to iron replacement therapy (with significant improvements in hematological parameters such as serum ferritin values). Another interesting clinical manifestation related to *H. pylori* infection is growth retardation. A possible pathogenic mechanism to explain the relationship relates to the increased level of circulated cytokines, such as TNF α , associated with *H. pylori* infection.

Future studies should follow three directions. First epidemiological studies should be large enough for moderate-sized effects to be assessed or registered reliably. The effect of residual confounders should be kept to a minimum. Second pathogenic mechanism should be investigated by comparison of plasma concentration of inflammatory markers in patients positive and negative in serum for microorganism. Third, because antibiotics are not specific for *H. pylori* infection appropriate controls include those with the same condition and without *H. pylori* infection placebo controls and longer follow-up after therapy.

REFERENCES

1. Patel P, Mendall M, Northfield T. Manifestations extra-digestives associées à l'infection à *Helicobacter pylori*. In: Mûgraud F, Lamouliatte H, ed. *Helicobacter pylori*. Elsevier, Paris Vol 2. 1997; 309-316.
2. Mendall M, Goggin M, Molineaux N, et al. Relation of *Helicobacter pylori* infection and coronary heart disease. *Br Heart J* 1994; 71: 437-9.
3. Maier F, Auricchio A, Nilius M, et al. Lack of association between *Helicobacter pylori* infection and angiographically documented Coronary Heart Disease. *Gut* 1996; 39 Suppl 2: A92 (abstract).
4. Delaney BC, Hobbs FDR, Holder R. Association of *Helicobacter pylori* infection with coronary heart disease - Eradication of the infection on grounds of cardiovascular risk is not supported by current evidence. *BMJ* 1996; 312: 251-252.
5. Danesh J, Collins R, Peto R. Chronic infections and coronary heart disease: is there a link? *Lancet* 1997; 350: 430-436.
6. Martin de Argila C, Boixeda D, Canton R, et al. High seroprevalence of *Helicobacter pylori* infection in Coronary Heart Disease. *Lancet* 1995; 346: 310.
7. Morgando A, Sanseverino P, Perotto C, et al. *H. pylori* seropositivity in myocardial infarction. *Lancet* 1995; 345: 1380.
8. Murray LJ, Bamford KB, O'Reilly DPJ, et al. *Helicobacter pylori* infection: Relation with cardiovascular risk factors, ischaemic heart disease, and social class. *Br Heart J* 1995; 74: 497-501.
9. Ponsetto A, Larovere MT, Sanseverino P, et al. Association of *Helicobacter pylori* infection with coronary heart disease - Study confirms previous findings. *BMJ* 1996; 312: 251.
10. Treiber G, Schneider E, Adolph C, et al. *Helicobacter pylori* (HP), Coronary Heart Disease (CHD) and Plasma Fibrinogen: is there a causal link? *Gut* 1996; 39 Suppl 2: A29 (abstract).
11. Neri M, Reale M, Febbo CD, et al. Increased levels of soluble tumour necrosis factor receptor I (sTNF RI) in serum of *H. pylori* positive IHD patients. *Gastroenterology* 1996; 110: 209 (abstract).
12. Crabtree J. Cytokine responses to *Helicobacter pylori* infection. In: Riecken EO ZM, Stallmach A, Heise W, ed. *Malignancy and chronic inflammation in the gastrointestinal tract: new concepts*. Lancaster: Kluwer Academic Publisher, 1995: 25-36.
13. Lindberg G, Eklund G, Gullberg B, et al. Serum sialic acid concentration and cardiovascular mortality. *BMJ* 1991; 302: 143-6.
14. Yamell J, Baker I, Sweetnam P, et al. Fibrinogen, viscosity, and white blood cell count are major risk factors for ischaemic heart disease. The Caerphilly and Speedwell collaborative heart disease studies. *Circulation* 1991; 83: 836-44.
15. Parente F, Bianchi Porro G. The association between *Helicobacter pylori* infection and ischemic heart disease: facts or fancy? *Helicobacter* 1997; 2: S67-S72.
16. Mendall M, Patel P, Asante M, et al. Relation of serum cytokine concentrations to cardiovascular risk factors and coronary heart disease. *Heart* 1997; 78: 273-277.
17. Ridker PM, Cushman M, Stampfer MJ, et al. Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men. *N Engl J Med* 1997; 336: 973-979.
18. Pellicano R, Mladenova I, Broutet N, Salmi LR. Is there an association between *H.pylori* infection and coronary heart disease? *Eur J Epidemiol* 1999;15:611-699.
19. Gasbarrini A et al. Association of *H.pylori* infection with primary phenomenon. *Lancet* 1996;348:966-967.
20. Gasbarrini A. et al. *H.pylori* and Reindau phenomenon. *Gastroenterol Int* 1997 (suppl 1):18-19.
21. Gasbarrini et al. Beneficial effects of *H.pylori* on migraine. *Hepatogastroenterology* 1998;21:765-770.
22. De Luis DA. et al. *H.pylori* infection is markedly increased in patients with autoimmune atrophic thyroiditis. *J Clin Gastroenterol* 1998;26:259-263.
23. Di Campli et al. Beneficial effects of *H.pylori* eradication on idiopathic chronic urticaria. *Dig Dis Sci* 1998; 43:1226-1229.
24. Wustlich S. et al. *H.pylori* as a possible focus of chronic urticaria. *Dermatology* 1999;198:130-132.
25. Sharma VK et al. A study of the prevalence of *H.pylori* infection and other markers of upper gastrointestinal tract disease in patients with rosacea. *Am J Gastroenterol* 1998;93:220-222.
26. Marignani M. et al. Reversal of long -standing iron deficiency anaemia after eradication of *H.pylori* infection. *Scand J Gastroenterol* 1997;32:617-622.
27. Kepczyk T et al. A prospective multidisciplinary evaluation of premenopausal women with iron deficiency anaemia. *Am J Gastroenterol* 1999;94:109-115.
28. Gasbarrini A, Ponsetto A et al. *H.pylori* infection and extradigestive diseases. *Curr Op Gastroenterol* 1998;14(suppl 1):65-69.
29. Gasbarrini A, Franceschi F. et al. Role of *H.pylori* in extradigestive diseases. *Curr Op Gastroenterol* 1999;15(suppl 1):29-33.
30. Danesh J, Gasbarrini A, et al. *H.pylori* infection and extradigestive diseases. *Curr Op Gastroenterol* 2000;16(suppl 1):52-55.

TRANSMISSION OF HELICOBACTER PYLORI: EVIDENCE FOR FECAL- ORAL ROUTE

I. Mladenova-Hristova

Dept. of Epidemiology, Medical Faculty, Stara Zagora

SUMMARY

The evidence for the fecal-oral route of transmission of *H. pylori* is still controversial. *H. pylori* DNA has been detected in feces of infected subjects by some researchers but not others. It has been demonstrated that *H. pylori* can live for several days in milk and tap water in its infectious bacillary form and in river water for several months in a coccoid form. Transmission has been linked to consumption of vegetables grown in regions where human feces is used as fertilizer. Although the principal reservoir for *H. pylori* infection appears to be people, *H. pylori* has been isolated from nonhuman primates and domestic cats. The most recent reservoir suggested for *H. pylori* transmission is the housefly. However, evidence is lacking that *H. pylori* can be isolated from flies that have been in contact with *H. pylori* infected feces and that *H. pylori* can be transmitted from contaminated flies to food in a quantity sufficient to infect humans. In fact, nosocomial transmission of *H. pylori* is the only proven mode of transmission. Understanding the route(s) by which *H. pylori* infection spreads is fundamental for development of strategies for the infection's control.

Key words: *H. pylori*, transmission, fecal-oral route

Although it is now clear that the reservoir of *Helicobacter pylori* is limited to the stomach of humans, the routes of transmission still remain unproven. As for other pathogens of the digestive tract, it is logical to consider a fecal-oral transmission as a probable "path". However, the evidence is still controversial. Several studies performed DNA fingerprinting to determine the specific strains of *H. pylori* harbored by family members' (8). They found the same strain of *H. pylori* to be present in a small percentage of spouses and siblings. These studies, taken as a whole, lend support to the concept of **interfamilial clustering** of *H. pylori* infection. They suggest that person-to-person transmission occurred in these families possibly because of close interpersonal contact, that family members shared a genetic predisposition to *H. pylori* infection, that family members were exposed to a common source of infection, or that spouses' childhood socioeconomic class was similar. *H. pylori* DNA has been **detected in feces** of infected subjects by some researchers (9,10) but not others (13). Recently Gramley et al found detectable *H. pylori* DNA in the feces of 73% of infected subjects. Isolation of *H. pylori* by fecal culture has been performed by a number of investigators from around the world (9,10,14). Recently, Parsonnet et al were able to culture *H. pylori* from cathartic- induced diarrheal stools in 7 of 14 *H. pylori* infected subjects but not from normal stools (11). Studies by Hazell et al. in China (15) and Webb et al in United Kingdom noted serum anti-

bodies to *H. pylori* and Hepatitis A, a sensitive marker of fecal- oral exposure, but did not find strong evidence supporting community- wide fecal- oral spread of *H. pylori* via food or water.

It has been demonstrated that *H. pylori* can live for several days in **milk and tap water** in its infectious bacillary form (16,17) and in river water for several months in a coccoid form. However, the idea of coccoid forms is very controversial. While experimental tests have shown that, under physical or chemical stress, *H. pylori* is able to convert to a viable but nonculturable coccoid form (18), researchers have failed to convert a coccoid to a bacillary form in culture (19). In addition, it has not yet been determined whether *H. pylori* can revert from its coccoid to its infectious form in humans. Support for waterborne transmission comes from epidemiological studies conducted in Colombia, rural China, and Lima, Peru, that found that water source may be related to risk of *H. pylori* infection (21). The finding of *H. pylori* positive drinking and sewage water samples by PCR assays in Peru and Japan (15,20) provides additional evidence that waterborne transmission may be important, especially in areas of the world that have high rates of *H. pylori* infection and less than adequate water quality.

Transmission has been linked to consumption of **vegetables** grown in regions where human feces are used as fertilizer.

It has been suggested that *H. pylori* infection may, in some instances, be a zoonosis. Although the principal reservoir for *H. pylori* infection appears to be people, *H. pylori* has been isolated from **non-human's primates and domestic cats** (23,24,26). Under controlled laboratory conditions, human *H. pylori* has been shown to infect monkeys. However, even if *H. pylori* occurs naturally in monkeys, they are unlikely to represent a major route of transmission to humans, since close contact between nonhuman primates and humans is limited in most of the world. Handt et al was the first to report isolation of *H. pylori* from domestic cats (24). Their laboratory was able to experimentally infect naive cats with *H. pylori*, to culture *H. pylori* from feline salivary and gastric sections, and to find *H. pylori* DNA in feline feces and dental plaque. A study in Germany by Rothenbacher et al. (25) found that adults who owned a cat as a child had a significantly higher prevalence of *H. pylori* infection.

Gnotobiotic pigs have been successfully infected with *H. pylori*. However, there is currently no convincing evidence that swine are a reservoir for *H. pylori*, even though the monogastric pig stomach is anatomically and physiologically similar to human and nonhuman primate stomachs (26).

Exposure to sheep was implicated in two recent epidemiological studies. In a study of children from the Colombian Andes, Goodman et al. (21) reported that children who "played with sheep" had a higher risk of *H. pylori* infection (OR=4.5). A study by Dore et al. (2) revealed that the prevalence of *H. pylori* was significantly higher among Sardinians shepherds occupationally exposed to sheep than among their nonexposed family members or among Sardinian blood donors. While tending sheep shepherds live usually in small enclosures by themselves or sometimes with their families, far from villages. The enclosures lack elementary sanitation and are potentially at high risk for diseases transmitted by the fecal- oral route.

Nevertheless, we can not exclude the possibility that *H. pylori* could survive in sheep. This hypothesis is currently under investigation.

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CORRESPONDING AUTHOR:

Dr. I. Mladenova - Hristova, MD, PhD
Dept. of Epidemiology
Medical Faculty
17, Atanas Iliev Str.
6000, Stara Zagora, Bulgaria

H. pylori has been cultured from stool and has been identified in feces by PCR. *H. pylori* can survive in milk (31) for several days, suggesting that milk contaminated by feces containing feces could be potentially infectious to humans. Alternatively, animals might transmit the infection to humans through the environment contaminated by feces containing *H. pylori* or the coccoid forms. If animals, on occasion, play a role in transmission, the cycle of infection might include phases in the environment, animals /sheep or dogs/, and human beings.

The most recent reservoir suggested for *H. pylori* transmission is the **housefly**. In controlled experimental studies, Grubel et al. found that houseflies infected with *H. pylori* in the laboratory could harbor viable *H. pylori* bacteria in their intestines as well as on their body hairs (4,27). However, evidence is lacking that *H. pylori* can be isolated from flies that have been in contact with *H. pylori* infected feces and that *H. pylori* can be transmitted from contaminated flies to food in a quantity sufficient to infect humans. In a recent experimental study by Osato et al. researchers were unable to recover *H. pylori* from houseflies that were fed human feces infected with *H. pylori* (3). The fact that they were unable to recover *H. pylori* from houseflies that were exposed to stool containing approximately 9×10^7 CFU per g of stool indicates that even higher levels of viable organisms must be present in nature to ensure positive recovery from flies. Previous data have shown that such high levels of viable *H. pylori* may not be present in the extragastric environment (3,4). Therefore, it seems less likely that the domestic housefly serves as a vector for *H. pylori* transmission.

Endoscopy is a common medical procedure used to diagnose and manage gastrointestinal disease. Because of the complex structure of the endoscope and difficulty in disinfecting it, the possibility of iatrogenic infection in-patients following endoscopy is a potential risk factor not only for *H. pylori* but also for other infectious diseases such as hepatitis C, hepatitis B, tuberculosis and possibly HIV (28,29). **In fact, nosocomial transmission of *H. pylori* is the only proven mode of transmission.** According to Tytgat et al, the rate of iatrogenic infection may approximate four per 1000 endoscopies (0.4%) when the prevalence of *H. pylori* is around 60% (30). In 1990 the working party report to the World Congresses of Gastroenterology recommended that the endoscope be soaked in 2% activated glutaraldehyde for at least 5-10min. Furthermore, because biopsy forceps typically penetrate the gastric mucosa and are difficult to clean sterilization of the forceps or, preferably use of disposable forceps is essential (30).

Understanding the route(s) by which *H. pylori* infection spreads is fundamental for development of strategies for the infection's control.

Knowledge of the Epidemiology and mode of transmission of *H. pylori* may be useful in identifying high- risk populations, especially in areas that have high rates of gastric lymphoma, gastric cancer and gastric ulcer. Different studies show contradictory results for a fecal- oral route as well as for other types of transmission. The difficulty is linked to the lack of adequate markers for each route of transmission and to the problems in accurately diagnosing *H. pylori* infection, especially in children, where symptoms are commonly nonexistent. Studies have to be designed in families from high *H. pylori* infection prevalence areas.

REFERENCES

- Warren JR., Marshall B. Unidentified curved bacilli on gastric epithelium in active chronic gastritis. *Lancet* 1983;1:1273-5.
- Dore MP, Bilotta M, Vaira D et al. High prevalence of *H. pylori* infection in shepherds. *Dig Dis Sci* 1999; 44:1161-4.
- Osato MS, Le HH et al. Houseflies are an unlikely reservoir or vector for *H. pylori*. *J Clin Microbiol* 1998; 36:2786-8.
- Grubel P, Huang L, Masubuchi N et al. Detection of *H. pylori* DNA in houseflies (*Musca domestica*) on three continents. *Lancet* 1998; 352:788.
- Rothenbacher D, Bode G, Berg G et al. *H. pylori* among preschool children and their parents: evidence of parent-child transmission. *J Infect Dis* 1999; 179:398-402.
- Malaty H, Kumagai T, Tanaka E et al. Evidence from a nine -year birth cohort study in Japan of transmission pathways of *H. pylori* infection. *J Clin Microbiol* 2000; 38:1971-1973.
- Parente F, Maconi G, Sangaletti O et al. Prevalence of *H. pylori* infection and related gastroduodenal lesions in spouses of *H. pylori* positive patients with duodenal ulcer. *Gut* 1996;39:629-33.
- Schultze K, Hentchel E, Dragosics B et al. *H. pylori* reinfection with identical organisms : transmission by the patients' spouses. *Gut* 1995;36:831-3.
- Namavar F, Roosendaal R, Kuipers EJ et al. Presence of *H. pylori* in the oral cavity, oesophagus, stomach and faces of patients with gastritis. *Eur J Clin Microbiol Infect Dis* 1995; 14- 234-7.
- Shimada T, Ogura K, Ota S et al. Identification of *H. pylori* in gastric specimens, gastric juice, saliva and faeces of Japanese patients. *Lancet* 1994;343:1636-7.
- Parsonnet J, Shmueli H, Haggerty T. Fecal and oral shedding of *H. pylori* from healthy infected adults. *JAMA* 1999, 282:2240-5.
- Gramley WA, Asghar A, Frierson HFJ et al. Detection of *H. pylori* DNA in fecal samples from infected individuals. *J Clin Microbiol* 1999;37:2236-40.
- Van Zwet AA, Thijs JC, Kooistra - Smid AM et al. Use of PCR with feces for detection of *H. pylori* infection in patients. *J Clin Microbiol* 1994;32:21346-8.
- Kelly SM, Pitcher MC, Farmery SM et al. Isolation of *H. pylori* from feces of patients with dyspepsia in the United Kingdom. *Gastroenterology* 1994;107:1671-4.
- Hazell SL, Mitchell HM, Hedges M et al. Hepatitis A and evidence against the community dissemination of *H. pylori* via feces. *J Infect Dis* 1994;170:686- 9.
- Hulten K, Han SW, Enroth H et al. *H. pylori* in the drinking water in Peru. *Gastroenterology* 1996; 110:1031-5.
- Fan XG, Chua A, Li TG et al. Survival of *H. pylori* in milk and tap water. *J Gastroenterol Hepatol* 1998;13:1096-8.
- Bode G, Mauch F, Malfredheiner P. The coccoid forms of *H. pylori*. Criteria for their viability. *Epidemiol Infect* 1993;111:483-90.
- Sorberg M, Nilson M, Hanberger H et al. Morphologic conversion of *H. pylori* from bacillary to coccoid form. *Eur J Clin Microbiol Infect Dis* 1996;15:216-19.
- Sasaki K, Tajiri Y, Sata M et al. *H. pylori* in the natural environment. *Scand J Infect Dis* 1999;31:275-9.
- Goodman KJ, Correa P, Tengana Aux HJ et al. *H. pylori* infection in the Colombian Andes: a population - based study of transmission pathways. *Amer J Epidemiol* 1996;144:290-9.
- Klein PD, Graham DY, Gailour A et al. Water source as risk factor for *H. pylori* infection in Peruvian children. *Gastrointestinal Physiology Working group. Lancet* 1991;337:1503-6.
- Dubois A, Berg DE, Incecik ET et al. Transient and persistent experimental infection of nonhuman primates with *H. pylori* : implication for human disease. *Infect Immun* 1996;64:2885-91.
- Handt LK, Fox JG, Dewhirst FE et al. *H. pylori* isolated from the domestic cat: public health implications. *Infect Immun* 1994;62:2367-74.
- Rothenbacher D, Bode G, Peschke F et al. Active infection with *H. pylori* in an asymptomatic population of middle aged to elderly people. *Epidemiol Infect* 1998;120:297-303.
- Fox JG. Non-human reservoirs of *H. pylori*. *Aliment Pharmacol Ther* 1995;9(suppl 2):93-103.
- Grubel GS, Hoffman JS, Chong FK et al. Vector potential of houseflies (*Musca Domestica*) for *H. pylori*. *J Clin Mikrobiol* 1997;35:1300-3.
- Axon AT. Disinfection of endoscopic equipment. *Baillieres Clin Gastroenterol* 1991;5:61-77.
- Fantry GT, Zheng QX et al. Conventional cleaning and disinfection techniques eliminate the risk of endoscopic transmission of *H. pylori*. *Am J Gastroenterol* 1995;90:227-32.
- Tytgat GN. Endoscopic transmission of *H. pylori*. *Aliment Pharmacol Ther* 1995;9(suppl 2):105-10.
- Dore M, Sepulveda A, Osato M et al. *H. pylori* in sheep milk. *Lancet* 1999, 354:132.

CALGEVAX

COMPOSITION

It is composed of lyophilized living bacteria of Calmette-Guerin (BCG). Each ampoule contains 37.5 mg living BCG ($1.0 - 3.0 \times 10^8$ live units). As stabilizer 8% sodium glutamate is added. The preparation contains no conservants.

INDICATIONS

CALGEVAX is prescribed as an additional treatment for all types of malignant tumors requiring adjuvant therapy, especially for vesicle tumors, malignant melanoma, etc.

CALGEVAX is especially active in case of superficial tumors of the bladder. It is applied intravesically in the following cases:

- for prophylaxis of recidivations after transurethral resection of superficial tumors;
- for treatment of carcinoma in situ of the bladder.

CALGEVAX is used for treatment of malignant melanoma as well as for non-specific immunostimulating adjuvant therapy in other malignancies (tumors of the lungs, mammary glands, acute leukemia, lymphosarcoma, osteosarcoma, etc.)

ADMINISTRATION AND DOSAGE

Intravesical administration: CALGEVAX is applied most early 14 days after a biopsy, transurethral resection or traumatic catheterization.

It is carried out following a strictly aseptic technique and under specialist's control. Three or four ampoules of CALGEVAX are needed for each intravesical application. The standard application scheme consists of one intravesical infusion per week during six weeks (induction therapy). The maintaining therapy depends on the individual case.

The following schema may be applied: monthly infusion in the course of at least 6-12 months or weekly, during 3 consecutive weeks, at 3, 6, 12, 18, 30 and 36 months after the first infusion.

Percutaneous application: On the spot of scarification 0.5 ml of the preparation is applied. One ampoule of CALGEVAX is dissolved in 0.5 ml saline. For one scarification is used one ampoule.

PACKAGE

A box with five or ten ampoules CALGEVAX.

On each ampoule and box are marked: the name of the producer, the preparation name, the BCG contents in mg, the number of the lot and expiry term.

SHELF LIFE

Not more than 24 months from the date of control test and not more than 30 months from the date of production.

STORAGE

In a dry and dark place at $+2^{\circ}\text{C} - +8^{\circ}\text{C}$.

PRODUCER

BB-NCIPD, Ltd.

Bulgaria, 1504 Sofia; 26, Yanko Sakazov Blvd.

General Manager - phone: +359 2/ 944 61 91

Marketing - phone: +359 2/ 944 81 19

fax: +359 2/ 943 30 75

PRODUCTS INSPIRED FROM LIFE!

CALGEVAX

UROSTIM

COMPOSITION

Each tablet UROSTIM contains 25 mg for children and 50 mg for adults of freeze-dried with excipient lysate and killed bacterial bodies of the following microbial species: *Escherichia coli*; *Proteus mirabilis*; *Klebsiella pneumoniae*; *Enterococcus faecalis*.

INDICATIONS

UROSTIM is intended for peroral immunotherapy and immunoprophylaxis of acute recurrent or chronic non-specific infections of the urinary tract in children and adults, regardless of the agent type. It is recommended for the treatment of cystitis, pyelonephritis, urethritis, asymptomatic bacteriuria, etc. Can be used with antibiotic therapy and especially for treatment of patients with suppressed immune reactivity obtained in result of a long antibiotic therapy.

CONTRAINDICATIONS

The drug has an excellent tolerance and no contraindications.

ADMINISTRATION AND DOSAGE

For the purposes of immunotherapy UROSTIM is administered as follows:

Adults: 50 mg daily. Basic course of treatment: one tablet of 50 mg in the morning before breakfast, every day for 10 days.

Maintenance treatment: for long-term therapeutic effect 1 tablet of 50 mg UROSTIM is recommended in the morning before breakfast in three consecutive months.

Children: 25 mg daily for children 3 to 14 years of age. One tablet of 25 mg daily according to the above dosage scheme.

In case in which antibiotic therapy is needed UROSTIM should be given in addition to the antibiotic treatment.

DOSAGE FORM AND PACKAGES

tablets of 25 mg for children and 50 mg for adults, 30 tablets in one package.

EXPIRY DATE

2 years

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fax: +359 2/ 943 30 75

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UROSTIM

DENTAVAX

COMPOSITION

Each tablet DENTAVAX contains 36.0 mg freeze-dried lysate and killed bacterial bodies of the following microbial species: *Lactobacillus acidophilus*, *Streptococcus pyogenes*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Candida albicans*.

INDICATIONS

DENTAVAX has a prophylactic and therapeutic action in case of inflammatory and infectious diseases of the oral cavity and parodontite:

- gingivitis erythematosa and ulcerosa;
- superficial or profound parodontitis;
- stomatitis, glossitis, aphthae, etc.;
- before and after tonsillectomies and after dental extractions;
- Ulcerations and secondary infections caused by artificial dentures.

ADMINISTRATION AND DOSAGE

Tablets for oral administration, which are left to dissolve in the mouth by sucking. In case of acute inflammation - 4-6 tablets daily during 15-30 days. In case of chronic inflammation - 3-4 tablets daily during 15-30 days.

Depending on the clinical data the treatment can be carried out 2-3 times per year.

ATTENTION

Eating and drinking are not allowed for one hour after the preparation has been taken in order not to disturb its local effect.

EXPIRY DATE

2 years

DOSAGE FORM AND PACKAGES

40 tablets of 36 mg in blisters in a box or 40 tablets of 36 mg in a plastic sealed bottle.

PRODUCER

BB-NCIPD, Ltd.

Bulgaria, 1504 Sofia; 26, Yanko Sakazov Blvd.

General Manager - phone: +359 2/ 944 61 91

Marketing - phone: +359 2/ 944 81 19

fax: +359 2/ 943 30 75

PRODUCTS INSPIRED FROM LIFE!

Dentavax

bulbio@bulbio.com

RESPIVAX!

Choose Health! Choose -

RESPIVAX

COMPOSITION

Each tablet RESPIVAX contains 25 mg for children and 50 mg for adults of freeze-dried lysate and killed bacterial bodies of the following microbial species: *Streptococcus pneumoniae*, *Haemophilus influenzae*, *Staphylococcus aureus*, *Klebsiella pneumoniae*.

INDICATIONS

RESPIVAX is intended for oral immunotherapy and immunoprevention of non-specific respiratory diseases and has a very good therapeutic effect in children and adults with recurrent and chronic infections of the respiratory pathways, throat and nose included.

RESPIVAX is practically suitable for patients with hypersensitivity to antibiotics and for infections caused by antibiotics-resistant bacteria. It can be used in combination with any other treatment and allows repeated dose administration without risk of dependence.

CONTRAINDICATIONS

The drug has an excellent tolerance and no contraindications.

ADMINISTRATION AND DOSAGE

For the purposes of immunotherapy RESPIVAX is administered as follows:

Adults: 50 mg daily. Basic course of treatment: one tablet of 50 mg in the morning before breakfast, every day for 10 days.

Maintenance treatment: for long-term therapeutic effect 1 tablet of 50 mg RESPIVAX is recommended in the morning before breakfast for 10 days in three consecutive months.

Children: 25 mg daily for children 3 to 14 years of age. One tablet of 25 mg daily according to the above dosage scheme.

In case in which antibiotic therapy is needed RESPIVAX should be given in addition to the antibiotic treatment.

DOSAGE FORM AND PACKAGES

tablets of 25 mg for children and 50 mg for adults, 30 tablets in one package.

EXPIRY DATE

2 years

PRODUCER

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Marketing - phone: +359 2/ 944 81 19

fax: +359 2/ 943 30 75

PRODUCTS INSPIRED FROM LIFE!

RESPIVAX